



RECEIVED

JUL 03 1996

OSTI

ORNL/ER-366

**ENVIRONMENTAL
RESTORATION
PROGRAM**

**Waste Area Grouping 2
Phase I Task Data Report:
Ecological Risk Assessment and
White Oak Creek Watershed Screening
Ecological Risk Assessment**

MASTER

This document has been approved by the
K-25 Site Technical Information Office
for release to the public. Date: 5/13/96

ENERGY SYSTEMS



MANAGED BY
LOCKHEED MARTIN ENERGY SYSTEMS, INC.
FOR THE UNITED STATES
DEPARTMENT OF ENERGY

DISTRIBUTION OF THIS DOCUMENT IS UNLIMITED *ot*

This report has been reproduced directly from the best available copy.

Available to DOE and DOE contractors from the Office of Scientific and Technical Information, P.O. Box 62, Oak Ridge, TN 37831; prices available from 423-576-8401 (fax 423-576-2865).

Available to the public from the National Technical Information Service, U.S. Department of Commerce, 5285 Port Royal Rd., Springfield, VA 22161.

Energy Systems Environmental Restoration Program

**Waste Area Grouping 2
Phase I Task Data Report:
Ecological Risk Assessment and
White Oak Creek Watershed Screening
Ecological Risk Assessment**

R. A. Efroymsen
B. L. Jackson
D. S. Jones
B. E. Sample
G. W. Suter II
C. J. E. Welsh

Date Issued—May 1996

Prepared for the
U.S. Department of Energy
Office of Environmental Management
under budget and reporting code EW 20

Environmental Management Activities at the
OAK RIDGE NATIONAL LABORATORY
Oak Ridge, Tennessee 37831-8169
managed by
LOCKHEED MARTIN ENERGY SYSTEMS, INC.
for the
U.S. DEPARTMENT OF ENERGY
under contract DE-AC05-84OR21400

Author Affiliations

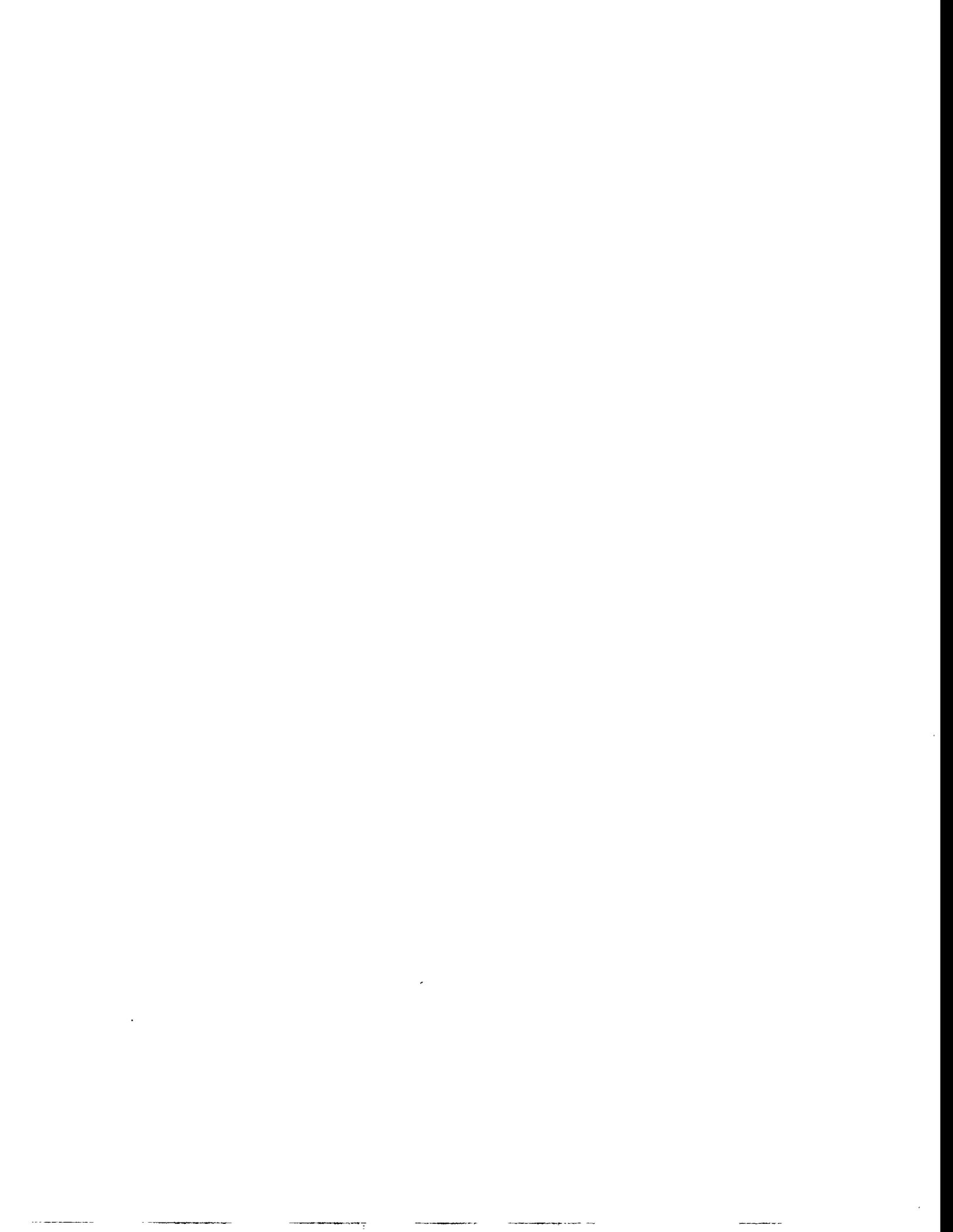
G. W. Suter, B. E. Sample, R. A. Efroymsen, and D. S. Jones are members of the Environmental Sciences Division, Oak Ridge National Laboratory. C. J. E. Welsh is a member of the Health Sciences Research Division, Oak Ridge National Laboratory. B. L. Jackson is a member of the Computer Sciences and Mathematics Division.

PREFACE

This report, *Waste Area Grouping 2 Phase I Task Data Report: Ecological Risk Assessment and White Oak Creek Watershed Screening Ecological Risk Assessment (ORNL/ER-366)*, was prepared in accordance with requirements under the Comprehensive Environmental Response, Compensation, and Liability Act. This work was performed under Work Breakdown Structure 1.4.12.6.1.02.40.08.05 (Activity Data Sheet 3326). Publication of this document meets a project deliverable of May 15, 1996. This document is one of five reports issued in 1996 that provide follow-up information to the *Phase I Remedial Investigation Report for Waste Area Grouping (WAG) 2 at the Oak Ridge National Laboratory*. This report presents results of a screening ecological risk assessment for WAG 2 and, as far as available data permit, for other areas in White Oak Creek watershed. A screening assessment is intended to screen out chemicals that are not hazardous and receptors that are not at risk. A relatively small number of chemicals in water, sediment, and floodplain soils were identified as chemicals of potential ecological concern. However, no receptors could be screened out, and some tributaries and all upland areas except WAG 5 could not be assessed because of lack of data.

DISCLAIMER

This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor any of their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof.



CONTENTS

TABLES	ix
FIGURES	xi
ABBREVIATIONS	xiii
EXECUTIVE SUMMARY	xv
1. INTRODUCTION	1-1
1.1 PURPOSE	1-1
1.2 BACKGROUND	1-1
1.3 ORGANIZATION OF THE ECOLOGICAL RISK ASSESSMENT	1-3
2. ECOLOGICAL PROBLEM FORMULATION	2-1
2.1 ENVIRONMENTAL DESCRIPTION	2-1
2.2 SOURCES	2-1
2.3 ECOLOGICAL ENDPOINTS	2-5
2.3.1 Assessment Endpoints	2-5
2.3.2 Measurement Endpoints	2-7
2.4 CONCEPTUAL MODELS	2-9
3. RISKS TO FISH	3-1
3.1 EXPOSURE ASSESSMENT FOR FISH	3-1
3.1.1 Aqueous Chemical Exposure	3-1
3.1.2 Radiation Exposures	3-1
3.2 EFFECTS ASSESSMENT FOR FISH	3-2
3.2.1 Single Chemical Aqueous Toxicity	3-2
3.2.2 Radiological Effects	3-2
3.2.3 Ambient Water Toxicity	3-2
3.2.4 Fish Community Survey	3-6
3.2.5 Indicators of Fish Reproduction	3-6
3.3 RISK CHARACTERIZATION FOR FISH	3-6
3.3.1 Single Chemical Aqueous Toxicity	3-6
3.3.2 Single Chemical Internal Toxicity	3-10
3.3.3 Radiation Risks	3-10
3.3.4 Ambient Water Toxicity	3-10
3.3.5 Biological Indicators	3-11
3.3.6 Biological Surveys	3-11
3.3.7 Weight Of Evidence For Fish	3-11
3.3.8 Summary of Risk Characterization for Fish	3-13
3.4 UNCERTAINTIES CONCERNING RISKS TO FISH	3-14
4. RISKS TO BENTHIC INVERTEBRATES	4-1
4.1 EXPOSURE ASSESSMENT FOR BENTHIC INVERTEBRATES	4-1
4.1.1 Chemicals In Sediment	4-1

4.1.2 Sediment Radiation Exposures	4-1
4.2 EFFECTS ASSESSMENT FOR BENTHIC INVERTEBRATES	4-2
4.2.1 Single Chemical Sediment Toxicity	4-2
4.2.2 Invertebrate Community Surveys	4-5
4.3 RISK CHARACTERIZATION FOR BENTHIC INVERTEBRATES	4-5
4.3.1 Single Chemical Sediment Toxicity	4-5
4.3.2 Sediment Radiation Characterization	4-8
4.3.3 Benthic Community Surveys	4-8
4.3.4 Weight of Evidence for Benthic Invertebrates	4-8
4.4 UNCERTAINTIES CONCERNING RISKS TO BENTHIC INVERTEBRATES	4-9
5. RISKS TO SOIL INVERTEBRATES	5-1
5.1 EXPOSURE ASSESSMENT FOR SOIL INVERTEBRATES	5-1
5.2 EFFECTS ASSESSMENT FOR SOIL INVERTEBRATES	5-1
5.3 RISK CHARACTERIZATION FOR SOIL INVERTEBRATES	5-2
5.3.1 Single Chemical Soil Toxicity	5-2
5.3.3 Summary of Risk Characterization for Soil Invertebrates	5-3
5.4 UNCERTAINTIES CONCERNING RISKS TO SOIL INVERTEBRATES	5-3
6. RISKS TO PLANTS	6-1
6.1 EXPOSURE ASSESSMENT FOR PLANTS	6-1
6.1.1 Soil Exposures	6-1
6.1.2 Soil Solution Exposures	6-1
6.2 EFFECTS ASSESSMENT FOR THE PLANT COMMUNITY	6-2
6.3 RISK CHARACTERIZATION FOR THE PLANT COMMUNITY	6-2
6.3.1 Single Chemical Soil and Solution Toxicity	6-2
6.3.2 Summary of Risk Characterization for the Plant Community	6-4
6.4 UNCERTAINTIES CONCERNING RISKS TO PLANTS	6-4
7. RISKS TO WILDLIFE	7-1
7.1 RISKS TO WILDLIFE FROM CHEMICALS	7-2
7.1.1 Exposure Assessment	7-2
7.1.2 Chemical Effects Assessment for Wildlife	7-6
7.1.3 Characterization Chemical Risks to Wildlife	7-9
7.1.4 Uncertainties Concerning Risks to Wildlife	7-31
7.2 RISKS TO PLANTS AND EARTHWORMS FROM EXPOSURE TO RADIONUCLIDES	7-32
7.2.1 Exposure Assessment	7-32
7.2.2 Effects Levels for Radionuclides	7-38
7.2.3 Risk Characterization for Terrestrial Receptors Exposed to Radionuclides	7-38
7.7.4 Uncertainties in the Radiological Risk Assessment	7-40
8. SUMMARY AND CONCLUSIONS	8-1
9. REFERENCES	9-1

APPENDIX A.
Benthic Macroinvertebrate Community Study
of Waste Area Grouping 2 Deposition Areas A-1

APPENDIX B.
Data Tables for Risks to Fish, Invertebrates, and Plants B-1

APPENDIX C.
Data Tables for Risks to Wildlife C-1

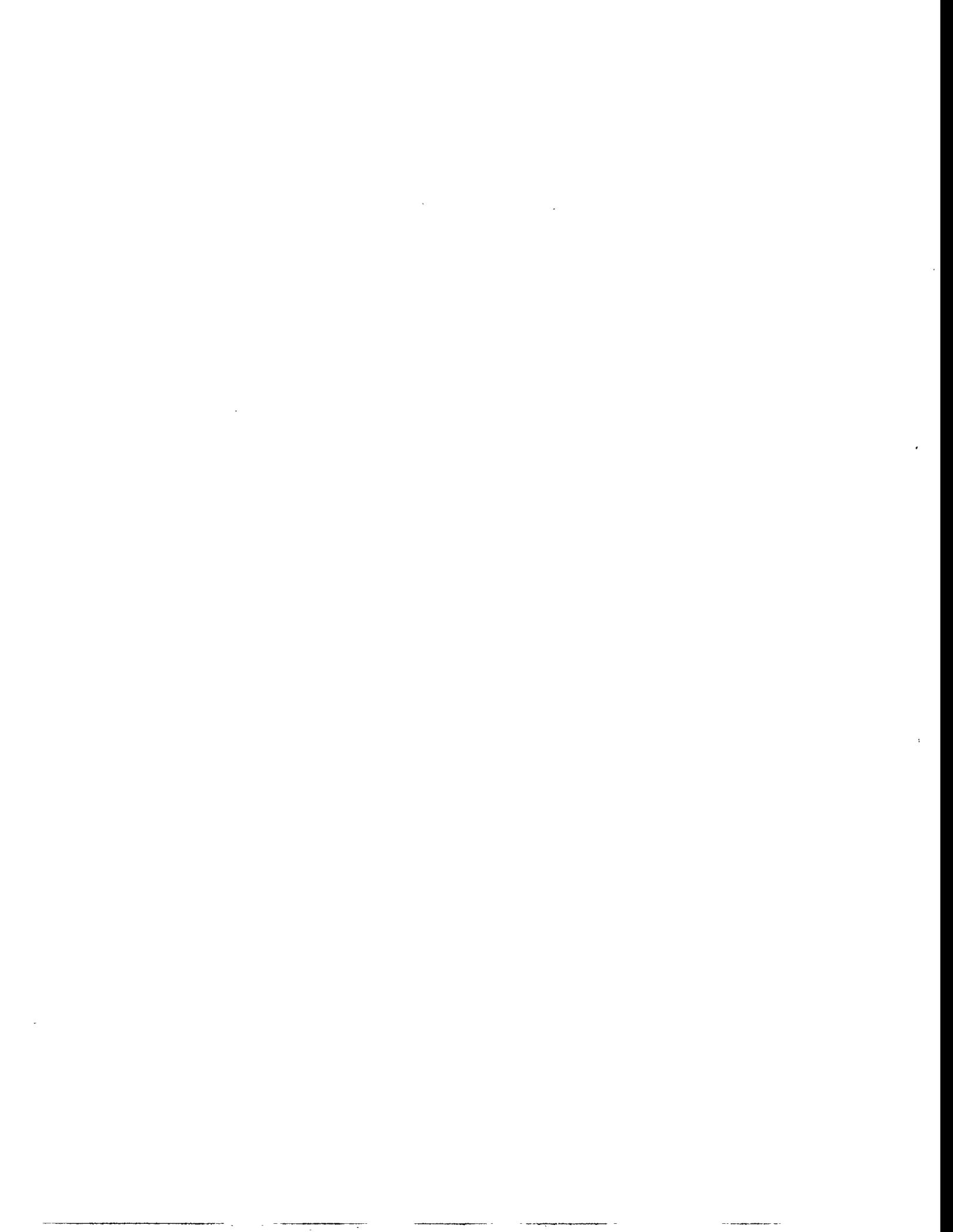
TABLES

2.1	Definition of reaches in the White Oak Creek watershed	2-3
3.1	Descriptions of the ecotoxicological screening benchmarks for aquatic biota	3-3
3.2	Exposure factors for the radionuclides detected in WAG 2 sediment and surface water and their short-lived progeny	3-5
3.3	Results of screening of chemicals that exceed benchmarks in whole or filtered water for chemicals of potential ecological concern (COPECs)	3-8
3.4	Availability of biological data related to the fish community endpoint in the White Oak Creek watershed reaches	3-13
4.1	Descriptions of the ecotoxicological screening benchmarks for benthic biota exposed to contaminated sediments	4-3
4.2	Results of screening of chemicals that exceed benchmarks in sediments for COPECs	4-6
7.1	Metal concentration in hair of mink from the Oak Ridge Reservation and from off-site reference samples	7-7
7.2	Comparison of exposure distributions	7-19



FIGURES

1.1	Map of White Oak Creek watershed and its tributaries	1-2
2.1	Map of White Oak Creek watershed with anthropogenic features including the waste area groups	2-2
2.2	Map of the White Oak Creek watershed showing the stream reaches	2-10
2.3	Conceptual model of the mechanisms of transport and exposure by which ecological receptors are exposed to contaminants in White Oak Creek, its tributaries, and its floodplain	2-11
2.4	Conceptual model of the mechanisms by which wide-ranging wildlife species are exposed to contaminants in White Oak Creek, its tributaries, and its floodplain	7-10
7.1	Summary of lowest observed adverse effects level-based toxic units for short-tailed shrews in WAG 2	7-11
7.2	Summary of lowest observed adverse effects level-based toxic units for white-footed mice in WAG 2	7-12
7.3	Summary of lowest observed adverse effects level-based toxic units for red fox in WAG 2	7-13
7.4	Summary of lowest observed adverse effects level-based toxic units for mink in WAG 2	7-14
7.5	Summary of lowest observed adverse effects level-based toxic units for mink in WAG 2 (exposure through ingestion of fish only)	7-15
7.6	Summary of lowest observed adverse effects level-based toxic units for white-tailed deer in WAG 2	7-16
7.7	Summary of lowest observed adverse effects level-based toxic units for wild turkey in WAG 2	7-17
7.8	Summary of lowest observed adverse effects level-based toxic units for red-tailed hawks in WAG 2	7-17
7.9	Summary of lowest observed adverse effects level-based toxic units for belted kingfisher in WAG 2 (exposure through ingestion of fish only)	7-18
7.10	Cumulative binomial probability of short-tailed shrews experiencing exposure to Aroclor-1260 in WAG 2 in excess of the lowest observed adverse effects level	7-21
7.11	Cumulative binomial probability of short-tailed shrews experiencing exposure to cadmium in WAG 2 in excess of the lowest observed adverse effects level	7-22
7.12	Cumulative binomial probability of short-tailed shrews experiencing exposure to selenium in WAG 2 in excess of the lowest observed adverse effects level	7-23
7.13	Cumulative binomial probability of red fox experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level	7-24
7.14	Cumulative binomial probability of belted kingfisher experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level	7-25
7.15	Cumulative binomial probability of wild turkey experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level	7-26



ABBREVIATIONS

BCV	Bear Creek Valley
BMAP	Biological Monitoring and Abatement Program
BWC	Background White Oak Creek
COPECs	chemicals of potential ecological concern
CV	chronic values
DOE	U.S. Department of Energy
DQO	Data Quality Objectives
EPA	U.S. Environmental Protection Agency
ER-L	effects range-low
EWC	White Oak Creek Embayment
FFA	Federal Facility Agreement
FY	fiscal year
HRT	Homogeneous Reactor Test
HQ	hazard quotient
IAEA	International Atomic Energy Agency
ICRP	International Council for Radiation Protection
IHP	Intermediate Holding Pond
LMB	Lower Melton Branch
LOAEL	lowest observed adverse effects level
LOEC	lowest observed effect concentration
LWC	Lower White Oak Creek
MWC	Middle White Oak Creek
NAWQ	National Ambient Water Quality Criteria
NCRP	National Council on Radiation Protection
NOAEL	no observed adverse effects level
NOEC	no observed effect concentration
NPDES	National Pollutant Discharge Elimination System
NWT	Northwest Tributary
ORNL	Oak Ridge National Laboratory
ORR	Oak Ridge Reservation
OU	operable unit
PAH	polycyclic aromatic hydrocarbons
PCB	polychlorinated biphenyl
PRL	probably effect level
RAC	Raccoon Creek
RI	remedial investigation
SAV	secondary acute value
SCV	secondary chronic values
TDEC	Tennessee Department of Environment and Conservation
TEL	threshold effect level
T&E	threatened and endangered
UCL	upper confidence limit
TIE	toxicity identification and evaluation
UCB	upper confidence bound
UMB	Upper Melton Branch
WAG	Waste Area Grouping

WOC	White Oak Creek
WOL	White Oak Lake
WS	West Seep
W4T	WAG 4 Tributary
1WC	WAG 1 White Oak Creek

EXECUTIVE SUMMARY

This report presents an ecological risk assessment for Waste Area Grouping (WAG) 2 based on the data collected in the Phase I remedial investigation (RI). It serves as an update to the WAG 2 screening ecological risk assessment that was performed using historic data. In addition to identifying potential ecological risks in WAG 2 that may require additional data collection, this report serves to determine whether there are ecological risks of sufficient magnitude to require a removal action or some other expedited remedial process.

WAG 2 consists of White Oak Creek (WOC) and its tributaries downstream of the Oak Ridge National Laboratory (ORNL) main plant area, White Oak Lake (WOL), the White Oak Creek Embayment of the Clinch River, associated flood plains, and the associated groundwater. The WOC system drains the WOC watershed, an area of approximately 16.8 km² that includes ORNL and associated WAGs. The WOC system has been exposed to contaminants released from ORNL and associated operations since 1943 and continues to receive contaminants from adjacent WAGs.

Since the WAG 2 program was planned, Federal Facility Agreement managers have decided to proceed with a remedial investigation for the WOC watershed. An early requirement of that activity is a screening ecological risk assessment for the watershed to identify hazards and data gaps. Since that assessment was needed at approximately the same time as the WAG 2 screening assessment and since the WAG 2 program is the primary source of data concerning the chemical composition of ambient media other than groundwater for the watershed, the assessments were combined. Therefore, this document used both WAG 2 program data and other data that was available to present a screening assessment of the WOC watershed. WAGs 5 and 6 and the WAG 1 surface impoundments have completed remedial investigations, and their results are not repeated here. Other WAGs have essentially no data concerning surface contamination, and they are scheduled for limited sampling and analysis.

This screening assessment is intended to indicate whether there are credible hazards to ecological endpoints in the WOC watershed due to contamination and whether there are data gaps that need to be filled. The following are conclusions concerning the hazards:

- Radionuclides in WAG 2 do not pose a hazard to fish or aquatic invertebrates. ⁹⁰Sr in three seeps exceed the screening benchmarks for aquatic life, but these seeps do not support communities of aquatic macroorganisms.
- Radionuclides in WAG 2 pose a hazard to terrestrial plants, wildlife, and soil invertebrates in all four reaches of WOC and in lower Melton Branch. Exposures were highest in the Intermediate Holding Pond (IHP) reach.
- The unfiltered water in all reaches exceeded water quality criteria and other toxicological benchmarks for aquatic life.
- Water from WOC adjacent to and below ORNL was toxic in a fish embryo-larval test but not in the standard subchronic tests.
- The fish community in WOC has low species richness, but this may be due to lack of recovery from past toxicity. However, the riffle invertebrate community that is exposed primarily to chemicals in water like the fish and has flying stages that should allow recovery also has low species richness.

- Sediments from the IHP, Middle WOC (MWC), and Lower Melton Branch (LMB) are contaminated to levels that have been associated with toxic effects at other sites. Contaminants include metals, polycyclic aromatic hydrocarbons, and polychlorinated biphenyls.
- The species richness of benthic invertebrates from WAG 2 sediments is lower than in background WOC sediments but not other reference streams. The exception is WOL, which has very low species richness. However, no comparable reference lakes were characterized.
- Chromium and mercury levels in IHP and Lower WOC (LWC) soils and mercury in MWC and LMB soils exceeded levels that were reported to be toxic to earthworms.
- Multiple metals were found in IHP, MWC, LWC, and LMB soils at concentrations that have been reported to be toxic to plants.
- Metals in seep waters in the WAG 4 Tributary and West Seep reaches are at concentrations that have been reported to be toxic to plants.
- Hazards to wildlife were found in all WOC reaches. However, the hazards were greatest for mercury among chemicals, for IHP among reaches, and for shrews among species.
- Kingfishers from WOC show elevated levels of contaminants, and the one adult found had tissue levels of mercury and selenium that are indicative of toxicity. Mink from WOC do not appear to be contaminated.

The following are major uncertainties that could be addressed by additional data collection:

- Background concentrations need to be better characterized for the seeps, surface waters, sediments, and floodplain soils.
- The contamination of soil and biota on burial grounds other than WAGs 5 and 6 are unknown.
- The bioavailable concentrations of metals in WOC are unknown.
- The toxicity of soil and sediment in the watershed are unknown.
- The toxicity of water is unknown for some reaches and has been irregular in others.
- The fish communities have not been quantitatively characterized for the smaller tributaries.
- The chemical composition of fish is unknown for some tributaries and reaches.
- Because of the high selenium levels in a kingfisher, selenium levels should be characterized in fish.

In sum, plausible hazards exist in all reaches of WAG 2 and to all ecological endpoints. Radionuclides, organic chemicals, and metals are all implicated. However, none of the risk estimates or observations of the state of the watershed suggest that there is a need for an accelerated response. That is, no threatened or endangered species are at risk and no wetlands or other highly valued populations or ecosystems are experiencing catastrophic effects. Some parts of the watershed are uncharacterized and should be surveyed and sampled before the RI is completed. These results imply that a more complete data set should be assembled and used to prepare a definitive ecological risk assessment for the WOC watershed.

1. INTRODUCTION

1.1 PURPOSE

This is one of five reports issued in 1996 that provide follow-up information to the Phase I Remedial Investigation (RI) Report for Waste Area Grouping (WAG) 2 at Oak Ridge National Laboratory (ORNL). The five reports address areas of concern that could cause immediate risk to public health at the Clinch River and ecological risk within WAG 2 at ORNL. The five reports that complete activities conducted as part of Phase I of the RI for WAG 2 are as follows:

- Waste Area Grouping 2, Phase I Task Data Report: Seep Data Assessment
- Waste Area Grouping 2, Phase I Task Data Report: Tributaries Data Assessment
- Waste Area Grouping 2, Phase I Task Data Report: Ecological Risk Assessment
- Waste Area Grouping 2, Phase I Task Data Report: Human Health Risk Assessment
- Waste Area Grouping 2, Phase I Task Data Report: Sediment Transport Modeling

In December 1990, the "Remedial Investigation Plan for Waste Area Grouping 2 at Oak Ridge National Laboratory" was issued (ORNL). The WAG 2 RI Plan was structured with a short-term component to be conducted while up-gradient WAGs are investigated and remediated, and a long-term component that will complete the RI process for WAG 2 following remediation of up-gradient WAGs. RI activities for the short-term component were initiated with the approval of the U. S. Environmental Protection Agency (EPA), Region IV, and the Tennessee Department of Environment and Conservation (TDEC).

This report presents a screening ecological risk assessment for WAG 2 based on the data collected in the Phase I Remedial Investigation. It serves as an update to the WAG 2 screening ecological risk assessment that was performed using historic data (Suter 1992). In addition to identifying potential ecological risks in WAG 2 that may require additional data collection, it serves to determine whether ecological risks are of sufficient magnitude to require removal or some other expedited remedial process.

1.2 BACKGROUND

WAG 2 consists of White Oak Creek (WOC) and its tributaries downstream of the ORNL main plant area, White Oak Lake, the WOC embayment of the Clinch River and the associated flood plains, and the associated groundwater (Fig. 1.1). The WOC system drains the WOC watershed, an area of approximately 16.8 km² that includes ORNL and associated WAGs. The WOC system has been exposed to contaminants released from ORNL and associated operations since 1943 and continues to receive contaminants from adjacent WAGs.

The WAG 2 RI Plan developed in 1990 was not a prototypical RI plan. It was recognized that full implementation of an RI was inappropriate while contaminants continue to enter the system. A phased effort was adopted in response to the need to take initial steps to protect the public and the environment and to characterize and assess risks associated with WAG 2 and the limitations imposed by changing contaminant input.

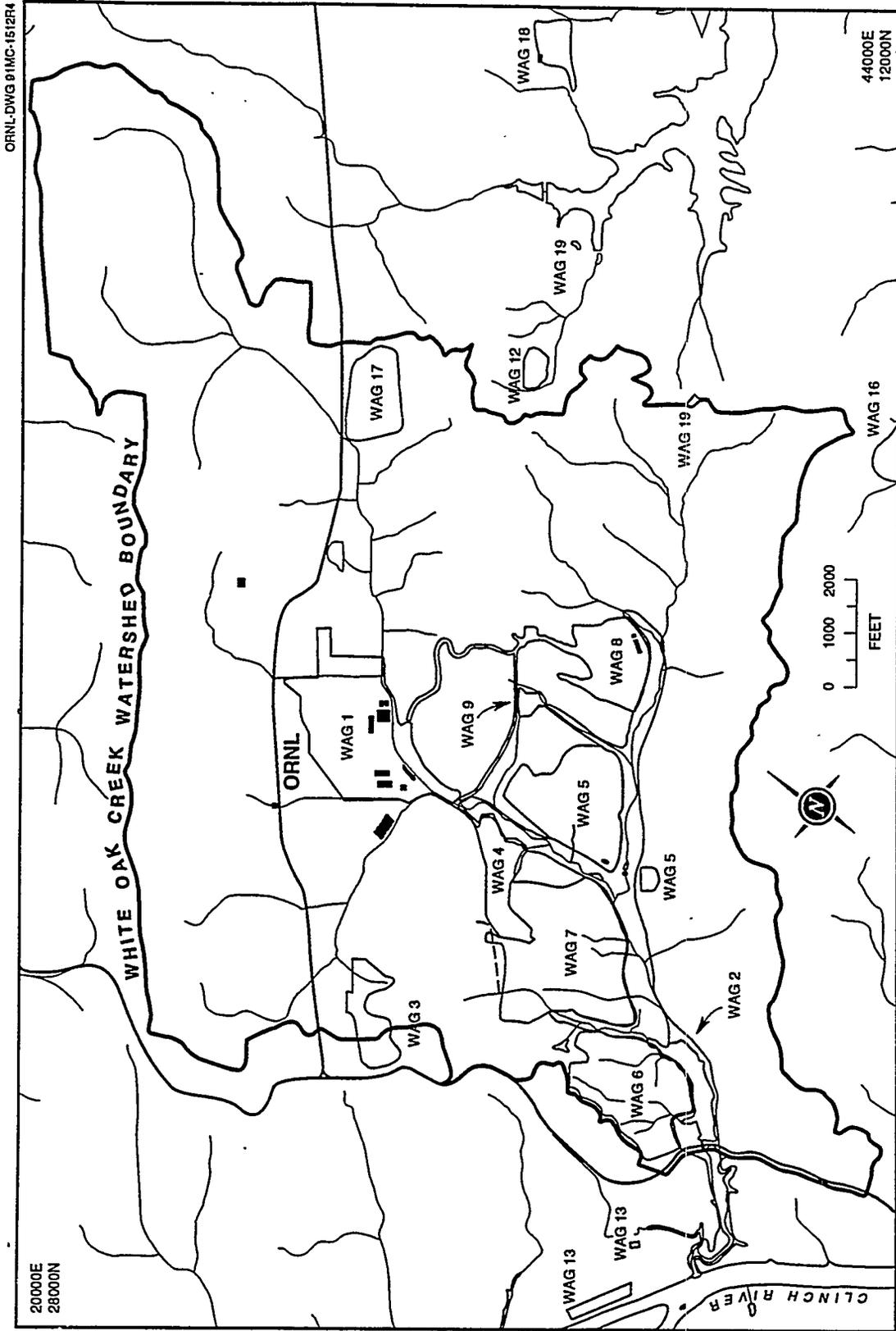


Fig. 1.1. Map of White Oak Creek and its tributaries.

Three phases were initially identified: Phase I was the initial scoping activity to determine the need for early action; Phase II included interim activities during remediation of up-gradient WAGs to evaluate potential changes in the contamination status of WAG 2 that would necessitate reevaluation of the need for early action; and Phase III would be completion of the Comprehensive Environmental Response, Compensation, and Liability Act process following remediation of the up-gradient WAGs. Field activities were initiated in fiscal year (FY) 1992 consistent with the RI Plan (ORNL 1990), and a report summarizing Phase I results to date was published in 1993 (DOE).

On June 20 and 21, 1994, a Data Quality Objectives (DQO) Workshop was held with representatives of the Department of Energy (DOE), EPA, and TDEC. The participants defined the nature and boundaries of the problems for the WAG 2 RI, determined decision criteria, and established inputs to be used for characterizing the site for decision-making purposes. During the workshop, the regulators made recommendations that would alter the initial WAG 2 RI plan. Consequently, the Federal Facility Agreement (FFA) managers from EPA, TDEC, and DOE directed that FY 1995 WAG 2 RI activities concentrate on meeting FFA requirements.

The FFA managers also directed that the WAG 2 RI be changed to a two-phase field program by eliminating Phase II activities and transferring needed elements into the newly formed ORNL Environmental Restoration Surface Water Program. A separate FY 1995 WAG 2 RI Work Plan was developed (DOE 1994) to replace previously identified planning and tasking documents. Emphasis was to be on analysis of existing data, data interpretation, and reporting of results. This document reports the results of the ecological risk assessment.

Finally, after the WAG 2 program was planned, the FFA managers decided to proceed with an RI for the WOC watershed. An early requirement of that activity is a screening ecological risk assessment for the watershed to identify hazards and data gaps. Since that assessment was needed at approximately the same time as the WAG 2 screening assessment and since the WAG 2 program is the primary source of data concerning the chemical composition of ambient media other than groundwater for the watershed, the assessments were combined. Therefore, this document used both WAG 2 program data and other data that were available to present a screening assessment of the WOC watershed. WAGs 5 and 6 and the WAG 1 surface impoundments have completed RIs, and their results are not repeated here. Other WAGs have essentially no data concerning surface contamination, and they are scheduled for limited sampling and analysis.

1.3 ORGANIZATION OF THE ECOLOGICAL RISK ASSESSMENT

This screening ecological risk assessment is organized in terms of the standard framework (EPA 1992). After a problem formulation the risks of chemicals to each of the ecological risk assessment endpoints are assessed separately (Sect. 3.7). Each includes an exposure assessment, effects assessment, characterization of risk, and characterization of uncertainty. Finally, ecological risks are summarized. The components of the assessment are explained in the following.

The problem formulation defines the scope and content of the assessment. It describes the site and potential contaminant sources, defines assessment and measurement endpoints, and presents the conceptual model.

Exposure assessment characterizes the distribution in space and time of the concentrations of contaminants to which organisms are exposed. Risk from undetected chemicals was not assessed, as instructed by the FFA. Exposure calculations are performed for each reach.

Effects assessment characterizes the evidence concerning effects of contaminant exposure. The principal lines of evidence concerning effects are biological survey data that indicate the actual state of the receiving environment, media toxicity data that indicate whether the contaminated media are toxic under controlled conditions, bioindicator data that are biochemical and histological indications of the potential mechanisms and causes of effects, and single chemical toxicity data that indicate the toxic effects of the concentrations measured in site media.

Risk characterization is the phase of risk assessment in which the information concerning exposure and the information concerning the potential effects of exposure are integrated to estimate risks (the likelihood of effects given the exposure). Because this is a screening assessment, emphasis is placed on screening the chemical concentrations in ambient media against screening benchmark values. Procedurally, the risk characterization is performed for each assessment endpoint by (1) screening all measured contaminants against toxicological benchmarks and background concentrations if available, (2) considering the implications of other types of data for the hypothesis that a hazard exists that requires further assessment, (3) logically integrating the screening results with the other evidence to determine whether a credible hazard exists to the endpoint, and (4) listing and discussing the major uncertainties in the assessment.

2. ECOLOGICAL PROBLEM FORMULATION

The problem formulation consists of describing the relevant features of the environment, describing the sources of contamination, identifying ecological endpoints, and summarizing that information in terms of a conceptual model of the hazard posed by the contaminants to the endpoint biota.

2.1 ENVIRONMENTAL DESCRIPTION

The environment considered in this assessment is the WOC watershed (henceforth, simply watershed) and in particular WAG 2 (Fig. 2.1). WAG 2 consists of Melton Branch and its floodplain below km 1.5 (just above the discharge from the High Flux Isotope Reactor) and WOC and its floodplain downstream of WAG 1 (km 3.45 at the 7500 Bridge).

The streams and floodplains of the watershed are divided into reaches, which are lengths of stream and associated floodplain that are relatively uniform with respect to exposure and ecology. The reaches within WAG 2 correspond to the areas defined for floodplain soil and sediment characterization (Ford et al. 1996). However, because this is also a preliminary assessment of the watershed, additional reaches are defined beyond the five WAG 2 reaches. These are defined in Table 2.1 and portrayed in Fig. 2.2. One stream that is not in the watershed, Raccoon Creek, is included because it was included in the WAG 2 Phase 1 program and it drains WAG 3 which is an ORNL site.

The reaches are divided into two categories. Mainstem reaches are those on WOC, including WOL, and tributaries are streams that feed WOC and are not ephemeral. Many of the water samples are from springs and small tributaries' sites, which include springs, seeps, and ephemeral tributaries. Each of these sites is associated with the reach to which it drains so that sources of chemicals can be identified, but its data are not averaged with the mainstem or tributary data.

No threatened or endangered species are known to occur in WAG 2, but some state-listed species (e.g., river otter) and federally listed species (e.g., bald eagle) are undergoing range expansions and may use WAG 2 in the future. Threatened or endangered species that are believed to occur on the Oak Ridge Reservation (ORR) are listed by Suter et al. (1996).

2.2 SOURCES

The proximate sources considered in this assessment are the contaminated water, sediment, and soil. The ultimate sources of contaminants are the National Pollutant Discharge Elimination System (NPDES) permitted point discharges at ORNL and releases from wastes in WAGs 1, 3-9, and 17. DOE's operations in the WOC watershed have included waste disposal, spills, and use of chemicals such as pesticides in the environment.

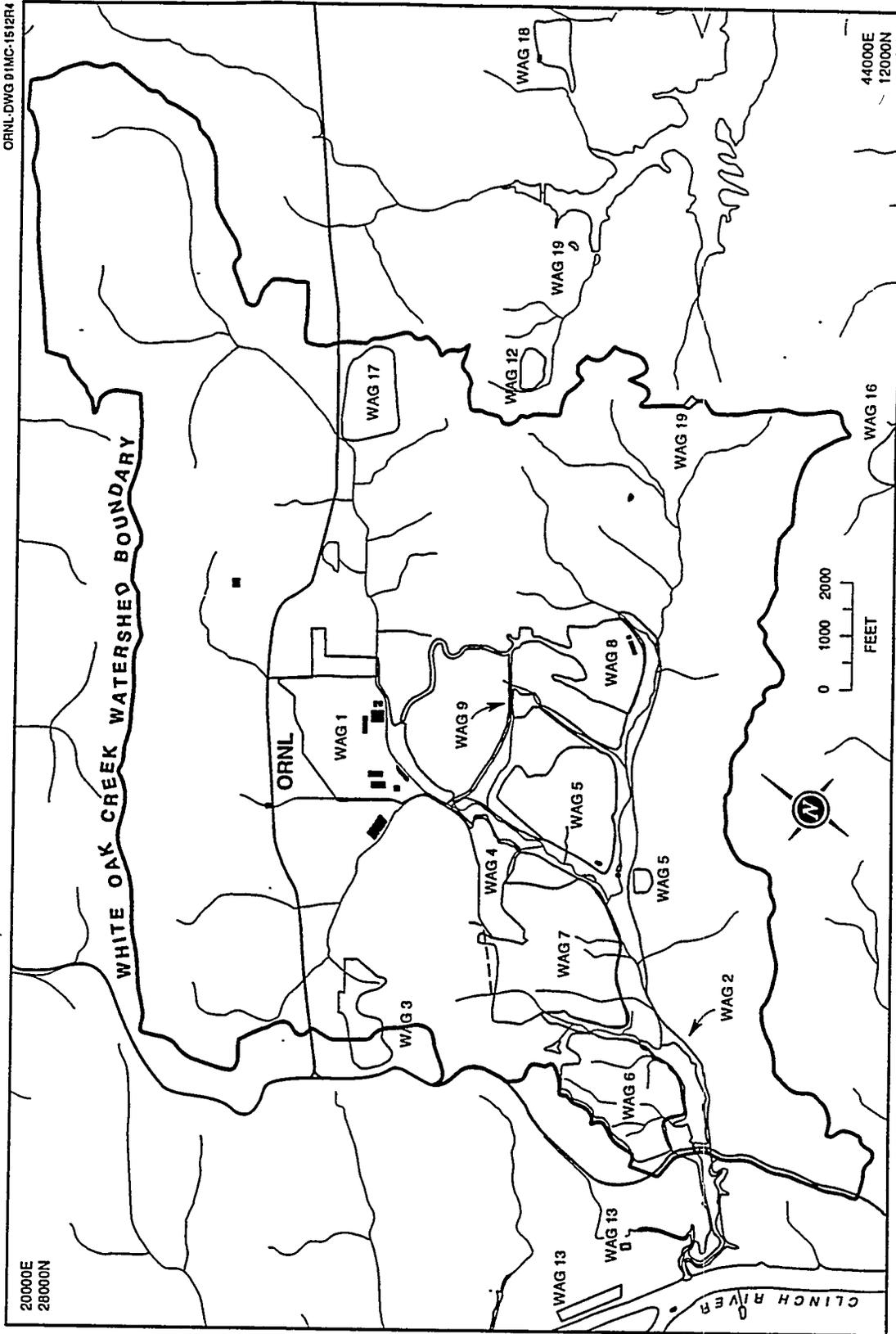


Fig. 2.1. A map of White Oak Creek watershed with anthropogenic features including the waste area groups.

Table 2.1. Definition of reaches in the White Oak Creek watershed

Reach code	Reach description
BWC	Background White Oak Creek (WOC): upstream of WAG 1
IWC	Waste Area Group (WAG) 1 WOC: the stream adjacent to WAG 1 and extending downstream to the 7500 Bridge
IHP	Intermediate Holding Pond: The first WOC reach in WAG 2, extending from the 7500 Bridge to the confluence of the WAG 4 Tributary; it corresponds to the area of the former Intermediate Holding Pond.
MWC	Middle WOC: extends from the WAG 4 Tributary to Melton Branch
LWC	Lower WOC: extends from the Melton Branch to White Oak Lake
WOL	White Oak Lake
EWC	Embayment of WOC: extends from the dam at Route 95 to the coffer dam at the confluence with the Clinch River
UMB	Upper Melton Branch: extends upstream from the Homogeneous Reactor Test (HRT) Tributary
LMB	Lower Melton Branch: extends downstream from the HRT Tributary
1C	First Creek: flows through western ORNL and enters the lower end of the Northwest Tributary
5C	Fifth Creek: flows through central ORNL and enters WOC
NWT	Northwest Tributary: drains the eastern end of WAG 3 and far western ORNL
W4T	WAG 4 Tributary
HRT	Homogeneous Reactor Test Tributary: the tributary of Melton Branch that drains WAG 9
WS	West Seep: a small tributary located between WAGs 6 and 7 that directly enters White Oak Lake
U	Unknown reach: two seeps that are not known to drain into any stream are assigned this designation
RAC	Raccoon Creek: outside the WOC watershed, drains WAG 3

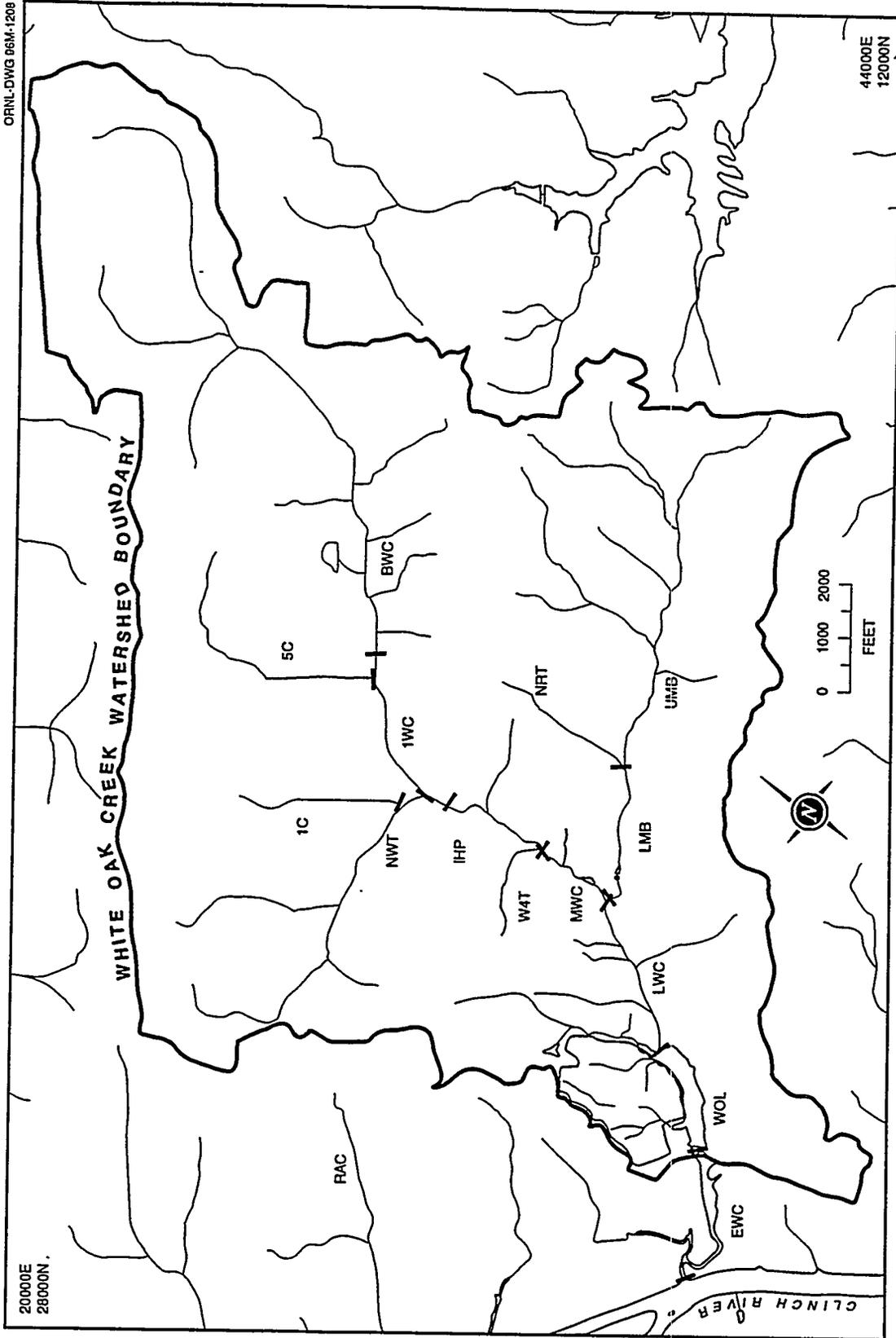


Fig. 2.2. Map of the White Oak Creek watershed showing the stream reaches.

CRNL-DWG 06M-1208

20000E
28000N

44000E
12000N

Chemicals of potential ecological concern (COPECs) for ecological risks have been identified in a previous screening assessment for WAG 2 (Blaylock et al. 1992). However, because the large number of COPECs, the revisions that have occurred in the ecological screening benchmarks, the questionable quality of the existing data used by Blaylock et al., and the new contaminant concentration data that have been obtained, chemicals are rescreened against benchmarks for this assessment.

2.3 ECOLOGICAL ENDPOINTS

The problem formulation must identify both the assessment endpoints, which are explicit statements of the characteristics of the environment that are to be protected, and the measurement endpoints, which are quantitative summaries of a measurement or series of measurements that are related to effects on an assessment endpoint.

2.3.1 Assessment Endpoints

The following assessment endpoints for aquatic and terrestrial risks have been selected for this assessment:

- reduction in species richness or abundance of fishes or increased frequency of gross pathologies in fish communities resulting from toxicity.
- reduction in species richness or abundance of benthic macroinvertebrate communities resulting from toxicity.
- reduction in abundance or production of earthworms resulting from toxicity.
- reduction in abundance or production of piscivorous wildlife populations (kingfisher and mink) resulting from toxicity.
- reduction in production of terrestrial plant communities resulting from toxicity.
- reduction in abundance or production of terrestrial wildlife populations (short-tailed shrew, white-footed mouse, red fox, red-tailed hawk) resulting from toxicity.

The ecological assessment endpoints have been selected based on DQO meetings that included representatives of DOE, EPA Region IV, and TDEC and the strategy for ecological risk assessment on the ORR, which was also a product of a DQO process (Suter et al. 1995).

The endpoints are chosen on the basis of ecological importance, policy or societal significance, susceptibility to toxic effects, and appropriateness of scale. All of these are self explanatory except for scale. An endpoint has appropriate scale for a site if toxic effects on the site could have a significant effect on the endpoint. For example, there is a distinct plant community on the WOC floodplain so properties of that community have an appropriate scale. However, the WOC watershed supports only a few kingfishers, which form a very small fraction of the biological population to which they belong. Therefore, individual kingfishers have an appropriate scale, but the kingfisher population does not.

The following paragraphs explain the selected endpoints.

- The fish community is considered to be an appropriate endpoint community because it is ecologically and societally important, susceptible, and has a scale appropriate to the site. The societal importance is due to recreational fisheries; the ecological significance comes from the fact that much of the energy flow in temperate streams passes through fishes, and fishes are a major nutrient reservoir in those systems. In addition, the fish species in WAG 2 tend to move within an area smaller than the OU and their movement is restricted by weirs and dams, so the scale is appropriate.
- The benthic invertebrate community is considered an appropriate endpoint community because it is highly susceptible and has a scale appropriate to the site. The high susceptibility is due to the association of these organisms with the sediment, which is the repository of most of the COPECs. In addition, the insects and crustaceans that dominate this community are sensitive to a variety of contaminants. Because these organisms are sedentary for all or most of their lives, their scale is highly appropriate to the scale of the site. Benthic invertebrates may also be ecologically important, but the importance of the invertebrates in the depositional areas is unclear.
- Soil invertebrates are considered to be an appropriate endpoint assemblage because they are susceptible and ecologically important and have an appropriate scale. The susceptibility results from their intimate exposure to contaminated soils including, in the case of earthworms, ingestion. The ecological importance is due to their role in litter degradation, nutrient cycling, and maintaining soil structure. The appropriate scale results from their low mobility; biological populations could occupy an area as small as the reaches defined for this assessment.
- The terrestrial plant community is considered an appropriate endpoint community because it is ecologically important, susceptible, and has a scale appropriate to the site. The ecological significance comes from the fact that the plant community is responsible for primary production. This community is susceptible because it would be directly exposed to the contaminants in the floodplain. Finally, the scale is appropriate because plants are immobile and because a distinct plant community occurs on the floodplain.
- Piscivorous and terrestrial wildlife species are considered appropriate because individuals are potentially sensitive due to food-web magnification of chemicals and because of the known sensitivity of some species (e.g., mink). The watershed has an appropriate scale for small mammals (i.e., mice and shrews) in that it could support a biological population of those species. The watershed clearly does not support biological populations of birds or large (e.g., deer) or medium-sized (e.g., mink) mammals, but the regulators have decided that the organisms occurring in a watershed constitute a population for regulatory purposes.

Threatened and endangered species are not directly addressed in this assessment. This is because the other endpoint species are judged to be as sensitive or more sensitive than the endangered species that may come to use the site. In addition, because this is a screening assessment, it is conservative and affords the extra protection appropriate to T&E species.

Wetlands are assumed to be protected by assessing the risks to plants in the small wetland areas associated with seeps. These wetlands should be more highly exposed than those associated with the streams.

2.3.2 Measurement Endpoints

Three basic types of effects data are potentially available to serve as measurement endpoints: results of biological surveys, toxicity tests performed on media from the CR operable unit (OU), and toxicity test endpoints for chemicals found in the OU. Measurement endpoints are presented in the following paragraphs.

2.3.2.1 Fish

Biological Survey Data. No fish survey data were collected by the WAG 2 program. However, results of Biological Monitoring Abatement Program (BMAP) surveys will be cited as supporting evidence. The BMAP measurement endpoints are assumed to be direct estimates of that assessment endpoint.

Biological Indicators Data. No fish bioindicators data were collected by the WAG 2 program. However, published results of the BMAP biological indicators task will be cited as supporting evidence. Frequencies of gross pathologies are a direct measure of one aspect of the assessment endpoint. Measures of fish fecundity in largemouth bass and bluegill provide an indication of the potential contribution of reproductive toxicity to community effects. Measures of the levels of physiological and histological condition in redbreast sunfish help to confirm that exposures have occurred and may suggest mechanistic connections between exposure and effects on the fish community.

Media Toxicity Data. No fish aqueous toxicity tests were conducted by the WAG 2 program. However, published results of the BMAP tests will be cited as supporting evidence. Test endpoints include reductions in growth and survivorship of larval fathead minnows and in fecundity and survivorship of *Ceriodaphnia dubia* (*C. dubia*) in 7-day tests of ambient water and reductions in hatching and larval survival and increases in terata in Japanese medaka (*Oryzias latipes*) eggs and larvae exposed to ambient water from shortly after fertilization to 48 hours post-hatch. Responses that are statistically significantly different or are inhibited by 20% or greater relative to control or reference waters are assumed to be indicative of waters that are toxic to fish.

Single Chemical Toxicity Data. Chronic toxicity thresholds for freshwater fish are expressed as chronic EC20s or chronic values (CVs). These test endpoints correspond to the assessment endpoint for this community. That is, the sensitivity distribution of the test species is assumed to approximate the distribution of OU species, and exceeding the CVs and EC20s is assumed to correspond to 20% or greater reductions in abundance, with some uncertainty.

2.3.2.2 Benthic invertebrates

Biological Survey Data. Benthic invertebrate survey data were collected by the WAG 2 program from areas in which fine sediments had been deposited. In addition, results of BMAP surveys of benthic invertebrates in riffles will be cited as supporting evidence. The measurement endpoints for both surveys are assumed to be direct estimates of that assessment endpoint.

Media Toxicity Data. Sediment toxicity tests planned for WAG 2 could not be performed because of concerns for worker safety.

Single Chemical Toxicity Data. Chronic toxicity thresholds for freshwater invertebrates expressed as chronic EC20s or CVs. These test endpoints correspond to the assessment endpoint

for this community. That is, the sensitivity distribution of the test species is assumed to approximate the distribution of OU species, and exceeding the CVs and EC20s is assumed to correspond to 20% reductions in population abundance or greater, with some uncertainty. Toxic concentrations in ambient sediments reported by the state of Florida. Two types of values were extracted from that data set: (1) thresholds for modification of benthic invertebrate community properties based on co-occurrence analyses, which are assumed to correspond to the assessment endpoint.; (2) thresholds for lethality in toxicity tests of contaminated sediments, which are also assumed to correspond to the assessment endpoint effect but with greater uncertainty due to the extrapolation to the field.

2.3.2.3 Piscivorous wildlife

Biological Survey Data. Kingfisher reproduction was surveyed in WAG 2 and reference areas. Assuming that the kingfishers in the watershed constitute a population, this is a direct measure of the assessment endpoint for avian piscivores.

Media Toxicity Data. None were performed.

Single Chemical Toxicity Data. Chronic toxicity thresholds for contaminants of concern in birds and mammals gave greater weight to data from long-term feeding studies with wildlife species. Preference was given to tests that included reproductive endpoints. After allometric scaling for the endpoint species, these test endpoints are assumed to correspond to effects on individuals that could result in exceeding the population-level assessment endpoint. An extrapolation must be made to populations if effects on individuals are estimated to occur. In addition, body burdens of a kingfisher were compared to concentrations associated with toxic effects on birds.

2.3.2.4 Terrestrial wildlife

Biological Survey Data. None were performed.

Media Toxicity Data. None were performed.

Single Chemical Toxicity Data. Chronic toxicity thresholds for contaminants of concern in birds and mammals gave greater weight to data from long-term feeding studies with wildlife species. Preference was given to tests that included reproductive endpoints. After allometric scaling for the endpoint species, these test endpoints are assumed to correspond to effects on individuals that could result in exceeding the population-level assessment endpoint. An extrapolation must be made to populations if effects on individuals are estimated to occur.

2.3.2.5 Terrestrial plants

Biological Survey Data. None.

Media Toxicity Data. None.

Single Chemical Toxicity Data. This includes EC20s for growth or production of vascular plants or equivalent chronic toxicity thresholds for contaminants of concern in soil. These test endpoints are assumed to correspond to the assessment endpoint for this community. That is, the sensitivity distribution of the test species is assumed to approximate the distribution of species

that would colonize Clinch River dredge spoil; exceeding the test endpoints is assumed to correspond to 20% reductions in abundance or productivity with some uncertainty; and a distinct plant community is assumed to occur on a spoil disposal area.

2.3.2.6 Soil invertebrates

Biological Survey Data. Earthworms were collected by the WAG 2 program in a manner that produces only presence/absence information.

Media Toxicity Data. Earthworm toxicity tests planned for WAG 2 could not be performed because of concerns for worker safety.

Single Chemical Toxicity Data. Chronic toxicity thresholds for earthworms have been obtained from the literature. These test endpoints vary in their relevance, but they are assumed to correspond to the assessment endpoint for this assemblage if they include sublethal responses.

2.3.2.7 Threatened and endangered species

Biological Survey Data. None.

Media Toxicity Data. None.

Single Chemical Toxicity Data. The same chronic toxicity thresholds for contaminants of concern in invertebrates, fish, birds and mammals used as measurement endpoints for other species are used with the T&E species but are interpreted in the risk characterization so as to provide the higher level of protection specified in the assessment endpoint.

2.4 CONCEPTUAL MODELS

Conceptual models are graphical representations of the relationships among sources of contaminants, ambient media, and the endpoint biota. Figure 2.3 shows a conceptual model for the streams and floodplains of the WOC watershed. Figure 2.4 shows a conceptual model for wide-ranging wildlife that utilize the streams and floodplains as well as upland areas and nearby aquatic habitat. The exposure pathways shown are those that are included in the assessment. Note that different benthic invertebrate assemblages are exposed to water and sediments. That is, the invertebrates inhabiting soft sediments in depositional areas are assumed to be exposed to chemicals in those sediments and their pore water with negligible exposure to free water and invertebrates inhabiting the rocky substrates of riffles are assumed to be primarily exposed to chemicals in water. These conceptual models are derived from the generic models developed for the ORR and are discussed in detail in the strategy document for ecological risk assessment on the ORR (Suter et al. 1995).

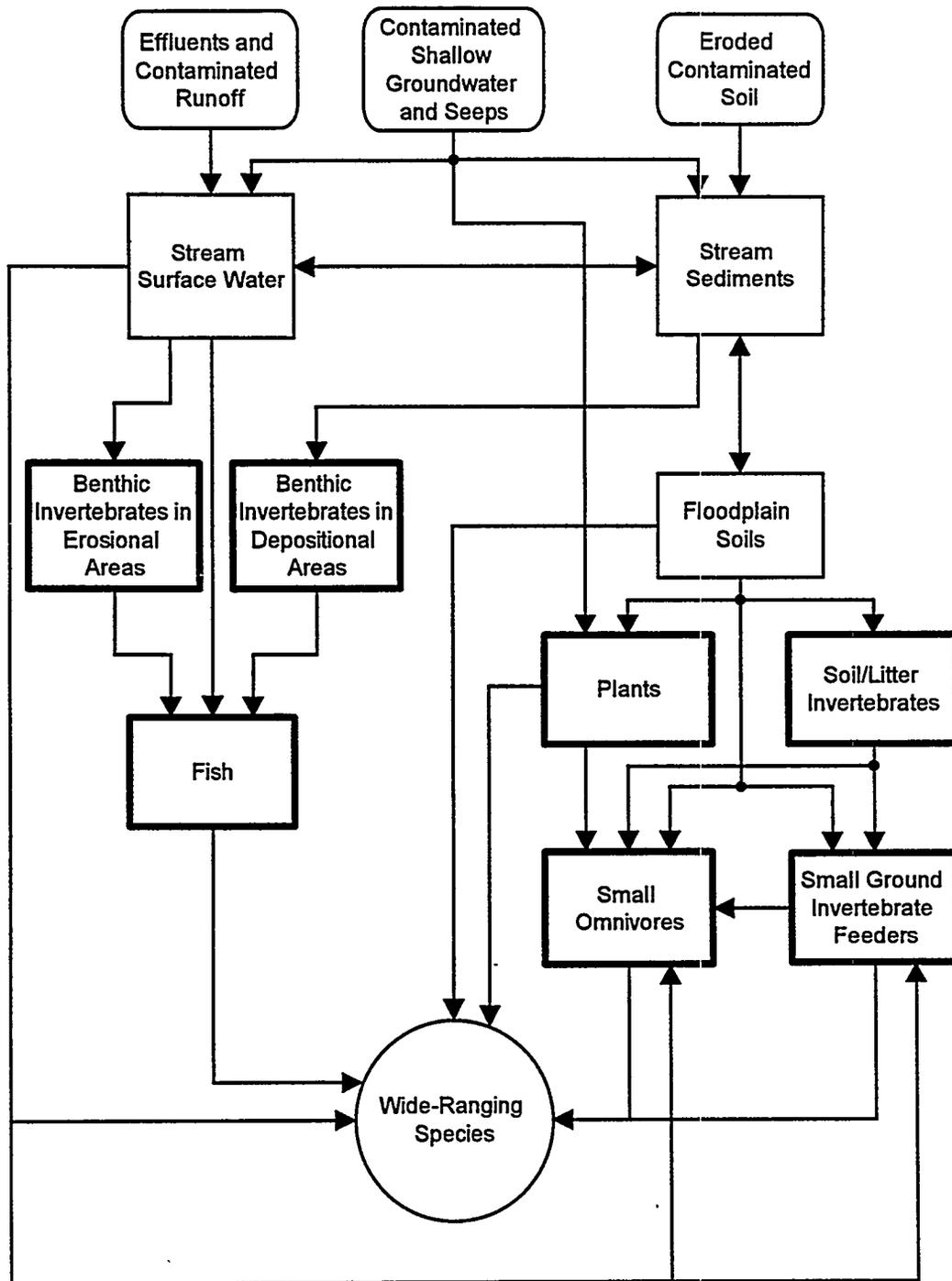


Fig. 2.3. Conceptual model of the mechanisms of transport and exposure by which ecological receptors are exposed to contaminants in White Oak Creek, its tributaries, and its floodplain.

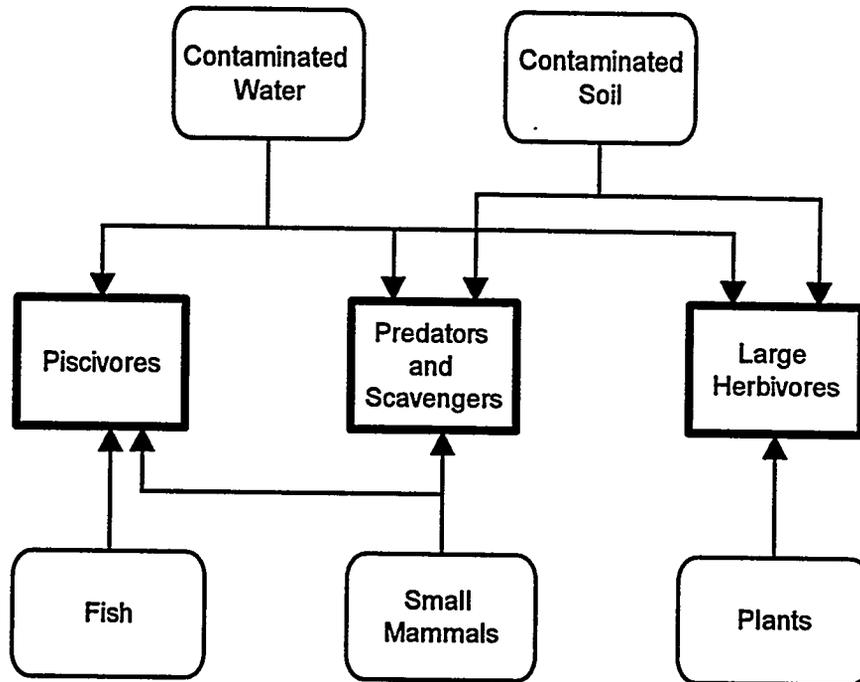


Fig. 2.4. Conceptual model of the mechanisms by which wide-ranging wildlife species are exposed to contaminants in White Oak Creek, its tributaries, and its floodplain.

3. RISKS TO FISH

3.1 EXPOSURE ASSESSMENT FOR FISH

3.1.1 Aqueous Chemical Exposure

Fish are exposed primarily to contaminants in water. Contaminants in water may come from upstream aqueous sources including the permitted outfalls, runoff, and the numerous seeps sampled and analyzed by the WAG 2 RI program. They may also come from internal cycling including exchange of materials between the surface water and contaminated sediments and exchange of contaminants between the biota and the water column. The consensus of the scientific community and of the EPA Office of Water is that aquatic biota should be assumed to be exposed to the dissolved fraction of the chemicals in water because that is the bioavailable form (Prothro 1993). However, EPA Region IV prefers to use total concentrations as conservative estimates of the exposure concentration. Therefore, total concentrations of metals are used in the exposure assessment for fish.

Because water in the OU is likely to be more variable in time than in space because of the rapid replacement of water, the mean water concentration within a subreach is an appropriate estimate of the chronic exposure experienced by fishes. The upper 95% confidence limit on the mean is an appropriately conservative estimate of this exposure for use in the contaminant screening. No distinction was made between storm flow and base flow samples in this screening analysis.

Many of the chemical concentrations in water were below analytical detection limits. Chemicals that were not detected in any sample in a reach were eliminated. If a reach contained some detects and some nondetects for a chemical, the product limit estimator was used to estimate the mean and its variance.

3.1.2 Radiation Exposures

Exposures to radionuclides are expressed as the dose rate received by the organism. The total dose rate is the sum of the internal and external dose rates, which are a function of exposure to the radionuclide and the characteristics of the radiation. Radiation exposures of fish in surface water are likely to be driven by internal exposures, especially for alpha emitters such as U^{234} and Th^{230} (Blaylock et al. 1993). Internal dose rates are based on the concentrations of the radionuclide in the organism, which were estimated by multiplying the water concentration times the biological concentration factors provided in Blaylock et al. (1993). External dose rates are based on the concentrations of the radionuclides in the surrounding water. Because the radiation exposure rate is expressed as absorbed dose per unit time (rads per day), the dose rates for each isotope and pathway can be added to determine the total dose rate.

As with estimates of aqueous chemical exposure, the 95% upper confidence limit (UCL) on the mean is an appropriately conservative estimate of exposure for use in the screening. As with chemical exposures, total concentrations of radionuclides were used in the radiation exposure assessment for fish. This results in conservative estimates of the internal dose and realistic estimates of external dose.

3.2 EFFECTS ASSESSMENT FOR FISH

3.2.1 Single Chemical Aqueous Toxicity

The screening benchmarks for aquatic biota are taken from Suter and Mabrey (1994). Because there are no standard screening benchmarks, sets of alternative benchmarks (described in Table 3.1) were calculated for each chemical. The benchmark preferred by the regulatory agencies is the chronic National Ambient Water Quality Criteria (NAWQC), but they are available for relatively few industrial chemicals. Secondary chronic values (SCV), which are conservative estimates of chronic NAWQC, were calculated for chemicals that do not have NAWQC. Other benchmarks are included to provide greater assurance of detecting all COPECs.

NAWQC that are functions of water hardness are corrected for site-specific conditions. For purposes of screening, we chose conditions that would constitute reasonable maximum toxicity, defined as conditions that would persist for seven days. This was a hardness of approximately 100 mg/L (Blaylock et al. 1992).

3.2.2 Radiological Effects

The recommended acceptable dose rate to natural populations of aquatic biota is 1 rad/d (NCRP 1991). Although there are no standard screening benchmarks for radionuclide concentrations in surface water, Blaylock et al. (1993) provide formulas and exposure factors estimating the dose rates to representative aquatic organisms. These formulas were used to calculate the water concentration that results in a total dose rate of 1 rad per day to a small fish for each radionuclide detected in the water samples from Bear Creek Valley (BCV). The exposure factors used in this assessment are from Blaylock et al. (1993) and are presented in Table 3.2. The screening values include internal and external exposures from the detected isotope and all short-lived daughter products. All major alpha, beta, and gamma emissions for each isotope were included, and the absorbed fraction of each was estimated from the graphs provided in Blaylock et al. (1993). Internal exposures from radionuclides in water were estimated using the biological concentration factors for the detected parent isotopes.

3.2.3 Ambient Water Toxicity

Toxicity tests of WAG 2 water are performed by the ORNL BMAP and are presented in their annual reports. The tests employed include the standard 7-day tests of growth and survival in fathead minnow larvae and fecundity and survival of *Ceriodaphnia dubia* and an early life stage test with Japanese medaka eggs and larvae. The standard tests did not show any aqueous toxicity in the last two years reported, 1992 and 1993. However, the more sensitive medaka embryo-larval test consistently showed toxicity in water collected adjacent to or downstream of ORNL relative to upstream and control waters (Ashwood 1994). Attempts to identify the causal agent determined that mortality was not affected by 0.1 micron filter to eliminate pathogens or by treatment with sodium thiosulfate to eliminate chlorine. However, toxicity was completely eliminated by filtration with activated charcoal, a nonspecific treatment for aqueous contaminants. Finally, in situ toxicity tests were conducted with caged snails (*Elimia clavaeformis*) exposed for seven days. Although snails are benthic invertebrates, they are discussed here because they were exposed to water and not sediment. Snails were killed in lower fifth creek in October 1992 and in WOC adjacent to ORNL in June and October 1992 (Ashwood 1993).

Table 3.1. Descriptions of the ecotoxicological screening benchmarks for aquatic biota

Benchmark	Abbreviation	Description
Acute National Ambient Water Quality Criteria	NAWQC_ACU	Current national criteria for protection of aquatic life from lethal effects in episodic exposures.
Chronic National Ambient Water Quality Criteria	NAWQC_CHR	Current national criteria for protection of aquatic life from lethal and sublethal effects in extended exposures. Criteria for uses of aquatic life (i.e., fish consumption) are not included.
Secondary acute value	S_ACU_V	Values estimated with 80% confidence to not exceed the unknown acute NAWQC. Used when data are inadequate to calculate the acute criterion.
Secondary chronic value	S_CHR_V	Values estimated with 80% confidence to not exceed the unknown chronic NAWQC. Used when data are inadequate to calculate the chronic criterion.
Lowest chronic value for fish	LCV_FISH	The lowest value, from acceptable fish chronic toxicity tests, of the geometric mean of the lowest observed effect concentration (LOEC) and the no observed effect concentration (NOEC).
Lowest chronic value for daphnids	LCV_DAPH	The lowest value, from acceptable daphnid chronic toxicity tests, of the geometric mean of the LOEC and the NOEC.
Lowest chronic value for nondaphnid invertebrates	LCV_ND	The lowest value of the geometric mean of the LOEC and the NOEC from acceptable chronic toxicity tests of nondaphnid invertebrate species.
Lowest plant value	LCV_AQPL	The lowest value from an acceptable daphnid chronic toxicity test of the geometric mean of the LOEC and the NOEC.
Lowest Test EC20 for fish	LTV_FISH	The lowest value, from acceptable fish chronic toxicity tests, of the lowest concentration causing at least a 20% reduction in the weight of young per female or the weight of young per egg.

Table 3.1 (continued)

Benchmark	Abbreviation	Description
Lowest test EC20 for daphnids	LTV_DAPH	The lowest value from an acceptable daphnid chronic toxicity test of the lowest concentration causing at least a 20% reduction in the product of survivorship, growth, and fecundity.

Note: More details are presented by Suter and Mabrey (1994).

Table 3.2. Exposure factors for the radionuclides detected in WAG 2 sediment and surface water and their short-lived progeny

Radionuclide ^a	Kd ^b	Biological Concentration Factor ^c	Emission Energies (MeV)				Absorption Factors			
			Average Alpha	Maximum Beta	Average Beta	Average Gamma	Beta for Small Invertebrates	Beta for Small Fish	Gamma for Small Invertebrates	Gamma for Small Fish
Cesium-137+Barium-137m	1000	2000		1.17e+00	2.52e-01	5.96e-01	0.6	0.99	0.003	0.015
Cobalt-60	45	300		1.48e+00	9.65e-02	2.50e+00	0.5	0.98	0.002	0.01
Hydrogen-3	N/A	1		1.80e-02	5.68e-03		1	1		
Strontium-90 + Yttrium-90	35	60		2.79e+00	1.13e+00	1.69e-06	0.25	0.83	0.4	0.6
Curium-244	2000	30	5.89e+00		8.59e-03	1.70e-03	1	1	0.4	0.6
Thorium-232	150000	100	4.07e+00		1.25e-02	1.33e-03	1	1	0.4	0.6
Radium-228					1.69e-02	4.14e-09	1	1	0.4	0.6
Actinium-228					4.60e-01	9.30e-01	0.95	1	0.003	0.015
Thorium-228	150000	100	5.49e+00		2.05e-02	3.30e-03	1	1	0.4	0.6
Radium-224			5.78e+00							
Radon-220			6.40e+00		8.19e-06	3.85e-04	1	1	0.4	0.6
Polonium-216			6.91e+00		1.61e-07	1.69e-05	1	1	0.4	0.6
Lead-212						1.75e-01	1.48e-01	1	1	0.002
Bismuth-212			2.22e+00	5.80e-01	4.69e-01	1.85e-01	0.95	1	0.002	0.012
Polonium-212			8.95e+00	2.25e+00			0.3	0.89		
Americium-241	700	30	5.57e+00		5.19e-02	3.24e-02	1	1	0.01	0.05

Table 3.2 (continued)

Radionuclide ^a	Kd ^b	Biological Concentration Factor ^c	Emission Energies (MeV)				Absorption Factors			
			Average Alpha	Maximum Beta	Average Beta	Average Gamma	Beta for Small Invertebrates	Beta for Small Fish	Gamma for Small Invertebrates	Gamma for Small Fish
Uranium-233	450	10	4.89e+00		6.08e-03	1.31e-03	1	1	0.4	0.6
Uranium-238	450	10	4.26e+00		1.00e-02	1.36e-03	1	1	0.4	0.6
Thorium-234					5.92e-02	9.34e-03	1	1	0.4	0.6
Protactinium-234m		10		2.29e+00	8.20e-01	1.13e-02	0.3	0.87	0.2	0.5
Plutonium-238	4500	30	5.58e+00		1.06e-02	1.81e-03	1	1	0.4	0.6
Uranium-234	450	10	4.84e+00		1.32e-02	1.73e-03	1	1	0.4	0.6
Thorium-230	150000	100	4.74e+00		1.46e-02	1.55e-03	1	1	0.4	0.6
Uranium-235	450	10	4.47e+00		4.80e-02	1.54e-01	1	1	0.002	0.012
Thorium-231					1.63e-01	2.55e-02	1	1	0.03	0.15

Sources: Soil-water partitioning coefficients are from Baes et al. (1984), biological concentration factors are from IAEA (1982), emission energies are from ICRP (1983), and absorbed fractions were estimated from graphs in Blaylock et al. (1993).

^a Designations of parent radionuclides and short-lived progeny are taken from Tables A.1 and A.2 in Blaylock et al. (1993). For naturally occurring alpha decay series, the parent is listed first, and progeny are listed immediately below in order of occurrence.

^b Soil-water partition coefficients are presented only for parent isotopes detected in WAG 2 sediments.

^c Biological concentration factors are presented only for parent isotopes detected in WAG 2.

3.2.4 Fish Community Survey

Analysis of fish population and community survey data provides a direct measure of impacts of human activities on aquatic ecosystems. The following results are taken from the most recent ORNL BMAP report (Ashwood 1993). The species composition of the fish community in the WOC watershed is stable at 13 species, which is less than other local streams. It is missing two families (Catastomidae and Percidae) and five genera (*Lythrurus*, *Luxilus*, *Notropis*, *Noturus*, and *Phoxinus*) that historically occurred in the stream.

3.2.5 Indicators of Fish Reproduction

Reproduction and indicators of reproduction were monitored in redbreast sunfish from 1989 to 1993. Indicators of reproductive condition of female redbreast sunfish improved over the period monitored until they nearly equal background values. However, reproductive success was highly variable with apparent reproductive failure occurring in lower WOC during the last reported year of monitoring. Since this failure was not associated with poor reproductive condition of the female fish prior to spawning, effects on the embryos or larvae were said to be the likely cause (Ashwood 1994).

3.3 RISK CHARACTERIZATION FOR FISH

3.3.1 Single Chemical Aqueous Toxicity

All chemicals detected in stream waters were screened against benchmarks. This was done by dividing the 95% UCL concentration (or maximum if the UCL could not be calculated because the chemical was detected only once) by each of the aqueous screening benchmarks (Table B.1). Chemicals that exceeded benchmarks were eliminated as COPECs if their UCL did not exceed twice the mean background concentration. EPA Region IV recommends this screening criterion. The reader should be aware that this practice in combination with the practice of using total metal concentrations, which is required by EPA Region IV, is potentially anticonservative because the concentration of particulate metal is being doubled along with the bioavailable dissolved fraction. This is potentially important because metals leached from wastes would be likely to be in soluble forms (e.g., chlorides or nitrates) that are potentially toxic, whereas, background concentrations of many metals in unfiltered water are likely to be predominately in the form of relatively insoluble minerals (e.g., sulfides or oxides), which are relatively nontoxic. If contamination doubled the bioavailable metal over background, this would not necessarily double the total metal concentration. The results of the screening are summarized in Table 3.3, and the COPECs for fish and chemicals that exceed NAWQC are in the following paragraphs.

In addition to stream waters, the WAG 2 program sampled waters from seeps and small ephemeral tributaries (Hicks 1996). Because these sites do not support fish communities, they are not assessed for risks to this endpoint. However, it is important to know which of these sources is contributing to risks in the streams. In addition, the state of Tennessee is concerned with whether the seep waters exceed water quality standards; therefore, the UCM or maximum concentrations for all chemicals detected in each seep are compared to their chronic NAWQCs or SCV (Table B.2). The use of the regulatory standard (NAWQC) or an estimate of the standard (SCV) addresses the regulatory concern and provides a means of highlighting those seeps that have high concentrations relative to toxicity without suggesting that the seeps support communities of fish and macroinvertebrates like the streams.

Table 3.3. Results of screening of chemicals that exceed benchmarks in whole or filtered water for chemicals of potential ecological concern (COPECs)

Chemical	COPEC	Reason for Inclusion or Rejection
Aluminum	Yes	Exceeds chronic National Ambient Water Quality Criteria (NAWQC) in all reaches and acute NAWQC in White Oak Lake (WOL) and Raccoon Creek Tributary reach (RAC). The chronic criterion is also exceeded in background water. Upper confidence limit (UCL) concentrations exceed twice background (0.34 mg/L) in 1C, WAG1 White Oak Creek (1WC), Northwest Tributary (NWT), RAC, West Seep (WS), and WOL; and the WOL mean exceeds twice background. It is entirely possible that the elevated Al is due to high particulate matter.
Barium	Yes	Exceeds the secondary chronic values (SCV) in all reaches and the secondary acute value (SAV) in Background White Oak Creek (BWC), Lower White Oak Creek (LWC), Waste Area Group 4 (W4T), and WS, but no other benchmarks. The upper confidence limit (UCL) concentration in BWC and both the mean and UCL concentrations in W4T exceed the twice background concentration (0.004 mg/L).
Beryllium	No	Exceeds the LTV daphnids but no other benchmarks in all reaches. That benchmark is apparently below the local background due to the compounding of effects across life stages in its derivation.
Boron	No	The Bo UCL exceeds the LTV for Daphnids in all reaches but no other benchmark. The LTV for Daphnids is much lower than other benchmarks for Bo. It is judged to be anomalous.
Cadmium	Yes	The Cd UCL concentration in W4T exceeds the chronic NAWQC, and the mean exceeds the lowest chronic value (CV) for daphnids. However, none of the concentrations exceed twice background (0.01 mg/L).
Calcium	No	The Ca UCL concentration in NWT and RAC slightly exceeds the CV for <i>Daphnia</i> but Ca would occur as nontoxic carbonates in those circumneutral and moderately hard streams rather than the toxic form (chloride) used in the test.
Chromium	Yes	The UCL exceeds the acute and chronic NAWQC, and the mean exceeds the chronic NAWQC in WOL. The WOL mean and the LMB UCL exceed the daphnid lowest CV. However, none of the concentrations exceed twice background (0.02 mg/L).
Cobalt	Yes	Exceeds the SCV in WS and Upper Melton Branch (UMB) and the daphnid LCV in WS in the one sample from each site in which it was detected. However, none of the concentrations exceed twice background (0.04 mg/L).

Table 3.3 (continued)

Chemical	COPEC	Reason for Inclusion or Rejection
Copper	Yes	Mean exceeds the chronic NAWQC in water from LWC, UMB, and W4T and exceeds the lowest CV for daphnids in all reaches where it was detected. However, none of the concentrations exceed twice background (0.023 mg/L).
Iron	No	Means exceeds chronic NAWQC in WS and WOL, and the daphnid LCV all reaches but BWC. However, none of the concentrations exceed twice background (0.81 mg/L).
Lead	Yes	Means in BWC and Intermediate Holding Pond (IHP) and the UCL in WOL exceed the chronic NAWQC. However, none of the concentrations exceed twice background (0.014 mg/L).
Manganese	Yes	Means in LMB, NWT, W4T, and WS exceed the SCV, and those in RAC and WS exceed the SAV and the lowest chronic values (CVs) for fish and daphnids. Mean concentrations exceed twice background (0.21 mg/L) in RAC, W4T, and WS.
Nickel	Yes	Means exceed daphnid and plant LCVs in LMB, W4T, and WS. However, none of the concentrations exceed twice background (0.04 mg/L).
Silver	Yes	Each of the single detected values at BWC, IHP, and W4T exceeds the SCV and lowest CVs for daphnids and fish. Silver was not detected at the background site.
Zinc	Yes	Mean in 1WC and the UCL in UMB exceed the chronic NAWQC. Lowest CVs for daphnids, fish, and plants exceeded in 1WC, IHP, UMB, and WOL. Only the UCL in 1WC exceeds twice background (0.22 mg/L).
Carbon disulfide	Yes	Exceeds the SCV in LMB, M.C., RAC, UMB, and WS. No background for organics.

The following text discusses those chemicals that meet all criteria for COPECs or all criteria except for the exceedence of twice background (Table 3.3). Those metals that exceed NAWQC but not background are retained in spite of the criteria because background needs to be better defined for the watershed and because of the potential problem, discussed above, of background particulate concentrations masking increases in bioavailable metals.

Aluminum. Aluminum is not known to be toxic in circumneutral streams, and one must suspect that the high concentrations are due to particulate aluminum. However, aluminum concentrations in streams clearly exceed background, so it cannot be eliminated. All but five of the 27 seeps exceed the NAWQC for Al, and RS-3a exceeds it by more than a factor of 100.

Barium. The exceedences of the SAV and SCV might be attributable to the conservatism of those benchmarks, which is a result of the poorly defined toxicity of this metal. However, the concentrations in W4T are quite high relative to background as well as benchmarks and therefore pose a credible risk to fish and other aquatic organisms. BTT, SW4-1, and SW4-2 are significant barium sources on W4T, with SW4-2 exceeding the SCV by more than a factor of 100.

Cadmium. Although cadmium does not occur at concentrations exceeding twice background in W4T and in two seeps (RS-3a and SW5-4, neither of which feeds W4T), it exceeds the chronic NAWQC.

Chromium. Although chromium does not occur at concentrations exceeding twice background, it exceeds the acute and chronic NAWQC in WOL and in the following seeps: East Seep, RS-3a, SW7-3, and WAG 4 T2A.

Copper. Although copper does not occur at concentrations exceeding twice background, it exceeds the chronic NAWQC in LWC, UMB, and W4T.

Carbon disulfide. Carbon disulfide was seldom detected in streams. The highest observed concentration, which is from approximately a factor of 100 lower than the lowest observed toxic concentration. The exceedence of the SCV in the screening assessment is attributable to the large safety factors used with minimal data sets. Carbon disulfide exceeds the SCV in 22/33 seeps; but it is a natural constituent of ground water, and the concentrations are not exceptionally high. Hence, there appears to be negligible risk to aquatic biota from carbon disulfide

Manganese. The exceedences of the SCV in six reaches might be attributable to the conservatism of that benchmark, which is a result of the poorly defined toxicity of this metal. However, the concentrations in RAC, W4T, and WS exceed the SAV and chronic toxic concentrations for aquatic life. Seeps SW4-1 and SW7-1 are a conspicuous contributors of manganese to the W4T and WS.

Nickel. Nickel exceeded chronically toxic concentrations in LMB, W4T, and WS. Nickel in seep SW4-2, which feeds reach W4T, is much higher than at any other location and exceeds the chronic NAWQC by more than a factor of 50.

Silver. The combination of a detection limit that is barely lower than the lowest CV, the failure to detect silver at the background site, and the small toxicity data set make interpretation of the hazard posed by silver uncertain. However, there is no basis for rejecting it as a COPEC. Silver was detected in only two seeps, RS-3A and SW2-4.

Zinc. The mean concentration of zinc exceeds the chronic NAWQC, and the UCL exceeds twice background in the vicinity of ORNL (reach 1WC). Therefore it is a clear COPEC. Zinc concentrations are not high in seeps except for SW9-2 which exceeds the chronic NAWQC. However, it feeds HRT, which does not have high zinc concentrations. This, and the fact that zinc was not detected above ORNL (reach BWC), suggests that the source of zinc is an effluent or an unidentified seep in the vicinity of ORNL.

Chemicals that were detected in water but for which no benchmarks are available are also of concern. They are silicon and cis-1,2-dichloroethene.

3.3.2 Single Chemical Internal Toxicity

Mercury. Mercury concentrations in largemouth bass, but not sunfish, increased from a mean of 0.11 to 0.57 mg/kg wet weight during the period 1988–1992 for unknown reasons (Ashwood 1994). The maximum concentrations in largemouth bass in the last two reported years (0.70 and 0.71 mg/kg in 1991 and 1992) exceeded the 0.66 mg/kg concentration that corresponded to a CV for fathead minnows but was well below the value for brook trout (5.6 mg/kg). The median concentration for largemouth bass and all concentrations for bluegill and redbreast sunfish were below 0.66 mg/kg.

PCBs. PCBs have been detected in sunfish, largemouth bass, and channel catfish in WOC with the highest concentrations found in channel catfish from the embayment (reach EWC) (Ashwood 1994). However, both mean and maximum concentrations in catfish in 1993 were below the concentration of PCB-1242 that reduced growth and caused liver hypertrophy in channel catfish at a body burden of 14.3 mg/kg (Hansen et al. 1976).

3.3.3 Radiation Risks

The concentrations of detected radionuclides were compared with the screening values for small fish. A hazard quotient (HQ) was calculated for each radionuclide by dividing the 95% UCL concentration by the screening value. The screening value is the concentration resulting in a total dose rate of 1 rad/d from the major emissions of the measured parent radionuclide and all short-lived progeny. At each location the HQs for all detected radionuclides were summed. Radionuclides present a significant risk if the sum of HQs for a site exceeds 1.0. Table B.3 details the exposure estimates (95% UCL), screening values, and HQs for individual radionuclides for all radionuclides detected at each location. Based on these results, radionuclides in BCV surface water appear to present a negligible risk to fish. That is, the individual and sum of HQs were <1.0 at all reaches for which data are available. Seeps and small ephemeral tributaries do not support communities of fish or other aquatic macroorganisms. However, they were also screened to determine whether they were significant sources. Although seeps SW2-6 and -7 and SW5-4 exceeded the screening benchmark for strontium-90, the nearby reaches are far below the benchmark so they are not contributing to a significant risk to aquatic life.

3.3.4 Ambient Water Toxicity

Water from the watershed was found to be lethal to snails and to medaka embryos and larvae. Snails were tested only in the vicinity of ORNL, and toxicity was attributed to chlorine on the basis of correlation. Medaka toxicity was tested in WOC but not its tributaries (Ashwood 1993). Toxicity was not observed above WAG 1 but occurred consistently downstream (Ashwood 1994). In the period 1991–1993 the lowest survival (<10%) was at the WOL weir. This pattern is

consistent with an accretion of contaminants in the creek. Since water was not chemically analyzed in conjunction with the toxicity tests and the toxicity identification and evaluation (TIE) was not completed, it is not possible to determine which contaminants may be responsible for the toxicity. However, preliminary TIE results indicate that a chemical other than chlorine was responsible.

In contrast, WOC water was not found to be toxic in the 7-day fathead minnow and *C. dubia* tests. This lack of response relative to the snail test was attributed to the insensitivity of the standard static renewal test to volatile chemicals such as chlorine (Ashwood 1993). The lack of response relative to medaka may be attributed to the inherent sensitivity of medaka or the longer duration of the medaka test and the inclusion of two life stages.

3.3.5 Biological Indicators

Results of the studies of histological, physiological, and reproductive indicators are presented by Ashwood (1994). Although these indicators show improvement, reproductive indicators for redbreast sunfish can not be interpreted as indicative of effects on the fish community endpoint; they are suggestive of inhibited reproduction in individuals of one species.

3.3.6 Biological Surveys

Surveys of the fish community in WOC indicate a significant loss of species and no recent improvement. Although it seems likely that the original loss was due to contaminants, it is not clear that the current contaminant levels would cause the same effect. The dam and numerous weirs inhibit movement of fish and may be preventing recovery of the community which would otherwise occur.

3.3.7 Weight Of Evidence For Fish

The weighing of evidence is performed by asking the following questions concerning each reach in the watershed:

- Is the fish community less species-rich or abundant than would be expected?
- Do individual fish display injuries that are indicative of significant toxic effects?
- Is the water toxic to aquatic organisms?
- Does that water contain chemicals in toxic amounts?
- Do the fish contain chemicals in toxic amounts?
- What factors account for apparent discrepancies in the results?
- What is the likelihood that the fish community is at least 20% less species rich or abundant than it would be in the absence of contamination?

The available evidence for each reach is summarized below. However, some lines of evidence are not available for every reach due to lack of data (Table 3.4).

Table 3.4 Availability of biological data related to the fish community endpoint in the White Oak Creek watershed reaches

Reach	Fish community	Standard tests	Medaka test	Snail test	Redbreast bioindicators	Fish bioaccumulation
EWC						X
WOL			X			X
LWC	X		X		X	X
M.C.	X	X	X		X	X
IHP	X	X	X			
IWC	X	X	X	X		X
BWC	X	X	X	X		
LMB	X	X				X
UMB	X	X				
WS						
W4T						
HRT						
NWT	X	X				X
RAC						
1C	X			X		X
5C	X			X		X

Source: Ashwood 1993, 1994

3.3.7.1 Mainstem reaches

White Oak Creek Embayment (EWC). The WAG 2 program did not sample this reach and biological data are not available. Its water quality would be expected to be similar to WOL but with some dilution from the Clinch River.

White Oak Lake (WOL). Because there are no good reference lakes for WOL, the effects on the fish community of the lake cannot be directly evaluated. WOL effluent water has been lethal to medaka embryos and larvae. WOL water exceeds NAWQC values for Al, Cr, Fe, and Pb as well as levels of Ba, Cu, Mn, and Zn that are toxic to aquatic life.

Lower White Oak Creek (LWC). Reach LWC has a depauperate fish community relative to similar streams; it has experienced reproductive failures of redbreast sunfish, and its waters have been lethal to medaka embryos and larvae. LWC water exceeds chronic NAWQCs for aluminum and copper and contains potentially toxic concentrations for barium and iron.

Middle White Oak Creek (MC). Reach MC has a depauperate fish community relative to similar streams. It has experienced reproductive failures of redbreast sunfish, and its waters have been lethal to medaka embryos and larvae. MC water exceeds the chronic NAWQC for aluminum and has potentially toxic concentrations for Ba, Cu, Fe, and carbon disulfide.

Intermediate Holding Pond (IHP). Reach IHP has a depauperate fish community relative to similar streams. Its waters have been lethal to medaka embryos and larvae. LWC water exceeds the chronic NAWQC for aluminum and lead and has potentially toxic concentrations for Ba, Cu, Fe, Ag, and Zn.

WAG1 White Oak Creek (1WC). Reach 1WC has a depauperate fish community relative to similar streams, and its waters have been lethal to medaka embryos and larvae and to snails. 1WC water exceeds the chronic NAWQC for aluminum and lead and contains potentially toxic concentrations for Ba, Cu, Fe, Ag, and Zn. Chlorinated effluents have been a source of severe episodic toxicity in this reach.

Background White Oak Creek (BWC). Reach BWC waters have not been found to be toxic. BWC water exceeds the chronic NAWQC for aluminum and lead and has potentially toxic concentrations for Ba, Cu, and Ag.

Lower Melton Branch (LMB). Reach LMB has a depauperate fish community relative to similar streams. Reach LMB water exceeds the chronic NAWQC for aluminum and contains potentially toxic concentrations for Ba, Cr, Cu, Fe, Mn, Ni, and carbon disulfide.

Upper Melton Branch (UMB). Reach UMB has a depauperate fish community relative to similar streams. Reach UMB water exceeds the chronic NAWQC for Al, Cu, and Zn as well as containing potentially toxic concentrations for Ba, Cr, Co, Fe, and carbon disulfide.

3.3.7.2 Tributary reaches

West Seep (WS). WS water exceeds the chronic NAWQC for aluminum and iron as well as containing potentially toxic concentrations for Ba, Co, Cu, Mn, No, and carbon disulfide.

WAG 4 Tributary (W4T). W4T water exceeds the chronic NAWQC for Al, Cd, and Cu as well as containing potentially toxic concentrations for Ba, Cr, Fe, Mn, No, and Ag.

Homogeneous Reactor Test Tributary (HRT). HRT water exceeds the chronic NAWQC for aluminum as well as containing potentially toxic concentrations for Cr, Cu and Fe.

Northwest Tributary (NWT). Reach NWT has a depauperate fish community relative to similar streams. NWT water exceeds the chronic NAWQC for aluminum as well as having potentially toxic concentrations for Ba, Cu, Fe, and Mn.

Raccoon Creek (RAC). RAC water exceeds the chronic NAWQC for aluminum as well as containing potentially toxic concentrations for Cu, Fe, Mn, and carbon disulfide.

3.3.8 Summary of Risk Characterization for Fish

Because this is a screening assessment, the goal of the risk characterization is to determine whether risks to fish can be eliminated as an issue or whether there is a credible hazard that

should be assessed further. Even if the NAWQC for Al is set aside as inappropriate to this site, credible NAWQCs and CVs are exceeded in nearly all reaches. The toxicity to medaka embryos and larvae provides important supporting evidence that waters in WOC are toxic to fish. The low species richness of the stream is consistent with effects but could have physical causes. Although all of these lines of evidence are consistent with a hazard to fish, the evidence is not conclusive. The high metal concentrations could be nonbioavailable, the medaka could be much more sensitive than any of the fish species native to the stream, and the low species richness could be due to physical barriers to recovery. In particular, it should be noted that three years have passed since the collection of the most recent biological data that were available for this assessment. At that time, the bioindicators data suggested that the condition of fish in WOC was improving and was approaching that of fish in reference streams. It is possible that the fish community has largely recovered and that toxicity would no longer be observed in WOC at this time.

3.4 UNCERTAINTIES CONCERNING RISKS TO FISH

The following issues constitute the major sources of uncertainty in the risk assessment for the fish community.

The bioavailable concentrations of chemicals in water are unknown. In assessments that included analyses of filtered as well as unfiltered water, concentrations of Al and some other metals that were above criteria in the unfiltered water were shown to be associated with particles (i.e., not bioavailable).

The toxicity of water in seven reaches is unknown.

The high observed mortality in medaka embryos and larvae are believed to constitute toxic effects that would result in significant effects on the fish community. However, this test is sensitive and has not been validated against biological survey data at sites where clear toxic effects are occurring and fish community properties are clearly related to toxic effects as has been done with the standard 7-day tests.

4. RISKS TO BENTHIC INVERTEBRATES

4.1 EXPOSURE ASSESSMENT FOR BENTHIC INVERTEBRATES

4.1.1 Chemicals In Sediment

Benthic invertebrates are exposed to contaminants in water or sediments. Those benthic invertebrates that inhabit riffles live on rocks and organic debris and are primarily exposed to contaminants in water. As with fish, exposures of benthic invertebrates to water are conservatively estimated as total concentrations, although actual exposures are best estimated by dissolved phase concentrations. Benthic invertebrates that inhabit the soft sediments of pools are exposed to contaminants in the sediments. Two different expressions of sediment contamination are used in ecological risk assessments: whole sediment concentrations and filtered pore water concentrations. The use of pore water is based on the assumption that chemicals associated with the solid phase are largely unavailable and therefore sediment toxicity can be estimated by measuring or modeling the pore water concentration. This is the approach the EPA uses to calculate sediment quality criteria. Whole sediment concentrations do not account for effects of sediment properties on bioavailability; however, they are required by EPA Region IV and may provide a better estimate of risk for highly particle-associated chemicals, which may not be detectable in pore water.

Benthic invertebrates are exposed to surface sediments and not to deep sediments. In a compromise between realism and the desire to include data for all analytes, a maximum interval of 0 to 50 centimeters was used. Analyses of cores with a depth greater than 50 centimeters were discarded, and core sections other than the surface section were discarded.

Because sediment is likely to be more variable in space than in time (due to its relative immobility) and because the organisms are relatively immobile, it is not appropriate to think of benthic invertebrates as averaging their exposures to sediment over space or time. Therefore, the median sediment concentration is an appropriate measure of the central tendency of the contaminant data. An appropriate conservative estimate of this exposure for use in the contaminant screening is the maximum.

As with water, many of the chemical concentrations in sediments are below analytical detection limits. Chemicals that were not detected in any sample in a reach were eliminated. If a reach contained some detects and some nondetects for a chemical, the product limit estimator was used to estimate the mean, which is reported for reference.

Ford et al. (1996) presented a complete description of the sediment sampling and analysis in WAG 2. Note that sediment samples from the lower W4T were aggregated with the IHP samples and that samples from WS were aggregated with LWC samples for consistency with that report.

4.1.2 Sediment Radiation Exposures

Exposures to radionuclides are expressed as the dose rate received by the organism. As with radiation exposures in surface water, the total dose rate is the sum of the internal and external dose rates, which are a function of exposure to the radionuclide and the characteristics of the radiation. Radiation exposures to benthic invertebrates immersed in sediments are likely to be

driven by external exposures to contaminated sediments. The exception is for alpha emitters, such as uranium-234 and thorium-230. Internal dose rates are based on the concentrations of the radionuclide in the organism, which is a function of bioavailability. Pore water concentrations are assumed to best estimate the bioavailable fraction in sediments, and standard sediment to benthic invertebrate transfer factors are not available. Therefore, the concentration in benthic invertebrates was estimated by using the measured sediment concentrations and the soil-water partition coefficients (K_d) from Baes et al. (1984) to estimate the sediment pore water concentrations. Concentrations in the organism were then estimated based on the biological concentration factors provided in Blaylock et al. (1993). Because the radiation exposure rate is expressed as absorbed dose per unit time (rads/d), the dose rates for each isotope and pathway can be added to determine the total dose rate.

4.2 EFFECTS ASSESSMENT FOR BENTHIC INVERTEBRATES

4.2.1 Single Chemical Sediment Toxicity

Because there are no standard screening benchmarks and sediment quality criteria for only a few chemicals, sets of alternative sediment benchmarks were derived for each chemical (Table 4.1). Whole sediment concentrations are compared to these alternative benchmarks. The use of multiple benchmarks provides greater assurance of detecting all COPECs. Sediment quality criteria are corrected for site-specific conditions. Sediment benchmarks derived using the equilibrium partitioning method are calculated using location-specific percent organic carbon.

4.2.2.2 Radiological effects

The recommended acceptable dose rate to natural populations of aquatic biota is 1 rad/d (NCRP 1991). Although there are no standard screening benchmarks for radionuclide concentrations in sediments, Blaylock et al. (1993) provide formulas and exposure factors estimating the dose rates to representative aquatic organisms. These formulas were used to calculate the sediment concentration that results in a total dose rate of 1 rad/d to a small invertebrate for each radionuclide detected in the sediment samples from BCV. The exposure factors used in this assessment are from Blaylock et al. (1993) and are presented in Table 3.2. The benchmarks include internal and external exposures from the detected isotope and all short-lived daughter products. All major alpha, beta, and gamma emissions for each isotope were included; and the absorbed fraction of each was estimated from the graphs provided in Blaylock et al. (1993). Conservatism was practiced by assuming the representative invertebrate was immersed in the sediments at all times. Internal exposures from radionuclides in sediments were estimated using the soil-water partition coefficients and biological concentration factors for the detected parent isotopes.

Table 4.1 Descriptions of the ecotoxicological screening benchmarks for benthic biota exposed to contaminated sediments

Benchmark	Abbreviation	Description
Effects Range-Low	ER_L	The tenth percentile of estuarine sediment concentrations reported to be associated with some level of toxic effects.
Effects Range-Median	ER_M	The fiftieth percentile of estuarine sediment concentrations reported to be associated with some level of toxic effects.
Region IV Benchmark	REG_IV	The higher of two values, the EPA Contract Laboratory Program Practical Quantification Limit (CLP PQL) and the Effects Value which is the lower of the ER-L and the Florida NOEL.
Threshold Effect Level	TEL	The geometric mean of the fifteenth percentile of reported concentrations which were associated with some level of effects and the fiftieth percentile of reported concentrations which were associated with no adverse effects. All data are for marine and estuarine sediments.
Probable Effect Level	PEL	The geometric mean of the fiftieth percentile of reported concentrations which were associated with some level of effects and the eighty-fifth percentile of reported concentrations which were associated with no adverse effects. All data are for marine and estuarine sediments.
National Sediment Quality Criteria	EPASQC_A	Sediment quality criteria based on toxicity in water expressed as chronic water quality criteria (recalculated after adding some benthic species) and partitioning of the contaminant between organic matter (1% of sediment by weight) and pore water. Sediment quality criteria were adjusted to the site-specific percent total organic matter content.
Equilibrium Partitioning Benchmark	EQPSQB_A	Benchmarks derived in the same manner as sediment quality criteria except that the expression of aqueous toxicity is the chronic NAWQC or the secondary chronic value, and site-specific percent organic matter is used.

Table 4.1 (continued)

Benchmark	Abbreviation	Description
Ontario Ministry of the Environment Lowest Effect Level	LEL_MOE	Concentrations determined by the Ontario MOE to constitute thresholds for toxic effects in Ontario sediments.
Ontario Ministry of the Environment Severe Effect Level	SEL_MOE	Concentrations determined by the Ontario MOE to constitute thresholds for severe toxic effects in Ontario sediments.
Region V Benchmark for nonpolluted sediments	REG_V_L	A concentration determined to constitute background for sediments in Illinois.
Region V Benchmark for heavily polluted sediments	REG_V_M	A concentration determined to constitute the upper bound of moderately polluted sediments relative to background sediments in Illinois.
Apparent Effect Threshold	AET	A concentration above which toxic effects occurred at all sites in Puget Sound.

Note: More details are presented by Hull and Suter (1994), Long et al. (1995), MacDonald (1994), and Region IV (1994). The last five benchmarks are used only when none of the first seven are available

4.2.2 Invertebrate Community Surveys

4.2.2.1 Sediment communities

Surveys of benthic invertebrates in soft sediments behind weirs and dams were performed in 1995 (Appendix A). The surveys were conducted in large part to determine whether dredging these sediments to maintain pool depth would destroy communities that had any particular ecological value. However, they serve to indicate the effects of sediment contamination more generally including natural pools where sediments accumulate. The survey included two weirs on WOC within WAG 2, two on Melton Branch, and two sites in WOL. These were compared to reference pools behind weirs on upper First Creek, WOC above ORNL, and Clear Creek. The locations are described in Appendix A. The results are somewhat confounded by differences in sediment characteristics among the weirs. The reference weirs tended to have more coarse particulate organic matter (detritus) and less silt than the weirs in WAG 2. This is not surprising given the silt that would be expected to enter the streams from construction and remediation activities at ORNL. The two WOL sites are different from all of the weirs in the depth of water and associated physical characteristics.

The lowest taxonomic richness was found in WOL (~ 4 taxa per sample), and the highest was found in background WOC (~ 15 taxa/sample) and upper Clear Creek (~ 12 taxa/sample). The other two reference weirs and the WAG 2 weirs all had (~ 9-11 taxa/sample). Since chironomid midge larvae made up most of the taxa identified, they were tabulated separately and gave the same pattern of relative taxonomic richness. Hence, although taxonomic richness varies by more than a factor of three among sites, the difference could be attributed to physical habitat differences as well as contamination.

4.2.2.2 Riffle communities

Surveys of benthic invertebrates in riffles were performed beginning in 1987 with the last reported results from 1992 (Ashwood 1994). At that time, the taxonomic richness of all invertebrates and particularly of the Ephemeroptera, Plecoptera, and Trichoptera (EPT) was severely reduced at sites on WOC adjacent to and downstream of ORNL. They were also reduced in lower First and Fifth Creeks and in Melton Branch relative to reference sites. However, they were elevated in lower NWT relative to the upstream reference site. These comparisons of upstream and downstream sites are confounded by habitat differences as with the soft sediment communities. However, the differences in substrate are not as great, and because riffle communities have been much more studied than soft sediment communities, it is possible to judge which differences are attributable at least in part to contamination. Therefore, it can be judged with some confidence, that the reduced taxonomic richness was not due simply to stream habitat gradients (Ashwood 1994).

4.3 RISK CHARACTERIZATION FOR BENTHIC INVERTEBRATES

4.3.1 Single Chemical Sediment Toxicity

Screening against benchmarks. Chemicals detected in whole sediments were screened against benchmarks and evaluated as COPECs. An HQ was calculated for each chemical by dividing the maximum concentration by the sediment benchmarks (Table B.5). Chemicals that exceeded any benchmark (e.g., HQ>1) were examined further to determine whether they were

Table 4.2. Results of screening of chemicals that exceed benchmarks in sediments for COPECs

Chemical	COPEC	Reason for Inclusion or Rejection
2-methylnaphthalene	No	Detected only in two reaches and only one and two samples; it barely exceeds the effects range-low (ER-L) in one sample.
Acenaphthene	Yes	Exceeds all benchmarks in Middle White Oak Creek (MWC) and multiple benchmarks in Intermediate Holding Pond (IHP) and Lower White Oak Creek (LWC).
Anthracene	Yes	Exceeds all benchmarks in MWC and multiple benchmarks in IHP and LWC.
Aroclor - 12xx		See PCBs
Arsenic	Yes	Exceeds the threshold effect level (TEL) in all reaches and the ER-L in all but LMB but not the ER-M or PRE in any reach.
Barium	Yes*	Exceeds both Region V benchmarks
Benzo(a)anthracene	Yes	Exceeds multiple benchmarks in IHP and MWC
Benzo(a)pyrene	Yes	Exceeds multiple benchmarks in IHP and MWC
Cadmium	Yes	Exceeds multiple benchmarks in IHP and MWC
Chromium	Yes	Exceeds the ER-L in IHP and LMB
Chrysene	Yes	Exceeds multiple benchmarks in IHP and MWC
Copper	Yes	Exceeds multiple benchmarks in IHP, LMB, LWC, and MWC
Dibenz(a,h)anthracene	Yes	Exceeds the ER-L and TEL in IHP and MWC and the PEL in IHP, but detected in only one sample/reach.
Fluoranthene	Yes	Exceeds the ER-L and TEL in IHP and MWC and the TEL in LWC.
Fluorene	Yes	Exceeds the ER-L and PEL in IHP and MWC.
Iron	Yes*	Exceeds the Ontario MOE's SEL.
Lead	Yes	Exceeds the ER-L and TEL in IHP and MWC and barely exceeds the Region IV value in LWC and the PEL in MWC.

Table 4.2 (continued)

Chemical	COPEC	Reason for Inclusion or Rejection
2-methylnaphthalene	No	Detected only in two reaches and only one and two samples; it barely exceeds the effects range-low (ER-L) in one sample.
Acenaphthene	Yes	Exceeds all benchmarks in Middle White Oak Creek (MWC) and multiple benchmarks in Intermediate Holding Pond (IHP) and Lower White Oak Creek (LWC).
Anthracene	Yes	Exceeds all benchmarks in MWC and multiple benchmarks in IHP and LWC.
Aroclor - 12xx		See PCBs
Manganese	Yes*	Exceeds MOE benchmarks in all reaches.
Mercury	Yes	Exceeds multiple benchmarks in all reaches but particularly high in IHP and MWC.
Nickel	Yes	Exceeds multiple benchmarks in all reaches
PCB, total	Yes	Exceeds multiple benchmarks in all reaches but particularly high in IHP and MWC.
Phenanthrene	Yes	Exceeds multiple benchmarks in IHP, LWC and MWC but particularly high in MWC.
Pyrene	Yes	Exceeds the ER-L in IHP, and MWC
Silver	Yes	Exceeds multiple benchmarks in IHP and MWC.
Zinc	Yes	Exceeds multiple benchmarks in IHP, LMB, and MWC.

*These COPECs are based on weak benchmarks which are the only ones available.

credible COPECs (Table 4.2). Only 2-methylnaphthalene could be eliminated without comparison to background and a more detailed evaluation of their distribution among reaches and the relevant effects data. These comparisons will be included in the White Oak Creek watershed RI.

Chemicals that were detected in sediment but for which no benchmarks are available are also of concern. They are aluminum, benzo(p)fluoranthene, benzo(g, h, I) perylene, benzo(k)fluoranthene, beryllium, boron, butylbenzylphthalate, carbazole, cobalt, indeno(1,2,3-cd)pyrene, lithium, methoxychlor, molybdenum, osmium, selenium, strontium, thallium, tin, and hexachlorobenzene.

4.3.2 Sediment Radiation Characterization

The concentrations of detected radionuclides were compared with the screening values for small invertebrates immersed in sediments. An HQ was calculated for each radionuclide by dividing the measured concentration by the screening value. The screening value is the concentration resulting in a total dose rate of 1 rad/d from the major emissions of the measured parent radionuclide and all short-lived progeny. A total dose rate to small invertebrates at each site was calculated by summing the HQs for all detected radionuclides. Radionuclides present a significant risk to benthic invertebrates if the sum of HQs for a site exceeds 1.0. Table B.5 details the detected concentrations, screening values, HQs for individual radionuclides, and the sum of HQs for all radionuclides detected at each site. Based on these results, radionuclides in BCV sediments appear to present a negligible risk to benthic invertebrates. That is, the individual and sum of HQs were considerably <1.0 at all locations.

4.3.3 Benthic Community Surveys

The benthic community survey data cannot be directly related to the chemical analyses because WOL and the community reference sites were not subject to chemical analyses and because differences in the sampling would tend to obscure patterns. The communities in the four pools on WOC and MB within WAG 2 had total taxonomic richnesses that were within the range of reference communities although not as high as the best site on upper WOC (Appendix A). The only indication of a contaminant effect in the soft sediment community is chironomid taxonomic richness (by far the most abundant taxon) in MWC (the most contaminated sediment), which was lower than in reference pools.

4.3.4 Weight of Evidence for Benthic Invertebrates

PCBs (both totals and Arochlor mixtures), 10 polycyclic aromatic hydrocarbons (PAHs), and 12 metals were found at concentrations high enough to indicate a hazard to benthic invertebrates. Sediment toxicity is unknown. Community surveys in depositional areas do not indicate a large effect of contamination except possibly in WOL, but they are confounded by habitat differences. Sediment chemical concentrations at most benthic community survey sites are unknown and were not available for most tributaries. Hence, the sediments are contaminated, but the benthic community is not clearly different from reference communities; or in the case of WOL, the community is different, but contaminant levels are not known.

The communities of riffles have low species richness below ORNL, which is likely to be primarily due to contaminants in water (Chap. 3). The dominance of this mode of exposure is the reason that riffle invertebrate communities are monitored to determine whether water quality goals are being met (Ashwood 1994). However, the feeding habits of some species expose them

to the small amount of particulate material that is deposited in such areas so sediment contamination may make a small contribution to this effect.

4.4 UNCERTAINTIES CONCERNING RISKS TO BENTHIC INVERTEBRATES

The following issues are believed to constitute the major sources of uncertainty in the risk assessment for the benthic invertebrate assemblage.

- The benchmarks against which the sediment concentrations are screened are from studies of estuaries and large lakes that may differ considerably in their physical and ecological properties from the streams being assessed.
- The toxicity of the sediments is unknown.
- The chemical concentrations in WOL and in most tributaries are unknown.
- Chemical concentrations in sediments from most of the benthic invertebrate survey sites are unknown.

5. RISKS TO SOIL INVERTEBRATES

This chapter addresses risks from chemical soil invertebrates, which are represented by earthworms. Radiological risks to earthworms are presented in this report along with other terrestrial endpoints.

5.1 EXPOSURE ASSESSMENT FOR SOIL INVERTEBRATES

Earthworms absorb a variety of inorganic and organic soil contaminants through both feeding in soil and litter and burrowing in soil. It is assumed that these organisms represent highly exposed soil invertebrates. It is not possible or necessary to distinguish the exposure of earthworms to solid, liquid, and gaseous phases of the soil. Both in toxicity tests and the field, earthworms integrate exposures to all soil components by all uptake routes.

The bioavailability of the contaminants in soil and decomposing plant litter are controlled by soil, litter, and analyte characteristics; however, current knowledge is not sufficient to allow the prediction of bioavailability based on soil variables. An exception is the association of extremes in pH and metal availability (van Gestel 1992).

It is assumed that the earthworms spend their entire life span in soil with contaminant levels represented by the 0.5 m depth of soil sampled. The assumption is reasonable, though organisms feeding on well-decomposed organic material near the surface may be exposed to different quantities of organic chemicals than worms feeding on litter pulled down into burrows in the subsoil (Curl et al. 1987). Few earthworms are found in waterlogged soils so exposure to seeps is not considered. Because earthworms and other soil invertebrates may spend their entire lives at a single sampling location, the maximum detected contaminant concentrations were screened against toxicity benchmarks, as described in Sect. 5.2.1. These concentrations are used to assess potential negative impacts of contaminated soils on soil invertebrate populations.

5.2 EFFECTS ASSESSMENT FOR SOIL INVERTEBRATES

Toxicity tests and biological surveys were not performed at these sites. Earthworms were sampled, but quantitative surveys were not performed. Thus the only line of evidence in the risk assessment is the screening against benchmarks, described in the following paragraphs. Where benchmarks were not available, risks could not be evaluated.

Contaminant concentrations in soils were compared to toxicological benchmarks for earthworms in order to screen out chemicals that do not constitute a hazard to soil invertebrates and therefore do not require detailed risk characterization. Chemicals that were not screened out because the maximum detected concentrations were greater than benchmarks may potentially pose significant risks to soil invertebrates. References used for the derivation of benchmarks were reports of toxicity tests of individual chemicals in laboratory, greenhouse, or field settings. Tests conducted with natural soils in the laboratory were assumed to represent the exposures of worms in floodplain soils. However, chemicals freshly added to soils in toxicity tests may be more bioavailable than the chemical forms that occur in field soils. Only experiments in which earthworms were exposed to soil (natural or artificial mixture of natural components), soil/litter microcosms, or manure were considered. Twenty percent reduction in growth, reproduction, or

activity was used as the threshold for significant effects to be consistent with other screening benchmarks for ecological risk assessment and because the FFA parties adopted this level of effects for ecological endpoints (Suter et al., 1995).

The method used for deriving soil benchmarks are described in Will and Suter (1995a). Screening benchmarks for toxic effects of contaminants present in WAG 2 soils are presented with the hazard quotients in Table B.6. Benchmarks were not available for any of the organic compounds detected in the floodplain, except n-nitrosodiphenylamine.

5.3 RISK CHARACTERIZATION FOR SOIL INVERTEBRATES

The evidence concerning risks to soil invertebrates consists of analyses of inorganic, organic, and radionuclide contaminants in soil. This single line of evidence is summarized in the following text. Toxicity tests and quantitative biological surveys were not available.

5.3.1 Single Chemical Soil Toxicity

Contaminants of potential ecological concern (COPECs) for soil invertebrates in the WAG 2 floodplain were identified by comparing maximum concentrations in soil at each location to the following: (1) twice the mean background for soils from WAG 5 (DOE 1995); and (2) available benchmarks for toxicity to soil invertebrates (Will and Suter 1995a). Maximum detected concentrations in soil are used for screening purposes because earthworms and other soil invertebrates are relatively immobile and can therefore be exposed to the maximum concentration during the course of their lifetimes (Suter et al. 1995). If both background and benchmark criteria were exceeded, the contaminant was identified as a COPEC. The magnitude of exceedence of a benchmark is described by the hazard quotient, the maximum concentration in soil divided by the toxicity benchmark concentration. If background criteria were not available, exceedence of a benchmark was sufficient for the COPEC designation. Benchmarks, maximum concentrations of analytes at each sample location exceeding background for which benchmarks are available, and hazard quotients for the contaminants are given in Table B.6. The major COPECs for soil invertebrates were mercury and chromium. Mercury and chromium are among the few chemicals for which the benchmarks for toxicity to soil invertebrates is lower than the phytotoxicity benchmarks.

Intermediate Holding Pond. The maximum detected concentrations of chromium, mercury, and nickel were 100, 800, and 2 times the benchmarks for toxicity to earthworms.

Lower White Oak Creek. The maximum detected concentrations of chromium and mercury were 200 and 50 times the benchmarks for toxicity to earthworms. Although the maximum detected concentration of zinc is apparently equivalent to the benchmark if a single significant digit is used, the concentration of 190 mg/kg is lower than the benchmark concentration of 200 mg/kg. Also, the mean concentration of zinc in the soils of the IHP reach was about half that of the benchmark concentration. Thus, zinc is not a COPEC for toxicity to soil invertebrates.

Middle White Oak Creek. The maximum detected concentration of mercury was 30 times the benchmark for toxicity to earthworms.

Lower Melton Branch. The maximum detected concentration of mercury was 20 times the benchmark for toxicity to earthworms.

The strength in the literature toxicity line of evidence is in its ability to relate the concentration of a single contaminant in soil to observed effects. The screening is most useful if the chemical species and bioavailability of contaminants in WAG 2 floodplain soils are comparable to those of the chemical in laboratory toxicity tests in other soils. In these soils, the fractions of mercuric chloride, methyl mercury, mercuric sulfide, and some others were not determined. Thus, the hazard quotients for mercury may be quite conservative.

5.3.3 Summary of Risk Characterization for Soil Invertebrates

As in any screening assessment, potential hazard is indicated by the exceedence of toxicological benchmarks. COPECs for soil invertebrates in the WAG 2 floodplain include chromium, mercury and nickel. Further studies, such as toxicity tests (or other measures of bioavailability) and chemical speciation of mercury, would be advisable before concluding that these contaminants constitute hazards to soil invertebrates.

5.4 UNCERTAINTIES CONCERNING RISKS TO SOIL INVERTEBRATES

The following factors create uncertainty in assessing the risk posed by the potential contaminants of concern in soils:

'Bioavailability' of elements. The extraction methods used during sample collection may remove from the soil quantities of elements and compounds greater than those that are extracted from soil as they pass through the gut of the earthworm or those that are adsorbed dermally.

Variable response to contaminants. Various earthworm and other soil invertebrate species and earthworm growth stages tolerate contaminants to different degrees. The literature from which benchmarks were derived is not necessarily based on experiments using earthworm species known to be representative of those occurring in the soils of WAG 2.

Multiple contaminant exposure. Toxicity benchmark concentrations are derived from experiments in which earthworms are exposed to single contaminants. Typical soils of the WAG 2 expose earthworms to multiple contaminants which may not be adequately assessed on the basis of literature-derived benchmarks.

Lack of benchmarks for some metals and most organic compounds. Little research has been conducted on the toxic effects of the organic compounds, other than pesticides. Thus, it is not possible to assess the risk to earthworm populations posed by many of the analytes found in the soils.

Chemical speciation. Earthworm toxicity benchmarks for metals are often derived from studies with a single species of a contaminant.

6. RISKS TO PLANTS

This chapter addresses risk from chemicals to plants. Radiological risks to plants are assessed along with other terrestrial endpoints.

6.1 EXPOSURE ASSESSMENT FOR PLANTS

Vegetation growing in the WAG 2 floodplain is exposed to contaminants in soil and potentially in shallow groundwater near seeps and intermittent tributaries.

6.1.1 Soil Exposures

Concentrations of analytes in the soil derived from double acid extractions (nitric and hydrochloric) generally representing the total potential reservoir of contaminants, not the concentrations to which plants are exposed at any one time. Some metals are more readily available for uptake by plants than others, depending on the solubility and species of the compound, interactions with soil constituents (organic matter and clay) and with other contaminants, and mechanisms for bioaccumulation in particular plant species. Organic contaminants may interact strongly with soil organic matter and therefore only be partially available for plant uptake.

Contaminants have vertical distributions in soil that reflect interactions between the soil and the contaminant, controlled by the chemical and physical characteristics of the soil and the quantity and chemical characteristics of the contaminant. The exposure of plant roots to a contaminant in the soil depends on these abiotic factors and the growth characteristics of individual plants, such as rooting depth and density. Deep-rooted trees and shallow-rooted grasses may be exposed to different concentrations of contaminants, depending on the location of the source. In the absence of data on root distribution, it is assumed that the plants are rooted within the zone from which the soil was sampled for analysis.

Two other routes of exposure cannot be addressed because of a lack of data. These are exposure to volatile organic compounds in the air above and in the soil and contaminants in dust particles deposited on above-ground plant parts. These routes are assumed to contribute negligibly to plant exposure in the WAG 2 floodplain.

6.1.2 Soil Solution Exposures

Plants growing in seep areas are exposed to contaminants in the aqueous phase. Numerous seeps, springs, or intermittent tributaries in the WAG 2 floodplain were sampled for contaminants. It is assumed that the roots of plants near the springs may be exposed to water containing contaminants at similar concentrations. Although the discharge in many of these seeps is seasonal and heightened during rainfall, we make the conservative assumption that plants growing at these locations are exposed to contaminants in the water throughout the year. We also assume that the entire root systems of plants growing in seep areas are exposed to contaminants and that this type of exposure is the only significant source for the plants.

Filtered water samples from seeps are better representations than unfiltered samples of concentrations to which plants are exposed. Samples for this assessment were not filtered; thus the data are conservative estimates of exposure.

6.2 EFFECTS ASSESSMENT FOR THE PLANT COMMUNITY

Contaminant concentrations in soils and springs were compared to toxicological benchmarks (Will and Suter 1995b) in order to screen out chemicals that do not constitute a hazard to plants and therefore do not require detailed risk characterization. Tests conducted in natural soils in the laboratory were assumed to be representative of the exposure of plants to contaminants measured in floodplain soils, with the possibility that chemicals freshly added to soils in literature toxicity tests may be more bioavailable than contaminants in WAG 2 soils. Tests conducted in nutrient and mineral solutions are the best available representation of exposures of plants to contaminants in the vicinity of seeps and springs.

Benchmarks were not available for numerous organic contaminants, including polycyclic aromatic hydrocarbons.

6.3 RISK CHARACTERIZATION FOR THE PLANT COMMUNITY

The evidence concerning risks to the plant community consists of analyses of inorganic, organic and radionuclide contaminants in soil. This single line of evidence is summarized in the following text. Toxicity tests and biological surveys were not available.

6.3.1 Single Chemical Soil and Solution Toxicity

COPECs for terrestrial plants in the WAG 2 floodplain were identified by comparing maximum detected site concentrations of soil to the following: (1) twice the mean background for soils from WAG 5 (DOE 1995) and (2) available benchmarks for toxicity to terrestrial plants (Will and Suter 1995b). The screening of maximum concentrations against benchmarks is intended to protect plants that spend their entire lives in a single, highly contaminated location. If both background and benchmark criteria were exceeded, the contaminant was identified as a COPEC. The magnitude of exceedence of a benchmark is described by the hazard quotient, the maximum concentration in soil divided by the toxicity benchmark concentration. If background criteria were not available, exceedence of a benchmark was sufficient for the COPEC designation. Benchmarks for plants growing in soils, maximum concentrations of analytes at each reach exceeding background and for which benchmarks are available, and hazard quotients for the contaminants are given in Table B.7.

COPECs for plants potentially exposed to water from seeps and springs were identified by a similar comparison of contaminant concentrations with background and toxicity benchmarks (Table B.8).

6.3.1.1 Terrestrial plants

The screening of contaminant concentrations in soil against phytotoxicity benchmarks is provided in Table B.7.

Intermediate Holding Pond. The maximum detected concentrations of boron, chromium, lead, lithium, and manganese were 20, 70, 2, 20, and 8 times the phytotoxicity benchmarks (i.e., concentrations causing toxic effects when added to surface soil). Mercury, molybdenum, nickel, silver, and zinc were detected at 300, 3, 10, 8, and 3 times the benchmarks.

Lower White Oak Creek. The maximum detected concentrations of boron, chromium, lithium, manganese, mercury, molybdenum, silver, and zinc were 20, 100, 8, 3, 20, 2, 3, and 4 times the phytotoxicity benchmarks. Lead and selenium were detected at concentrations only slightly above benchmarks (hazard quotient of 1 if appropriate significant figures were used).

Middle White Oak Creek. The maximum detected concentrations of boron, lithium, mercury, molybdenum, selenium, silver, and zinc were 20, 7, 8, 2, 2, 2, and 2 times the phytotoxicity benchmarks.

Lower Melton Branch. The maximum detected concentrations of antimony, boron, lithium, manganese, mercury, and molybdenum were 600, 2, 20, 10, 20, 7, and 4 times benchmarks. Selenium and thallium were detected at concentrations only slightly above benchmarks (hazard quotient of 1 if appropriate significant figures were used).

The strength of the literature toxicity line of evidence is in its ability to relate the concentration of a single contaminant in soil to observed effects. The screening is most useful if the chemical species and bioavailability of contaminants in WAG 2 floodplain soils are comparable to those of the chemicals in laboratory toxicity tests in other soils. In these soils, the fractions of mercuric chloride, methyl mercury, mercuric sulfide, and others were not determined. Thus, the hazard quotients for mercury may be conservative.

6.3.1.2 Plants exposed to seeps and springs

The screening of contaminant concentrations in soil solution against phytotoxicity benchmarks is provided in Table B.8.

Embayment of White Oak Creek. The maximum detected concentration of aluminum at WOCET was 8 times the benchmark for plants in solution.

Homogeneous Reactor Test Tributary. The maximum detected concentration of aluminum in SW9-2 was 7 times the plant benchmark concentration for aluminum in solution.

Intermediate Holding Pond. No contaminants were detected above plant benchmarks in seeps or tributaries.

Lower Melton Branch. The maximum detected concentration of manganese at MID. DRAIN. was slightly above the benchmark for plants in solution. No contaminants were detected above plant benchmarks for the following seeps or tributaries: MBTRIB-3, SW2-5, SW5-2, and SW5-4.

Lower White Oak Creek. The maximum detected concentrations of aluminum from the East Seep, SW7-3, SW7-5, SW7-6, and WCTRIB-1 were 10, 4, 10, 4, and 5 times the plant solution benchmark. No other contaminants were detected above background and benchmarks in seeps in this region.

Middle White Oak Creek. No contaminants were detected above plant benchmarks in seeps or tributaries.

Upper Melton Branch. No contaminants were detected above plant benchmarks in seeps or tributaries.

Unknown reach. The maximum detected concentration of iron in SW2-7 was 2 times the plant solution benchmark.

WAG 4 Tributary. The maximum detected concentrations of arsenic and iron at SW4-1 were 6 and 3 times the plant solution benchmarks. The maximum detected concentration of nickel in SW4-2 was 50 times the plant solution benchmark. Aluminum was detected at 5 times the phytotoxicity benchmark in both WAG 4 MS1 and WAG 4 T2A. Chromium was detected at a concentration slightly above the plant solution benchmark in WAG4 T2A. No contaminants were detected above benchmarks in BTT.

West Seep. The maximum detected concentrations of aluminum and arsenic, chromium, and iron in RS-3A were 90, 20, 2, and slightly greater than 1 times the benchmark for toxicity to plants in solution. Aluminum was detected in RS-3B at up to 7 times the benchmark. Manganese in SW7-1 and aluminum and cobalt in SW7-2 were detected at 2, 3, and 2 times the phytotoxicity benchmark.

Aluminum and iron. Of the contaminants detected at seeps and springs at concentrations above benchmarks, aluminum and iron are unlikely to be of ecological concern. In unfiltered water, both of these contaminants are significantly associated with the particulate fraction, which is wholly or largely unavailable to plants. Also, aluminum and iron are typically found at high background concentrations (twice the mean background is 0.34 mg/kg for Al and 0.81 mg/kg for Fe) in this region.

6.3.2 Summary of Risk Characterization for the Plant Community

As in any screening assessment, potential hazards are indicated by the exceedence of toxicological benchmarks. COPECs for the plant community exposed to soils in the WAG 2 floodplain include: boron, chromium, lead, lithium, manganese, mercury, molybdenum, nickel, selenium, silver, thallium, and zinc. COPECs for the plant community exposed to seeps or springs in the WAG 2 floodplain include: arsenic, chromium, cobalt, manganese, and nickel. Further studies, such as toxicity tests (or other measures of bioavailability) and chemical speciation of mercury, would be advisable before concluding that these contaminants constitute hazards to the plant community.

6.4 UNCERTAINTIES CONCERNING RISKS TO PLANTS

The following factors create uncertainty in assessing the risk posed by the contaminants of potential concern in WAG 2 soils, seeps, and springs.

Bioavailability of elements in soil. The extraction methods used may remove from the soil quantities of chemicals greater than are available to plants. The double-acid extraction removes the exchangeable fraction of metals thereby giving a concentration that reflects the total potential pool of contaminants, not that to which the plant is exposed at any one time. Estimates of exposure are confounded by the concentration- and species-dependent synergistic and antagonistic interactions between metals during uptake by roots and inside plants.

Bioavailability of elements in groundwater and springs. Concentrations of contaminants such as aluminum in whole, unfiltered groundwater are probably higher than the concentrations

bioavailable to plants. In unfiltered water samples (i.e., containing soil and organic particles), acidification will bring into solution metals strongly bound to this particulate matter.

Variable response to toxicants. Various plant species and plant growth stages tolerate contaminants to different extents. The literature from which benchmarks were derived is not based on experiments using plants found in ecosystems representative of Bear Creek Valley. It is difficult to extrapolate from largely agricultural crops in early growth stages used in most of the published literature to trees and the varied ground vegetation found at WAG 2.

Multiple contaminant exposure. Because of our lack of understanding of the complex interactions between contaminants, benchmark levels are necessarily derived from experiments in which plants are exposed to single contaminants. Typical soils in WAG 2 expose plants to multiple contaminants which may not be adequately assessed on the basis of literature-derived benchmarks.

Lack of benchmarks for some metals and most organic compounds. Little research has been conducted on the phytotoxic effects of the organic compounds. Thus, it is not possible to assess the risk to plant growth posed by some of the analytes found in the Bear Creek Valley soils.

Chemical speciation. The analytical techniques did not generally differentiate among species of chemicals present in the soil which have variable toxicity to plants.

7. RISKS TO WILDLIFE

Risks from radionuclides and nonradionuclides (e.g., organic and inorganic chemicals) were estimated for small mammals, piscivores, and wide-ranging wildlife species within WAG 2. Because the methods for risk estimation differ greatly between radionuclide and nonradionuclide contaminants, each is addressed separately. Risks to wildlife from chemicals are presented in Sect. 7.1; risks from radionuclides to wildlife, terrestrial plants, and earthworms are presented in Sect. 7.2.

7.1 RISKS TO WILDLIFE FROM CHEMICALS

7.1.1 Exposure Assessment

Wildlife may be exposed to contaminants through ingestion of food, soil, and water. In this assessment, exposure through ingestion of food, soil, and water was estimated using exposure models (described in the following paragraphs. For areas where there was insufficient data to assess complete exposure, exposure through water was assessed by comparing unfiltered water concentrations to water benchmarks for wildlife (see Sect 7.1.2.1). Contaminant exposure through ingestion was estimated for small mammals (short-tailed shrew and white-footed mouse), wide-ranging species (white-tailed deer, wild turkey, red-tailed hawk, and red fox) and piscivores (mink and belted kingfisher). Exposure estimates were calculated using soil and soil-biota uptake factors for small mammals, earthworms, and herbaceous, canopy, and browse vegetation. Uptake factors for small mammals and herbaceous, canopy, and browse vegetation were derived from data from 15 locations within the Bear Creek watershed (four within BCOU1 and 11 within the Bear Creek floodplain). Uptake factors for earthworms were derived from data from Bear Creek, WAG 5 and two locations within WAG 2. Exposure to piscivores was assessed using fish data from eight locations in the WOC watershed.

7.1.1.1 Oral Ingestion Exposure Model

Oral exposure to contaminants experienced by wildlife may come from multiple sources. They may consume contaminated food (either plant or animal), drink contaminated water, or ingest soil or sediment. Soil or sediment ingestion may be incidental while foraging or grooming or purposeful to meet nutrient needs. The total oral exposure experienced by an individual is the sum of the exposures attributable to each source and may be described as follows:

$$E_{total} \approx E_{food} + E_{water} + E_{soil}, \quad (1)$$

where

- E_{total} = total exposure from all pathways,
- E_{food} = exposure from food consumption,
- E_{water} = exposure from water consumption,
- E_{soil} = exposure from soil consumption.

For exposure estimates to be useful in the assessment of risk to wildlife, they must be expressed in terms of a body weight-normalized daily dose or mg contaminant per kg body weight per day (mg/kg/d). Exposure estimates expressed in this manner may then be compared to toxicological

benchmarks for wildlife, such as those derived by Opresko et al. (1995), or to doses reported in the toxicological literature. Estimation of the daily contaminant dose an individual may receive from a

$$E_j = \sum_{i=1}^m P_{ik} \left(\frac{IR_i \times C_{ijk}}{BW} \right), \quad (2)$$

particular medium for a particular contaminant may be calculated using the following equation:
where

- E_j = total exposure to contaminant (j) (mg/kg/d),
- m = total number of ingested media (e.g., food, soil, or water),
- IR_i = ingestion rate for medium (I) (kg/d or L/d),
- P_{ik} = proportion of type (k) of medium (I) consumed (unitless),
- C_{ij} = concentration of contaminant (j) in type (k) of medium (I) (mg/kg or mg/L),
- BW = body weight of endpoint species (kg).

Exposure estimates were calculated for all contaminants detected in soil or water from WAG 2. Because wildlife are mobile, their exposure is best represented by the mean contaminant concentration in media. To be conservative, the 95% UCL is used in exposure estimates. These data were used in the initial exposure estimates. Exposure estimates for contaminants that may present a risk to wildlife [based upon comparisons to no observed adverse effects levels (NOAELs) and lowest observed adverse effects levels (LOAELs)] were recalculated using Monte Carlo simulations. (**Note:** because the purpose of the initial exposure estimate is to be conservative and to identify COPECs, the 95% UCL was used regardless of whether or not the value exceeded the maximum observed value. Overestimates of exposure that may occur at the screening level are addressed through the use of Monte Carlo simulation).

7.1.1.2 Piscivore exposures

Exposure of mink and belted kingfisher to mercury and PCBs in fish from WOC was estimated at eight locations within the watershed (and one reference location outside that watershed) as part of the *Preliminary Oak Ridge Reservation-wide Ecological Risk Assessment* (Sample et al. 1995). Estimates for mink and kingfisher are presented in Tables C.1 and C.2. Additional detail on these exposure estimates may be found in Sample et al. (1995).

7.1.1.3 Soil-biota uptake factors

Contaminant concentrations in biota were not available for WAG 2. To estimate contaminant concentrations in biota, we used uptake factors developed as part of the Bear Creek RI. Summary statistics for the biota uptake factors are presented in Table C.3. Uptake factors for small mammals, canopy, browse, and herbaceous vegetation were developed exclusively with data from the Bear Creek assessment. In addition to the Bear Creek data, data from two samples in WAG 2 and six from WAG 5 were used to develop the earthworm uptake factors. Contaminant concentrations in biota were estimated by multiplying the biota type-specific uptake factor by the soil concentration.

7.1.1.4 Contaminant concentrations in media

Contaminant concentrations in soil and water are needed to estimate exposure. Soil and water data were aggregated into four reaches: IHP, MWC, LWC, and LMB. Definitions of the bounds of these reaches may be found in Table 2.1.

Summary statistics) for contaminants detected in soil and water from WAG 2 are presented in Appendix C. For comparison, estimates of exposure to inorganic analytes were generated for all endpoints using the 2 × mean background soil and water concentrations obtained from the WAG 5 RI.

7.1.1.5 Exposure modeling using point estimates

Initial estimates of exposure of piscivorous wildlife to contaminants were performed for each of the four reaches using point estimates of parameters in the exposure model. Species-specific parameters necessary to estimate exposure using Eq. 1 are listed in Tables C.4 through C.11.

In estimating contaminant exposure experienced by short-tailed shrew, the following assumptions were made:

- body weight = 0.015 kg;
- food consumption = 0.009 kg/d (fresh weight);
- soil consumption = 0.00117 kg/d (dry weight);
- water consumption = 0.033 L/d;
- diet consists 100% of earthworms or soil invertebrates;
- contaminant concentration in earthworms is representative of that in other invertebrate prey.

To estimate contaminant exposure experienced by white-footed mouse, the following assumptions were made:

- body weight = 0.022 kg;
- food consumption = 0.0034 kg/d (fresh weight);
- soil consumption = 0.000068 kg/d (dry weight);
- water consumption = 0.0066 L/d;
- diet consists 50% of earthworms or soil invertebrates and 50% herbaceous plant material;
- contaminant concentration in earthworms is representative of that in other invertebrate prey.

To estimate contaminant exposure experienced by white-tailed deer, the following assumptions were made:

- body weight = 56.5 kg;
- food consumption = 1.74 kg/d (fresh weight);
- soil consumption = 0.0348 kg/d (dry weight);
- water consumption = 3.7 L/d;
- diet consists 33% of browse plants, 33% canopy plants, and 33% herbaceous plant material.

To estimate contaminant exposure experienced by red fox, the following assumptions were made:

- body weight = 4.5 kg;
- food consumption = 0.45 kg/d (fresh weight);

- soil consumption = 0.0126 kg/d (dry weight);
- water consumption = 0.38 L/d;
- diet consists 80.8% of small mammals and birds, 10.4% browse plant material, and 8.8% earthworms or other invertebrates;
- contaminant concentration in small mammals is representative of that in other vertebrate prey;
- contaminant concentration in earthworms is representative of that in other invertebrate prey.

To estimate contaminant exposure experienced by red-tailed hawk, the following assumptions were made:

- body weight = 1.126 kg;
- food consumption = 0.109 kg/d (fresh weight);
- soil consumption = 0 kg/d (dry weight);
- water consumption = 0.064 L/d;
- diet consists 100% of small mammals and other vertebrates;
- contaminant concentration in small mammals is representative of that in other vertebrate prey.

To estimate contaminant exposure experienced by mink, the following assumptions were made:

- body weight = 1 kg;
- food consumption = 0.137 kg/d (fresh weight);
- water consumption = 0.099 L/d;
- diet consists 54.6% of fish or other aquatic prey and 45.4% small mammals;
- contaminant concentration in fish is representative of that in other aquatic prey.

To estimate contaminant exposure experienced by kingfisher, the following assumptions were made:

- body weight = 0.148 kg;
- food consumption = 0.75 kg/d (fresh weight);
- water consumption = 0.016 L/d;
- diet consists 100% of fish.

To estimate contaminant exposure experienced by wild turkey, the following assumptions were made:

- body weight = 5.8 kg;
- food consumption = 0.174 kg/d (fresh weight);
- soil consumption = 0.0162 kg/d (dry weight);
- water consumption = 0.19 L/d;
- diet consists 90.3% of plant material and 9.7% invertebrates;
- contaminant concentration in earthworms is representative of that in other invertebrate prey.

Using Eq. 3 and the assumptions and data described above, we made point estimates of exposure to contaminants within each of the four WAG 2 reaches and the background for each endpoint (Tables C.12 through C.20).

7.1.1.6 Exposure modeling using Monte Carlo simulations

Employing point estimates for the input parameters in the exposure model does not take into account the variation and uncertainty associated with the parameters and therefore may over or under

estimate the contaminant exposure that endpoints may receive. In addition, calculating the model using point estimates produces a point estimate of exposure. This estimate provides no information concerning the distribution of exposures or the likelihood that individuals within the watershed will actually experience potentially hazardous exposures. To incorporate the variation in exposure parameters and to provide a better estimate of the potential exposure experienced by wildlife in WAG 2, the exposure model was re-calculated using Monte Carlo simulations.

Monte Carlo simulation is a resampling technique frequently used in uncertainty analysis in risk assessment (Hammonds et al. 1994). In practice, distributions are assigned to input parameters in a model; and the model is recalculated many times to produce a distribution of output parameters (e.g., estimates of contaminant exposure). Each time the model is recalculated, a value is selected from within the distribution assigned for each input parameter. As a result, a distribution of exposure estimates is produced that reflects the variability of the input parameters.

For all endpoints, Monte Carlo simulations were performed for the entire WAG 2 area. The percentiles of the resulting exposure distributions represent the likelihood that an individual within the modeled area will experience a given exposure level. It was assumed that each sampling location contributed equally to the overall mean exposure (i.e., individuals within each watershed do not preferentially forage at any one location within the watershed). While this assumption is not likely to be ecologically correct (foraging effort and therefore exposure is likely to be biased toward those locations with the most abundant food), data were not available to estimate the preferential use at each sampling location.

Simulations were performed for each contaminant where comparison of point estimates of exposure to LOAELs produced HQs ≥ 1 for at least one location (LOAELs are presented in Sect 7.1.2.; Screening of exposure estimates against LOAELs is presented in Sect. 7.1.3.).

Distributions were used for the following parameters in the exposure model: contaminant concentrations in soil and water and soil-biota uptake factors. All distributions were assumed to be normal. Because these wildlife are mobile, the contaminant concentration they are exposed to on a daily basis is best represented by the average concentration instead of the entire distribution. The standard error of the mean was used to describe variation in the average contaminant concentration.

Monte Carlo simulations were performed using the @Risk software. Samples from each distribution were selected using latin hypercube sampling. The number of iterations, or recalculations, of each exposure simulation was determined by the convergence criteria set in the software. Under these criteria, iterations are performed until the between-iteration percent change in the percentiles, mean, and standard deviation is below 1.5% (i.e., the percentile, mean, and standard deviation for the latest iteration is less than 1.5% different than the those from the previous iteration). Using this convergence criterion, from 600 to 2500 model iterations were performed for each exposure estimate. Monte Carlo estimates of contaminant exposures are presented in Table C.19.

7.1.2 Chemical Effects Assessment for Wildlife

7.1.2.1 Single chemical toxicity data

Single chemical toxicity data consist of no observed adverse effects levels (NOAELs) and lowest observed adverse effects levels (LOAELs) of toxicity studies reported in the literature. NOAELs and LOAELs for wildlife endpoints were estimated from these data using the allometric methods outline in Opresko et al. (1995). This methodology for toxicity extrapolation is equivalent to that the EPA

uses in their carcinogenicity assessments and Reportable Quantity documents for adjusting from animal data to an equivalent human dose. Using the allometric scaling factor recommended in EPA (1995), the equation for estimating mammalian NOAELs and LOAELs was:

$$NOAEL_w = NOAEL_t \left(\frac{bw_t}{bw_w} \right)^{1/4} \quad (3)$$

where $NOAEL_t$ and $NOAEL_w$ represent NOAELs for a mammalian test species and wildlife species, respectively. Toxicity values for birds were estimated using the scaling factor derived from Mineau (1996) where:

$$NOAEL_w = NOAEL_t \left(\frac{bw_t}{bw_w} \right)^0 = NOAEL_t (1) = NOAEL_t \quad (4)$$

To evaluate the potential risk that contaminants in water may present, water benchmarks were derived according to the methods outline in Opresko et al. (1995). NOAEL's and LOAEL's and water benchmarks were derived for all seven endpoints. Experimental information used to estimate mammalian benchmarks and NOAEL's and LOAEL's for mammalian endpoints are presented in Tables C.20 and C.21. NOAELs and LOAELs for avian endpoints are listed in Table C.22. Water benchmark values for all endpoints are listed in Table C.23.

7.1.2.2 Biological surveys

Mink Survey. Stevens (1995) investigated bioaccumulation of mercury in mink on the ORR in 1993 through 1995. The methods used in the mink survey, while indicating that mink are present on the Reservation, cannot be used to estimate abundance or density on mink on the ORR. A total of four male mink were live-trapped over the course of 6073 trapnights (trapnight=one trap set for 24 h). One juvenile was captured along East Fork Poplar Creek, two adults were captured along Bear Creek, and one adult was captured along WOC. Captured mink were fitted with an intraperitoneal radio transmitter (to monitor movements and home range) and were released. Prior to release, samples of hair were collected for metals analysis. An additional eight roadkill mink (five male and three female) were collected from the ORR and surrounding areas of Roane and Anderson counties. While one roadkill sample (a male) was collected on a bridge over Bear Creek and was assumed to be a resident of Bear Creek, all others were collected off the ORR and were used as references. Results of metals analysis are presented in Table 7.1.

Radiotelemetry data on home ranges and movements were obtained for 3 mink—one each from the East Fork Poplar Creek, Bear Creek, and WOC watersheds. Mean (\pm standard deviation) home range for these three individuals was found to be 7.5 ± 3 km of stream. The entire home range of the East Fork Poplar Creek mink was in a highly urbanized area; it included all of upper East Fork inside the Oak Ridge Y-12 Plant and all areas of East Fork upstream of the Oak Ridge Turnpike–Illinois Avenue intersection. The home range of the WOC mink included all of WOC from the headwater tributaries to the Clinch River, including the X-10 facility. This individual was observed to use dens within the X-10 facility and moved through the facility on several occasions.

Table 7.1. Metal concentrations in hair of Mink from the Oak Ridge Reservation and from off-site reference samples^a

Site	N	Hg	Se	As	Cd	Pb
East Fork Poplar Creek	1	104	0.69	ND ^b	ND	0.33
Bear Creek	3	10.97±3.42	1.88±1.41	0.15±0.09	0.04±0.02	0.97±1.28
White Oak Creek	1	8.8	1	ND	ND	0.37
Offsite	7	5.15±3.43	1.11±0.25	0.22±0.31	0.04±0.02	0.7±0.31

^aMean ± standard deviation mg/kg dry weight

^bND = Not Detected

Belted Kingfisher Survey. A field monitoring effort (Baron and Ashwood 1996) was initiated in 1994 to evaluate population parameters and contaminant bioaccumulation by belted kingfisher on the ORR. Areas surveyed included: WOC, WOL, WOL embayment, Melton Branch, Poplar Creek, portions of East Fork Poplar Creek (within the Y-12 Plant to downstream of Lake Reality and approximately 1 mile east of the confluence of Poplar Creek), and portions of Bear Creek.

Methods. Nest burrows were monitored for nesting activity. If activity were observed, samples of feathers and eggshells were collected. In addition to specimens collected from the burrows, three carcasses of adult kingfishers were found on the ORR (two from East Fork Poplar Creek and one from WOC). These carcasses were necropsied, and organs were extracted and analyzed for metals and radionuclides. Additional detail concerning methods are reported in Baron and Ashwood (1996).

Results. During April-July of 1994, a total of 27 potential kingfisher burrows were identified on the ORR, 11 of which contained swallow nests. Twenty-five of these burrows were found on the Clinch River. One kingfisher burrow, containing a single unhatched kingfisher egg, was found on WOC (downstream of WOC km 3.5).

One active burrow, containing a clutch of six to seven eggs, was found on the Clinch River. This burrow was later abandoned with no sign of the eggs or the parents. Another burrow, containing six nestlings was located on the Clinch River approximately 12 miles upstream of all DOE contaminant outfalls. It was, therefore, considered uncontaminated. Three weeks following the initial observation of this burrow, three nestlings had fledged and three had died. Feathers were collected and analyzed. Results of residue analysis for eggshells and feathers from nestlings are presented in Table C.24; results for adult carcasses are presented in Table C.25.

Nestling feathers collected from the burrow on the Clinch River, upstream of ORR outfalls (Table C.24), contained relatively low levels of metal and radioactive contaminants. Feathers from the carcasses of three fledglings accumulated similar concentrations of As, Cd, Pb, Se, and Hg. Mercury concentrations in feathers were approximately 1 mg/kg. Mercury concentrations found in fish downstream of the nesting site are approximately 0.04 ± 0.01 mg/kg (Peterson et al. 1994). Thus, biomagnification is occurring in kingfishers foraging in up-gradient areas of the ORR. However, these feather concentrations are much lower than those found in adult kingfishers on the ORR. While selenium concentrations in nestling feathers appear high, they are similar to selenium levels in adult kingfishers (Table C.25) and mink and raccoons collected at reference locations (Ashwood et al. 1994). The fourth feather sample presented in Table C.24 (CRU) represents a mixture of feathers

retrieved from the three nestlings. This sample was analyzed to provide additional information on the variability of chemical concentrations within the feathers.

A burrow on the Clinch River contained fragments of egg shells and fish vertebrae from regurgitant. Analysis of the egg shells indicated that minimal metal contamination was present (CRD, Table C.24). Another burrow on WOC contained an unhatched kingfisher egg (WOC, Table C.24). Metal concentrations in this egg were similar to that for the Clinch River egg, except for cesium-137. The presence of this radionuclide in the egg indicates that the parent kingfisher bioaccumulated cesium-137 from foraging within WOC or a nearby surface impoundment (cesium-137 is a typical contaminant of this stream and the impoundments).

In at least one kingfisher from the ORR, ¹³⁷Cs, Cd, Pb, Se, and Hg were each detected (Table C.25). Arsenic was analyzed for but was not detected. Feathers of adult kingfishers contained elevated mercury (Table C.25) relative to feathers from the nestlings (Table C.24). The greatest burdens of Hg, Se, Pb, and Cs-137 were observed in the bird from the WOC watershed (bird 3; Table C.25). In contrast, cadmium levels were higher in the birds from East Fork Poplar Creek (birds 1 and 2) than in the WOC bird (Table C.25).

7.1.3 Characterization Chemical Risks to Wildlife

Risk Characterization integrates the results of the exposure assessment (Sect. C.1.1) and effects assessment (Sect. C.1.2) to estimate risks (the likelihood of effects given the exposure) based on each line of evidence and then applies a weight of evidence inference logic to determine the best estimate of risk to each assessment endpoint. In an ideal risk assessment, there are three lines of evidence: literature-derived single chemical toxicity data (which indicate the toxic effects of the concentrations measured in site media); biological surveys of the affected system (these indicate the actual state of the receiving environment); and toxicity tests with ambient media (these indicate the toxic effects of the concentrations measured in site media). With the exception of the biosurvey data for mink and belted kingfisher (Sect 7.1.2.2), only one line of evidence was available to assess risk to wildlife in WAG 2— that is single chemical toxicity data.

7.1.3.1 Single chemical toxicity data

Exposure estimates generated by the exposure model (see Sect. 7.1.1) produced by both point estimates of parameter values and Monte Carlo simulation represent exposure at the individual level. The exposure estimates using point estimates of parameter values at each reach are used to identify COPECs and locations that contribute significantly to risk. In contrast, the WAG 2-wide exposure distributions generated by Monte Carlo simulation represent the likelihood that an individual within the WAG will experience a particular exposure.

Two types of single chemical toxicity data are available with which to evaluate wildlife contaminant exposure: NOAELs and LOAELs. NOAELs are used to screen exposure estimates generated from point estimates of exposure parameters; if the estimate is greater than the NOAEL, adverse effects are possible and additional evaluation is necessary. LOAELs are compared to the exposure distribution generated by the Monte Carlo simulation. If the LOAEL is lower than the 80th percentile of the exposure distribution, there is a >20% likelihood that individuals within the modeled location are experiencing contaminant exposures that are likely to produce adverse effects. By combining measured or literature-derived population density data with the likelihood or probability of exceeding the LOAEL, the magnitude of population-level impacts may be estimated.

Screening Point Estimates of Exposure. To determine if the contaminant exposures experienced by wildlife in each reach and throughout WAG 2 are potentially hazardous, the dietary contaminant exposure estimates (generated using point estimates of parameter values; see Tables C.1 and C.2, and C.12 through C.18) were compared to estimated NOAELs and LOAELs for these species (Tables C.21 and C.22). To quantify the magnitude of hazard, an HQ was calculated, where $HQ = \text{exposure}/\text{NOAEL}$ or LOAEL . HQs greater than 1 indicate that individuals may be experiencing exposures that are in excess of NOAELs or LOAELs. While exceeding the NOAEL suggests that adverse effects are possible, exceeding the LOAEL suggests that adverse effects are likely. HQs for all endpoints are presented along with the point estimates of exposure in Tables C.1 and C.2, and C.12 through C.18

While exposure of shrews (Table C.12), mice (Table C.13), deer (Table C.14), fox (Table C.15), and turkey (Table C.17) exceeded both NOAELs and LOAELs for aluminum in all four reaches, all WAG 2 estimates for each endpoint were less than that at the background. Because exposures to aluminum were less than background and background exposure is assumed to be nonhazardous, aluminum was dropped as a contaminant of concern for wildlife.

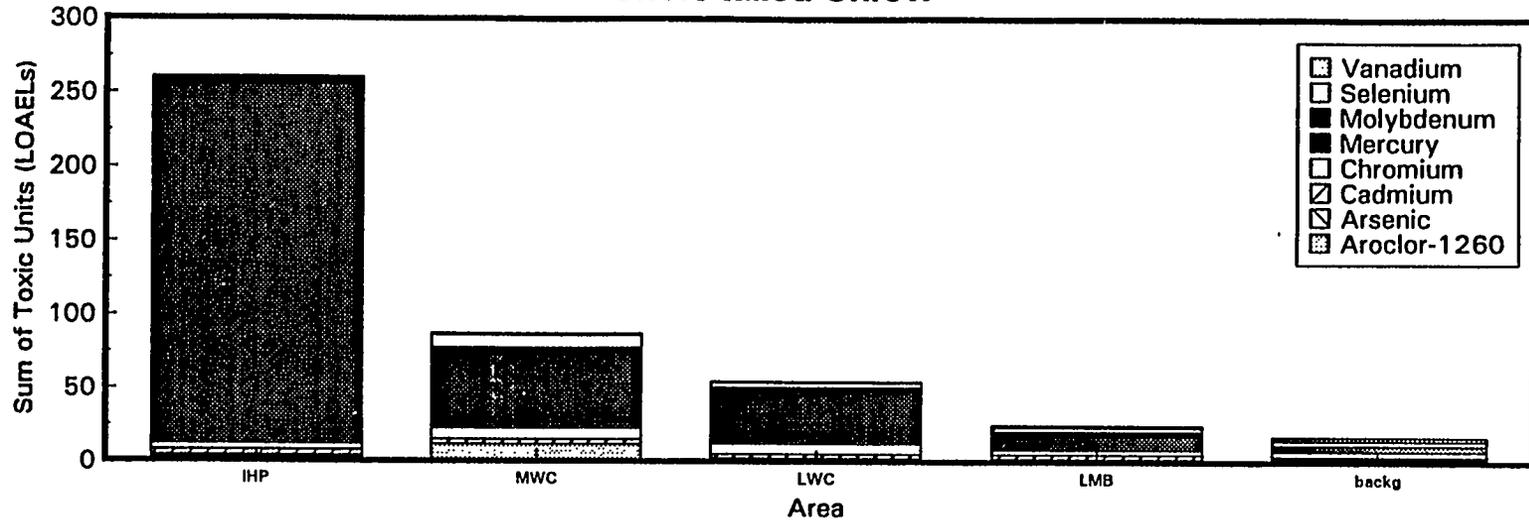
Mercury was the dominant contaminant presenting risks to wildlife in WAG 2. Mercury exposure in WAG 2 exceeded NOAELs, LOAELs, and background for all endpoints within all reaches (except for deer, where LOAELs were exceeded only within the IHP area, and turkey, where LOAELs were not exceeded in the LMB area). The next most important contaminants presenting risks were PCBs and selenium, which presented risks to shrews (Table C.12), mice (Table C.13), and fox (Table C.15). In general, few other contaminants presented significant risks. With the exception of short-tailed shrews, only one to at most four contaminants were identified as presenting risks to any endpoint. In the case of shrews, seven contaminants contributed significant risks (Table C.12).

The spatial distribution of contamination and potential risks to wildlife in WAG 2 are illustrated in Figs. 7.1 through 7.9. These figures display the sum of the LOAEL-based HQs (e.g., sum of toxic units or Σ TUs) for those contaminants where at least one LOAEL-based $HQ > 1$ was obtained. For all endpoint species, the greatest risks were identified in the mainstem of WOC. The highest Σ TUs are in the IHP area. Σ TUs decline with increasing distance downstream (IHP > MWC > LWC). Risks in the LMB were consistently lower than that in all three WOC reaches. The spatial patterns of contaminant risk were comparable for all endpoint species.

Screening Monte Carlo Simulation Estimates of Exposure. To incorporate the variation present in the parameters employed in the exposure model, Monte Carlo simulations were performed for the exposure estimates of each species to analytes where at least one LOAEL-based $HQ > 1$ was observed. For all endpoints, simulations were performed only at the WAG 2-wide level. The mean, standard deviation, and 80th percentile of the simulated exposures are presented in Table C.19.

By superimposing NOAEL and LOAEL values on the exposure distributions generated from the Monte Carlo simulation, the likelihood of an individual experiencing potentially hazardous exposures can be estimated, and the magnitude of risk may be determined. Interpretation of the comparison of exposure distributions to NOAELs and LOAELs is described in Table 7.2.

Short-tailed Shrew



Without Mercury

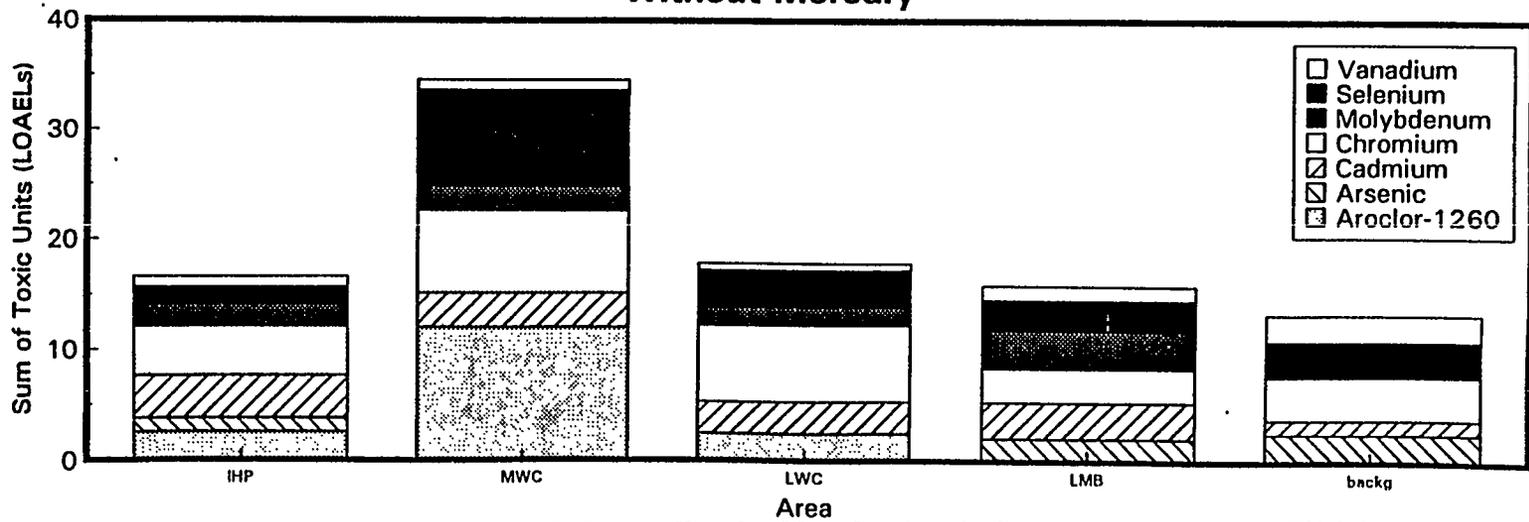


Fig. 7.1. Summary of lowest observed adverse effects level-based toxic units for short-tailed shrews in WAG 2.

White-footed Mouse

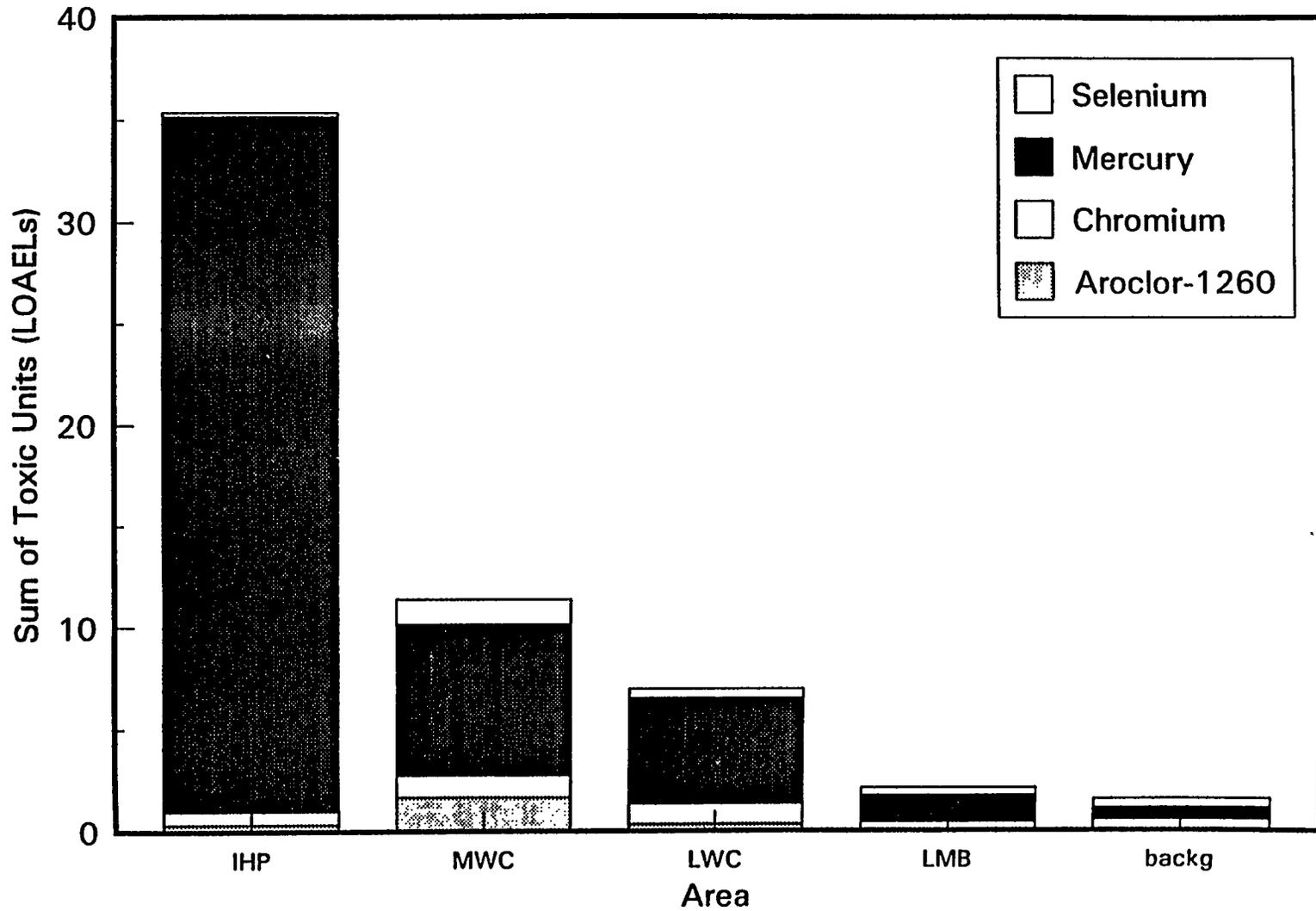


Fig. 7.2. Summary of lowest observed adverse effects level-based toxic units for white-footed mice in WAG 2.

Red Fox

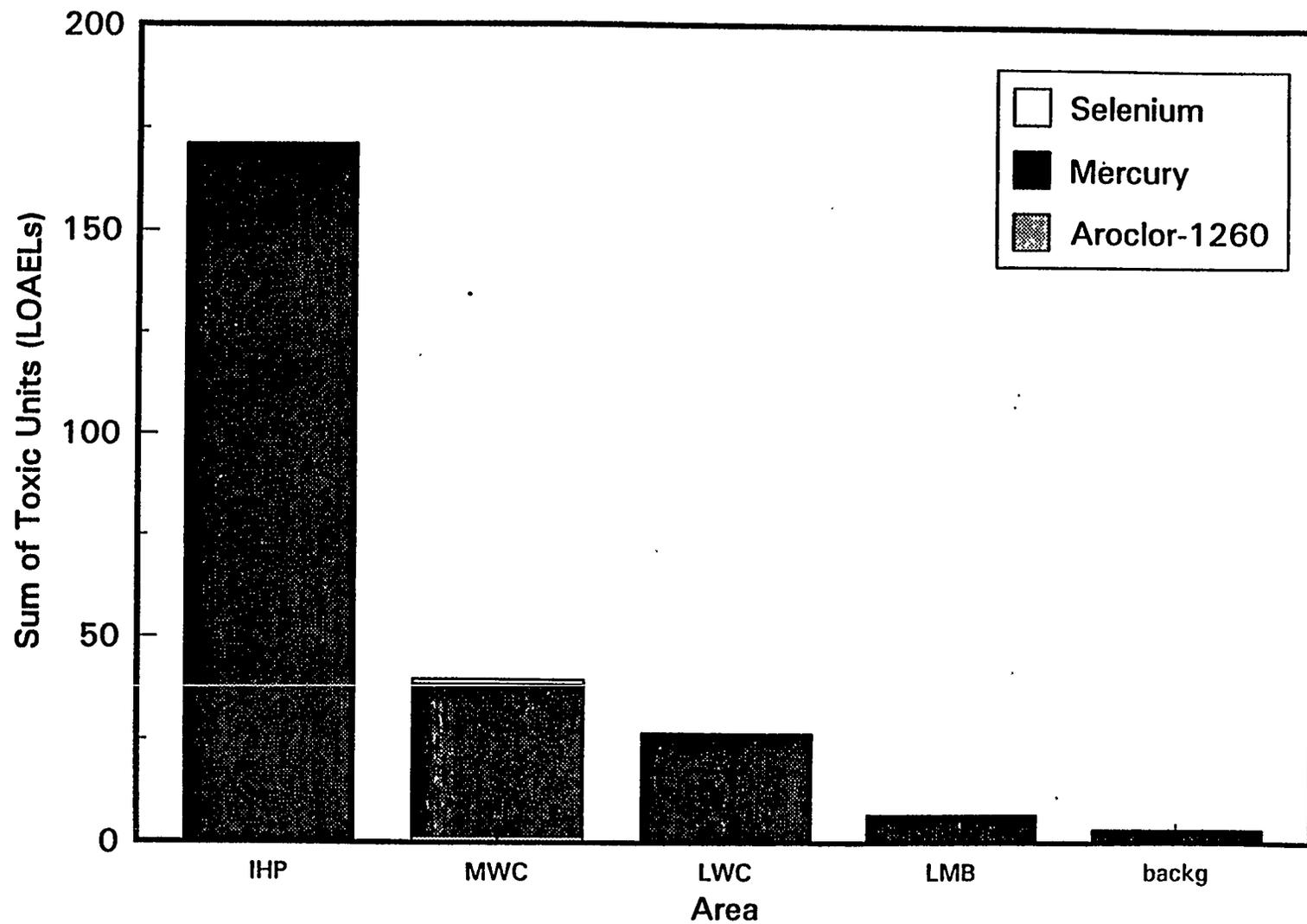


Fig. 7.3. Summary of lowest observed adverse effects level-based toxic units for red fox in WAG 2.

Mink

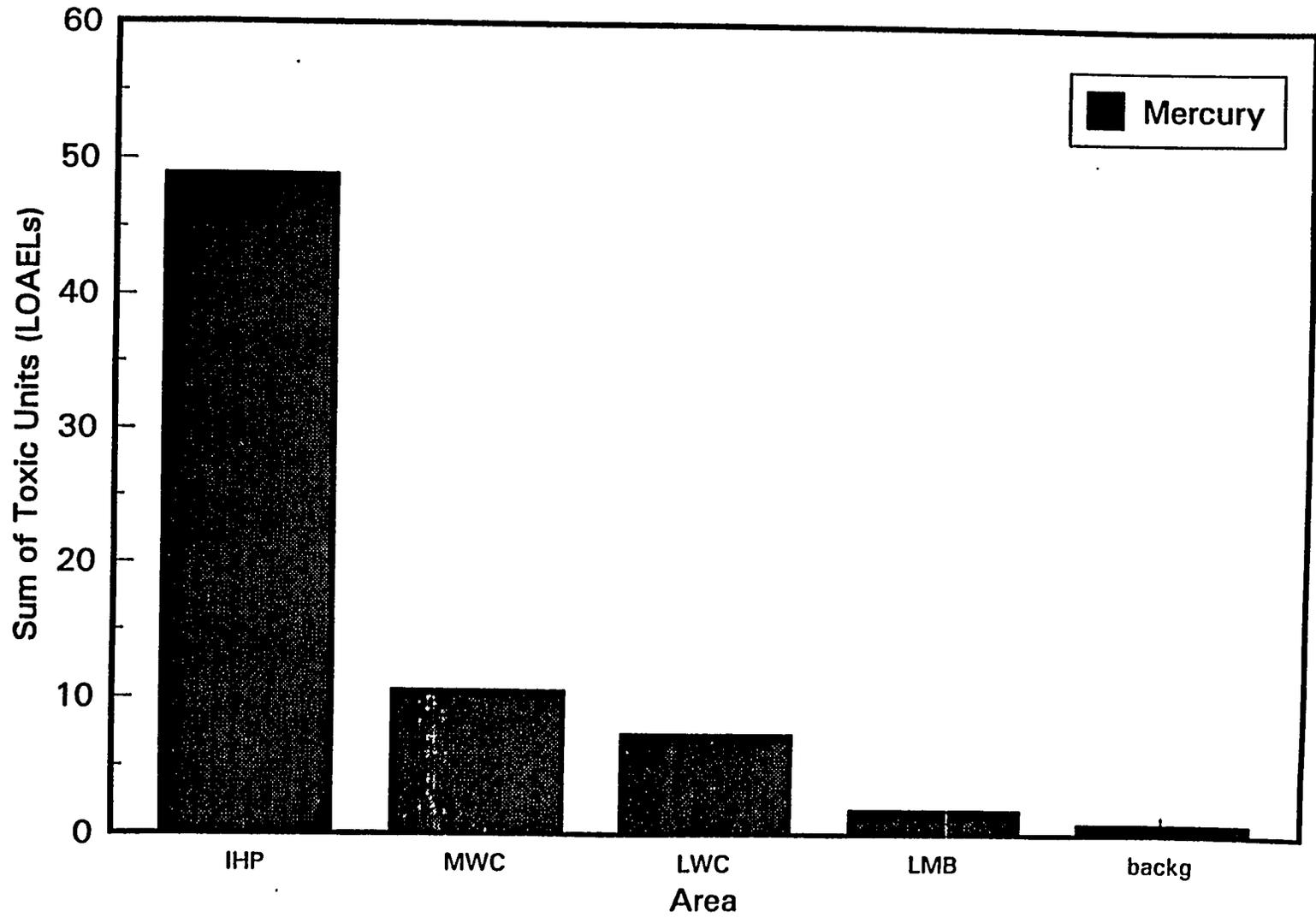


Fig. 7.4 Summary of lowest observed adverse effects level-based toxic units for mink in WAG 2.

Mink

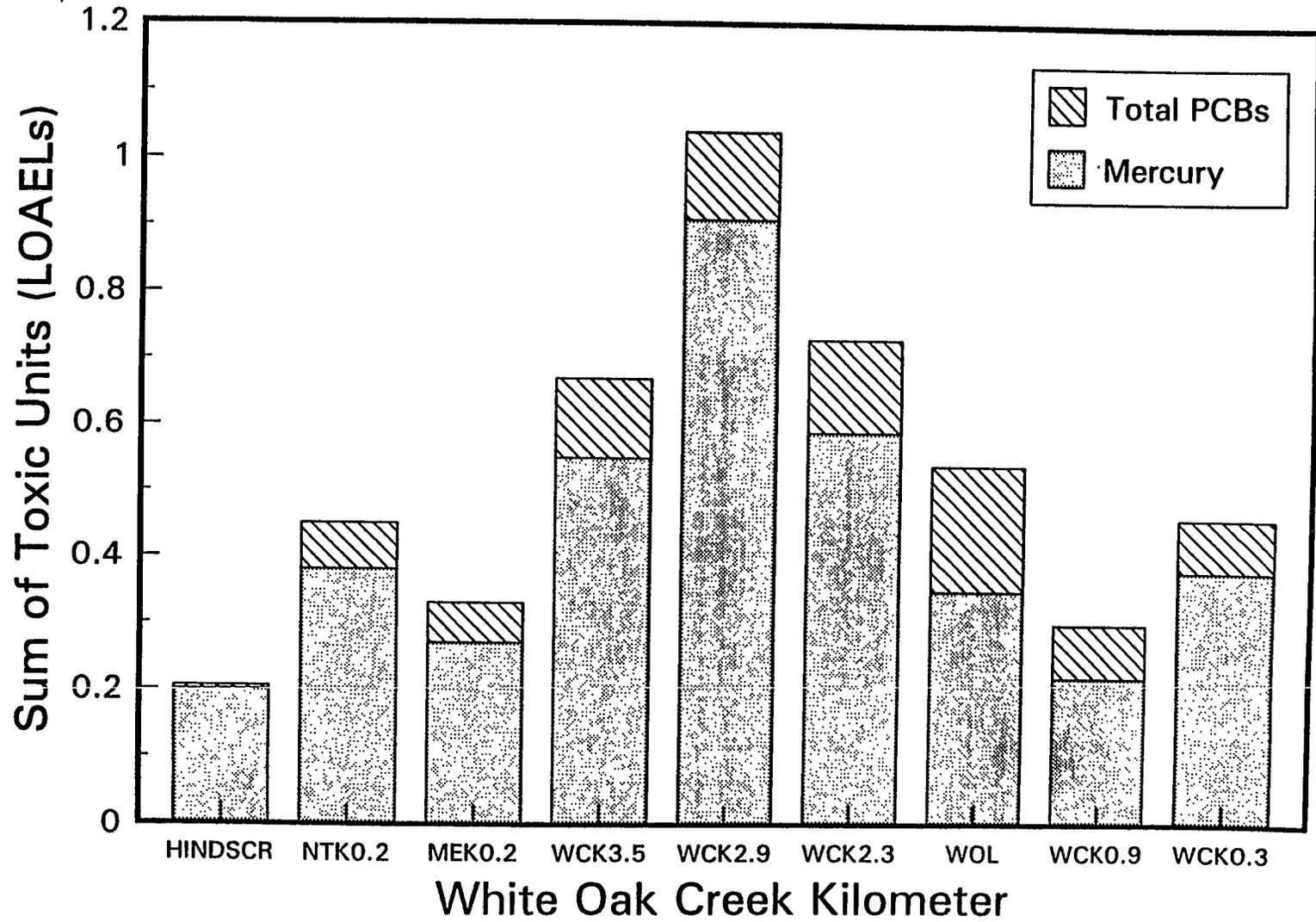


Fig. 7.5. Summary of lowest observed adverse effects level-based toxic units for mink in WAG 2 (exposure through ingestion of fish only).

White-tailed Deer

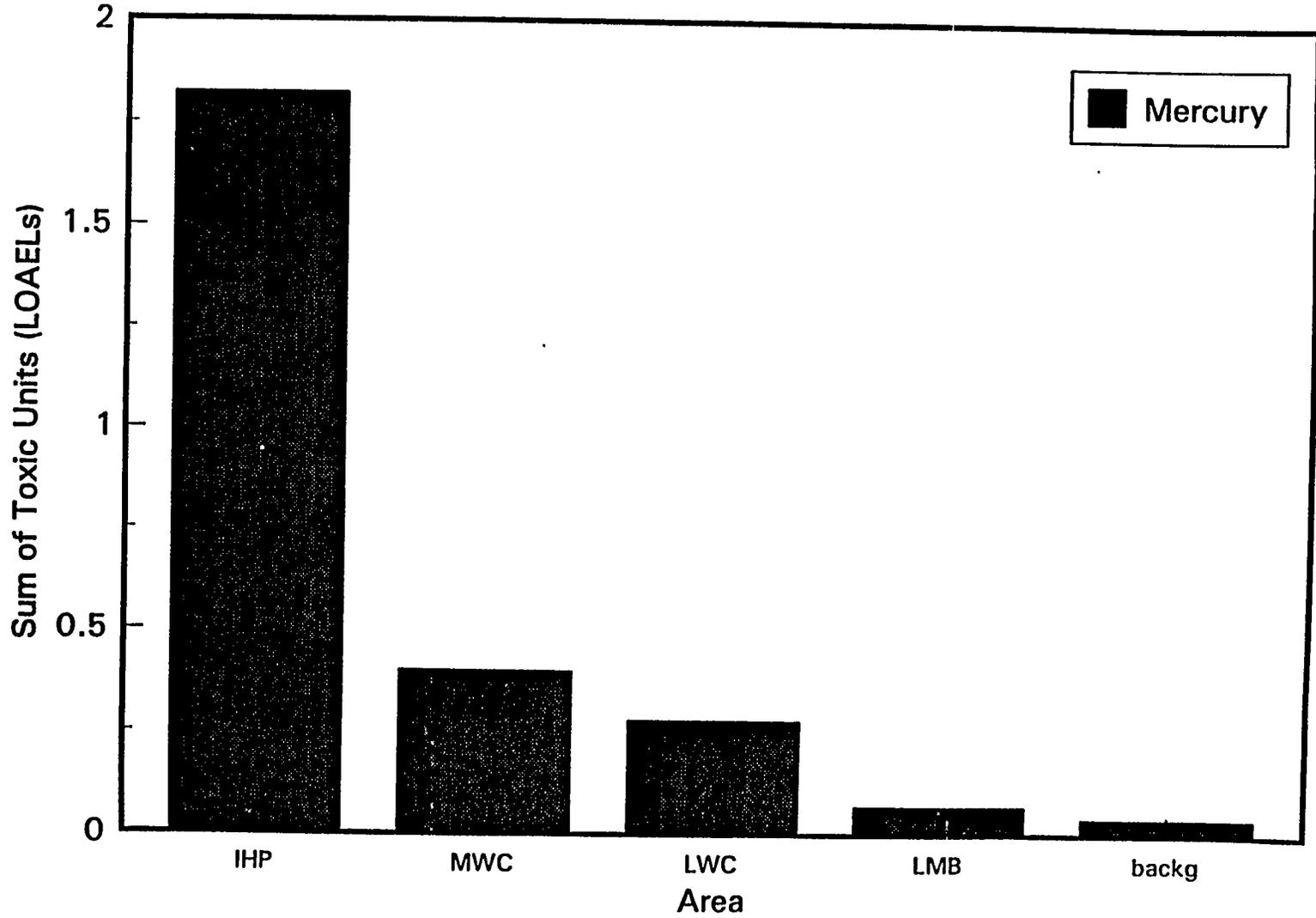


Fig. 7.6. Summary of lowest observed adverse effects level-based toxic units for white-tailed deer in WAG 2.

Wild Turkey

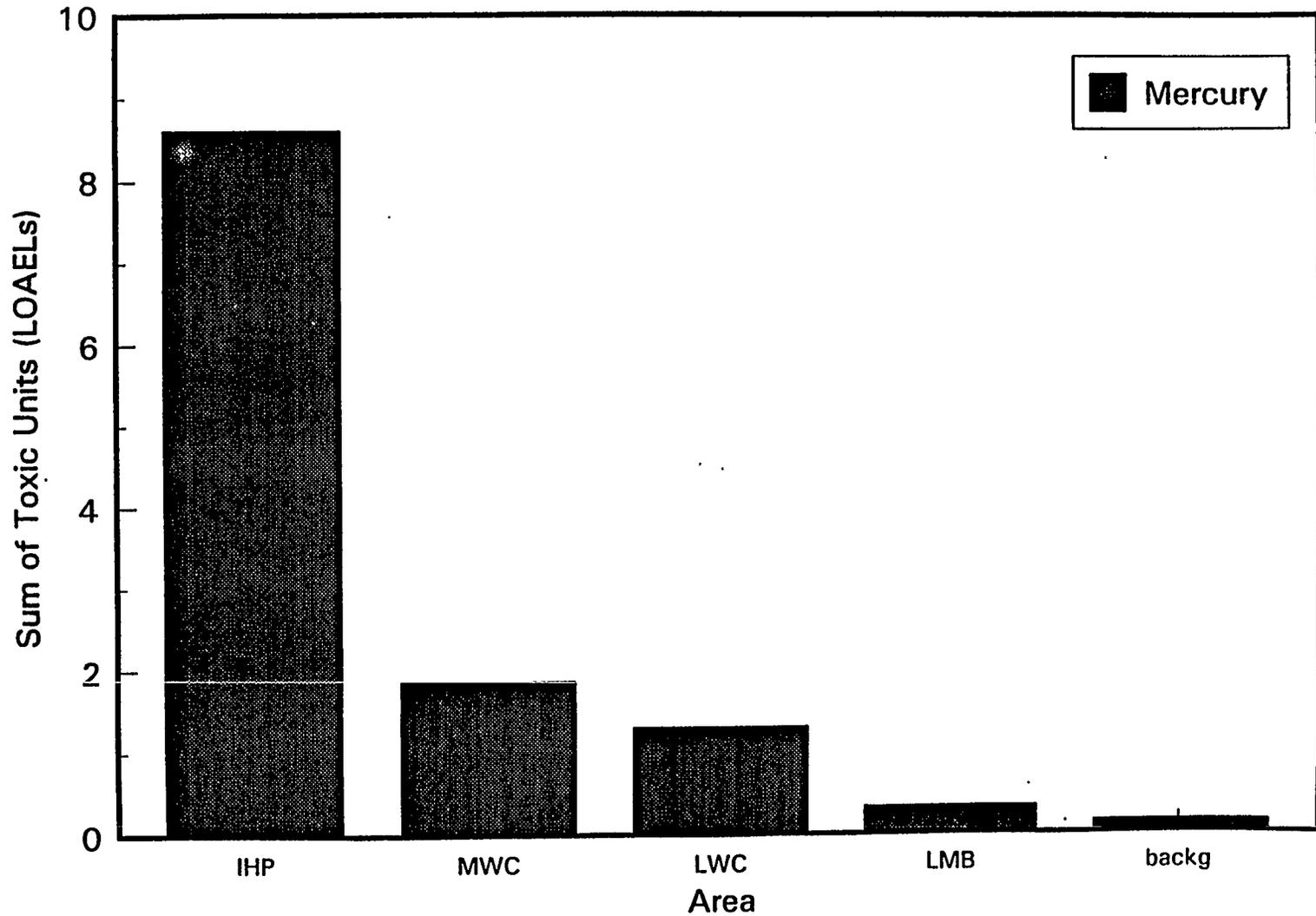


Fig. 7.7. Summary of lowest observed adverse effects level-based toxic units for wild turkey in WAG 2.

Red-Tailed Hawk

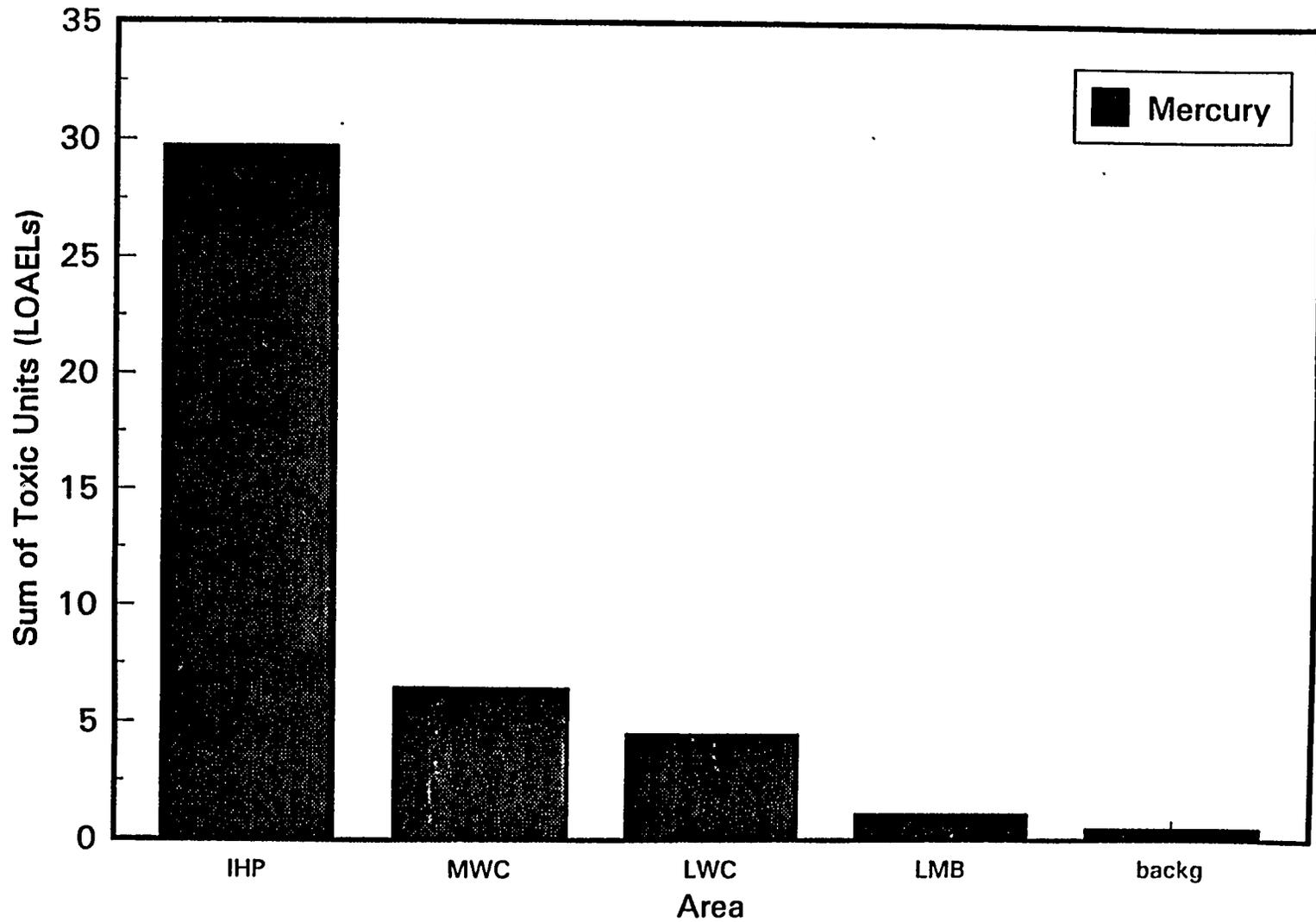


Fig. 7.8. Summary of lowest observed adverse effects level-based toxic units for red-tailed hawks in WAG 2.

Belted Kingfisher

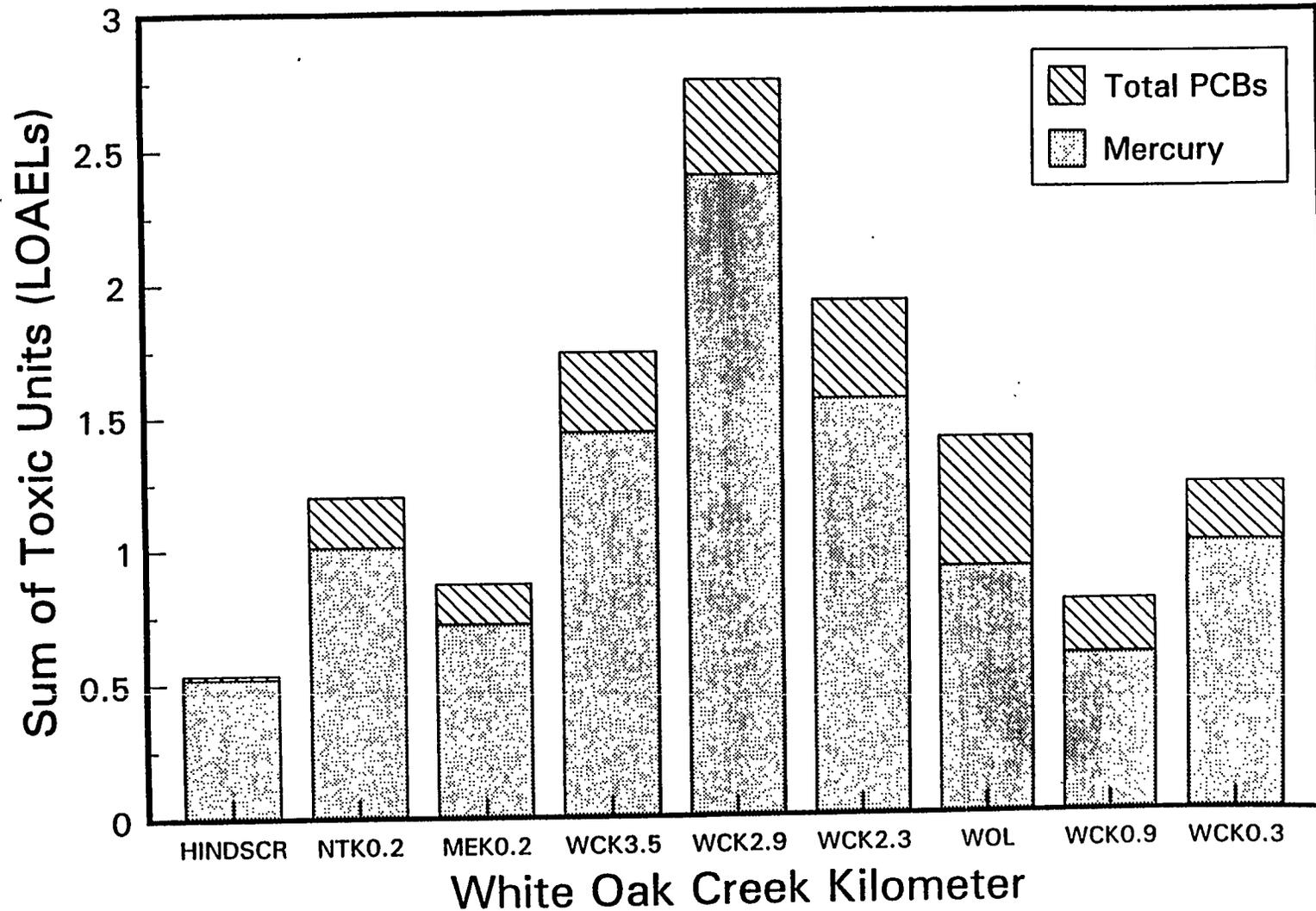


Fig. 7.9. Summary of lowest observed adverse effects level-based toxic units for belted kingfisher in WAG 2 (exposure through ingestion of fish only).

Table 7.2. Comparison of exposure distributions

Comparison	Meaning	Risk-based Interpretation
NOAEL > 80th percentile of exposure distribution	less than 20% of exposures > NOAEL	Individual- and population-level adverse effects are highly unlikely
NOAEL < 80th percentile < LOAEL	More than 20% of exposures > NOAEL, but less than 20% of exposures > LOAEL	Individuals experiencing exposures at the high end of the distribution may experience adverse effects, but those effects are unlikely to significantly contribute to effects on the Oak Ridge Reservation (ORR) population.
LOAEL < 80th percentile of exposure distribution	More than 20% of exposures > LOAEL	Effects on some individuals are likely and they may contribute significantly to effects on the ORR population.

Note: NOAEL=no observed adverse effects level; LOAEL=lowest observed adverse effects level

To evaluate the likelihood and magnitude of population-level effects on wildlife, literature-derived population density data (expressed as number of individuals/ ha or km of stream) were combined with hectares of suitable habitat (within WAG 2) to estimate the number of individuals of each endpoint species expected to be present in the watershed. For the terrestrial species (shrew, mice, deer, fox, turkey and hawk), habitat preferences follow those reported in the *Preliminary Reservation-wide Ecological Risk Assessment* (Sample et al. 1995). These habitat preferences were compared to the habitat types identified in WAG 2 in Washington-Allen et al. (1995). Because streams are the preferred habitats for mink and kingfisher, the length of White Oak Creek and its tributaries was assumed to represent suitable habitat. The estimated abundance of wildlife endpoint species is reported in Table C.26.

The number of individuals within WAG 2 likely to experience exposures >LOAELs can be estimated using cumulative binomial probability functions (Dowdy and Wearden 1983). Binomial probability functions are estimated using the following equation:

$$b(y; n; p) = \binom{n}{y} p^y (1-p)^{n-y}, \quad (5)$$

where

- y = the number (or percent) of individuals experiencing exposures >LOAEL,
- n = total number (or percent) of individuals within the watershed,
- p = probability of experiencing an exposure in excess of the LOAEL,
- b(y; n; p) = probability of y individuals out of a total of n, experiencing an exposure >LOAEL, given the probability of exceeding the LOAEL=p.

Solving Eq. 8 for y=0 to y=n generates a cumulative binomial probability distribution, which can be used to estimate the number of individuals within WAG 2 who are likely to experience adverse effects.

Binomial probability distributions were generated only for contaminant-endpoint combinations where the percent of the exposure distribution exceeding the LOAEL was 20% to 80% (these values

are reported in Table C.19). If the percent of the exposure distribution exceeding the LOAEL was <20%, it was assumed that no individuals within the area of interest were experiencing adverse effects. Conversely, if the percent of the exposure distribution exceeding the LOAEL was >80%, it was assumed that all individuals within the area of interest were experiencing adverse effects. Exposure estimates for six contaminant-endpoint combinations met the 20% to 80% exceedance criterion: PCB, cadmium, and selenium exposure to shrews and mercury exposure to fox, kingfisher, and turkey. Figures 7.10 through 7.15 graphically display the cumulative binomial probability distributions for each contaminant-endpoint combination. The total numbers of individuals for each endpoint species estimated to be experiencing adverse effects within WAG 2 are summarized in Table C.27.

Based on the Monte Carlo analysis, comparison to background exposure estimates, and binomial distribution analysis, the following conclusions may be made:

- Because <20% of the WAG 2 populations are estimated to be experiencing exposures >LOAEL, the following contaminants do not present significant risks:

Endpoint	Analytes
Short-tailed shrew	As, Ba, Cu, Mn, Mo, Ni, V, and Zn
White-footed mouse	Aroclor-1260, Cr, and Se
Red fox	Aroclor-1260 and Se
Mink	Hg (exposure through fish consumption only)
White-tailed deer	Hg

- Because >20% of the WAG 2 shrew population is estimated to be experiencing exposures >LOAEL, Aroclor-1260, cadmium, chromium, mercury, and selenium present a significant risk to shrews in WAG 2;
- Because >20% of the populations are estimated to be experiencing exposures >LOAEL, mercury presents a significant risk to white-footed mice, red fox, mink, belted kingfisher, red-tailed hawk, and wild turkey in WAG 2.

Screening Point Estimates of Exposure: Surface Water. To evaluate the potential risk that contaminants in surface water present to wildlife in reaches and seeps where total exposure was not estimated (due to lack of surface soil data), the 95% UCLs for concentrations in unfiltered water were compared to NOAEL and LOAEL water benchmarks for all species. HQs (water concentration/benchmark value) were calculated for all species. Comparisons of reach concentrations to NOAEL benchmarks are presented in Table C.28. NOAEL-based screening benchmarks were not exceeded within any reach, indicating that exposure to surface water within these WOC watershed reaches does not contribute to risk. These screening results indicated that exposure to seep water within the WOC watershed does not contribute to risk.

Comparisons of seep concentrations to NOAEL and LOAEL benchmarks are presented in Tables C.29 and C.30. Only one contaminant (aluminum) at one seep (RS-3A) exceeded NOAEL benchmarks (Table C.29); the LOAEL benchmarks, however were not exceeded (Table C.30). At all other seeps for all other analytes, NOAEL-based benchmarks were not exceeded.

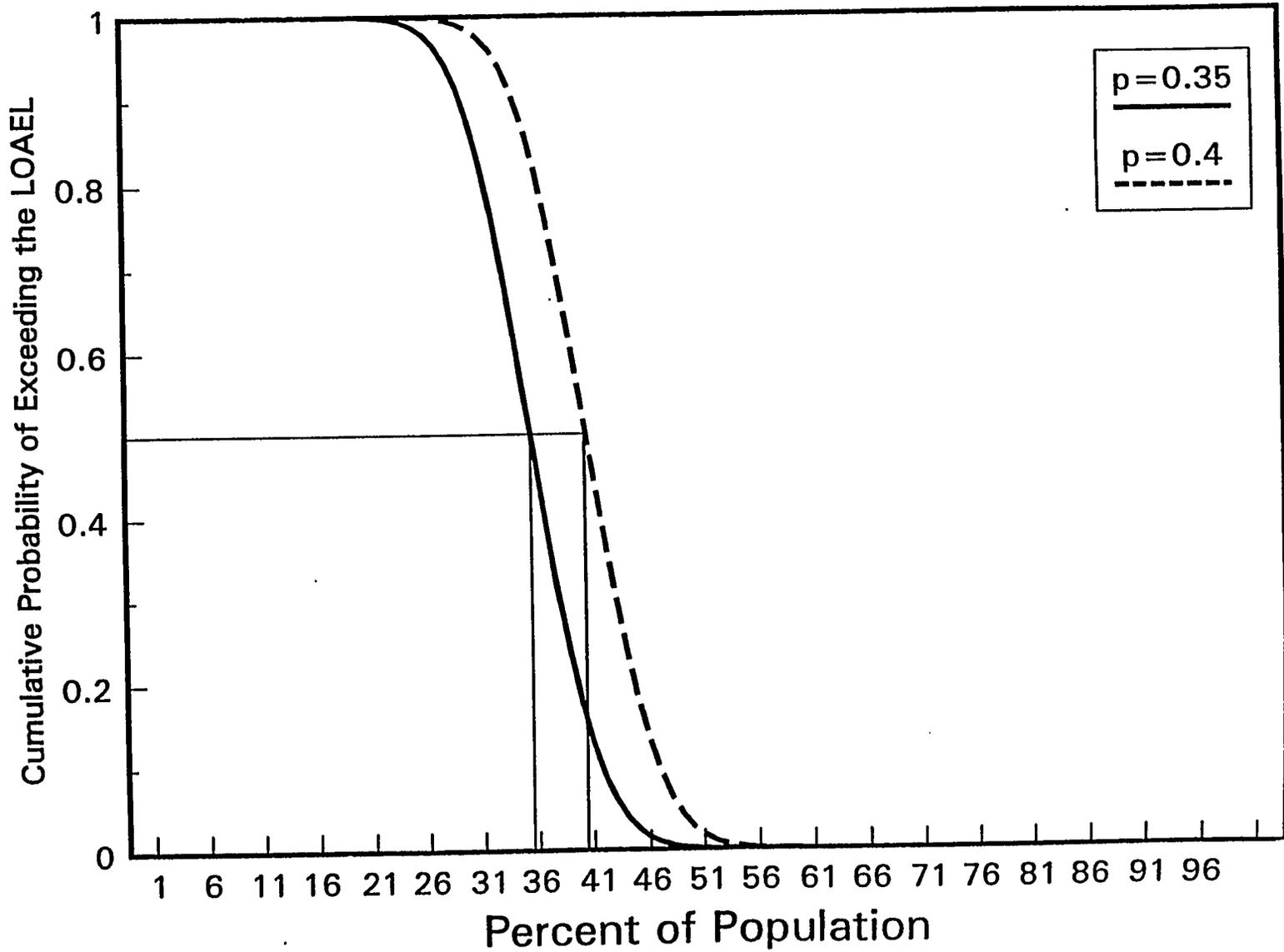


Fig. 7.10. Cumulative binomial probability of short-tailed shrews experiencing exposure to Aroclor-1260 in WAG 2 in excess of the lowest observed adverse effects level.

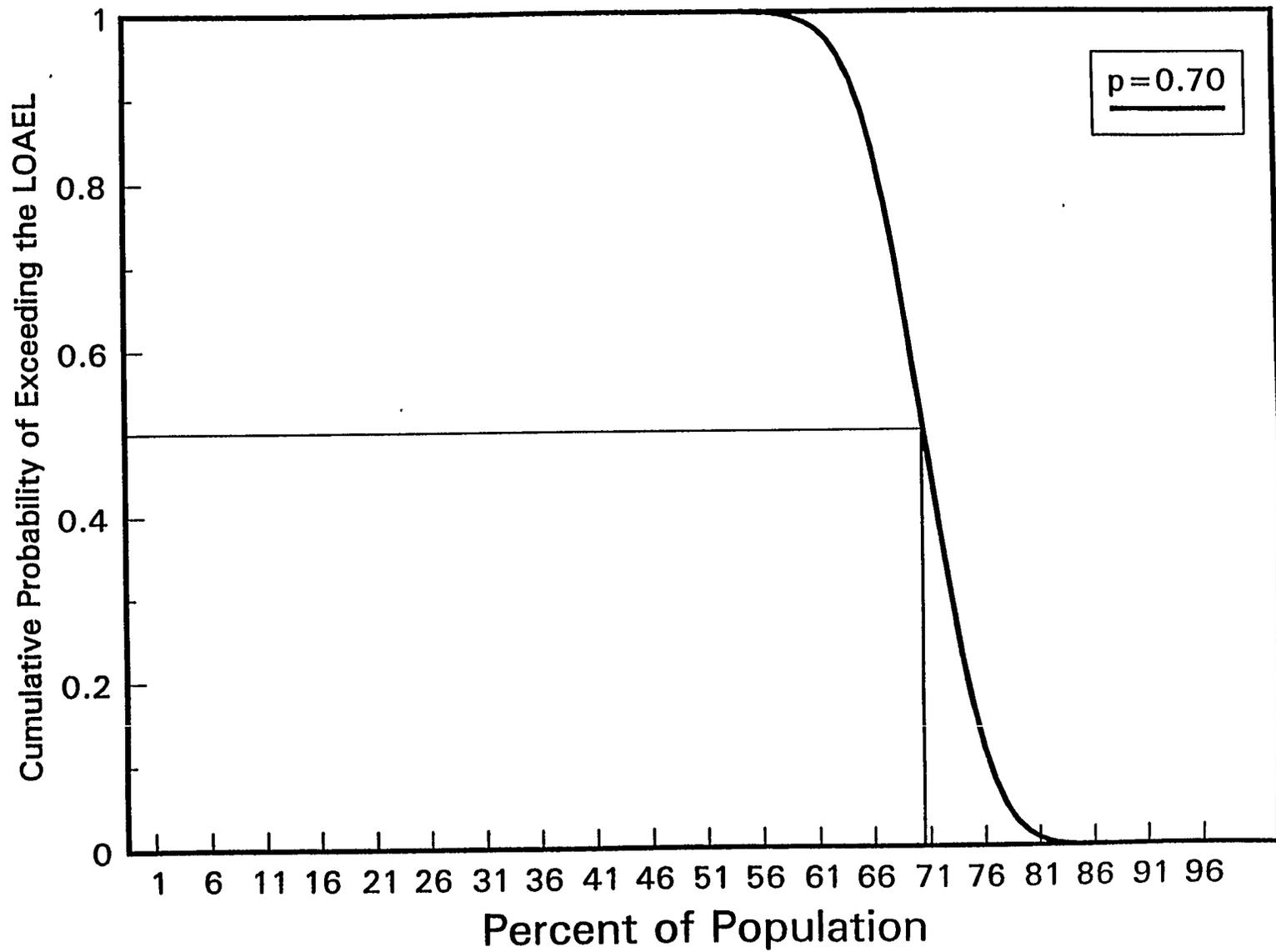
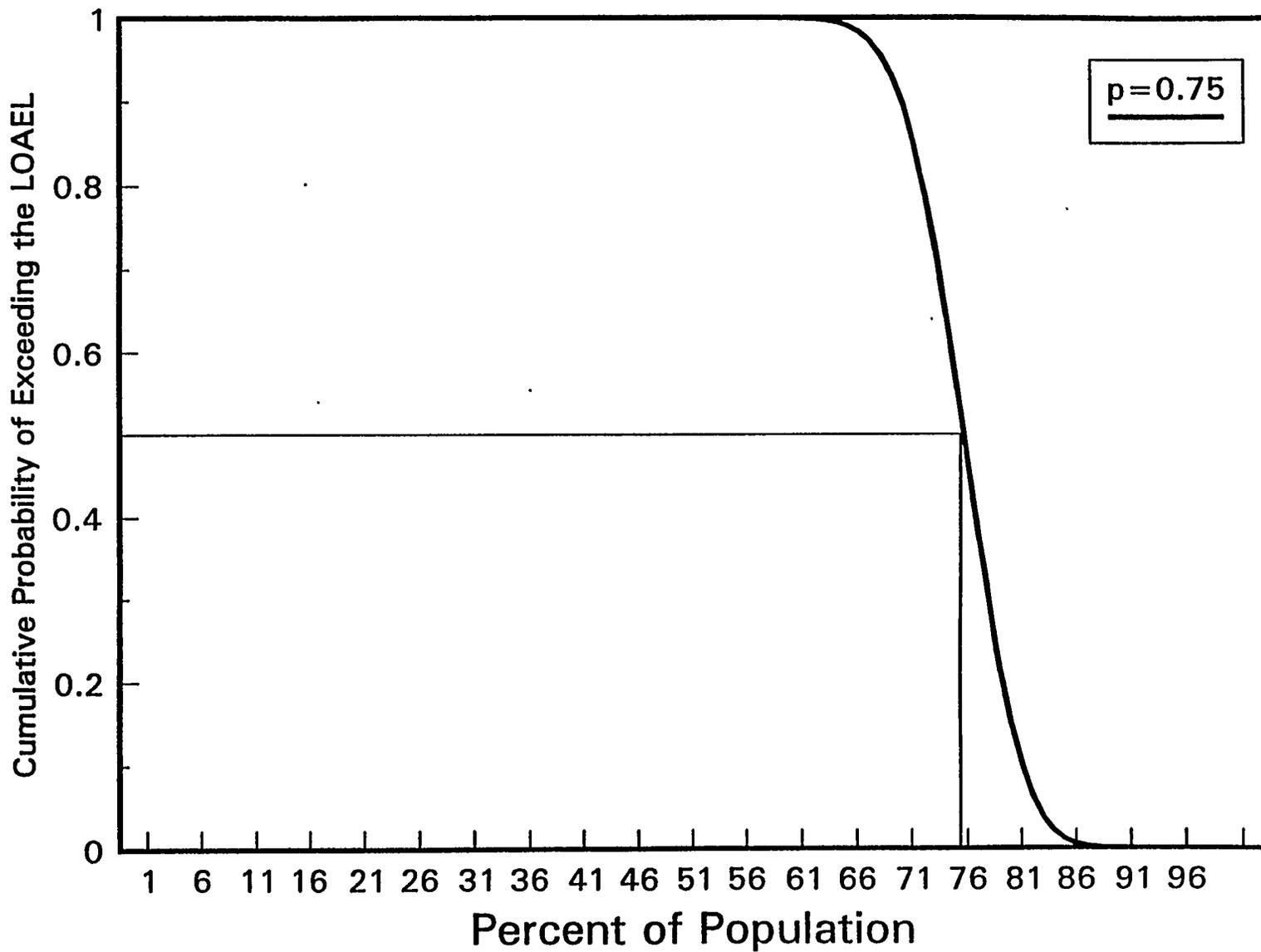


Fig. 7.11. Cumulative binomial probability of short-tailed shrews experiencing exposure to cadmium in WAG 2 in excess of the lowest observed adverse effects level.



7-23

Fig. 7.12. Cumulative binomial probability of short-tailed shrews experiencing exposure to selenium in WAG 2 in excess of the lowest observed adverse effects level.

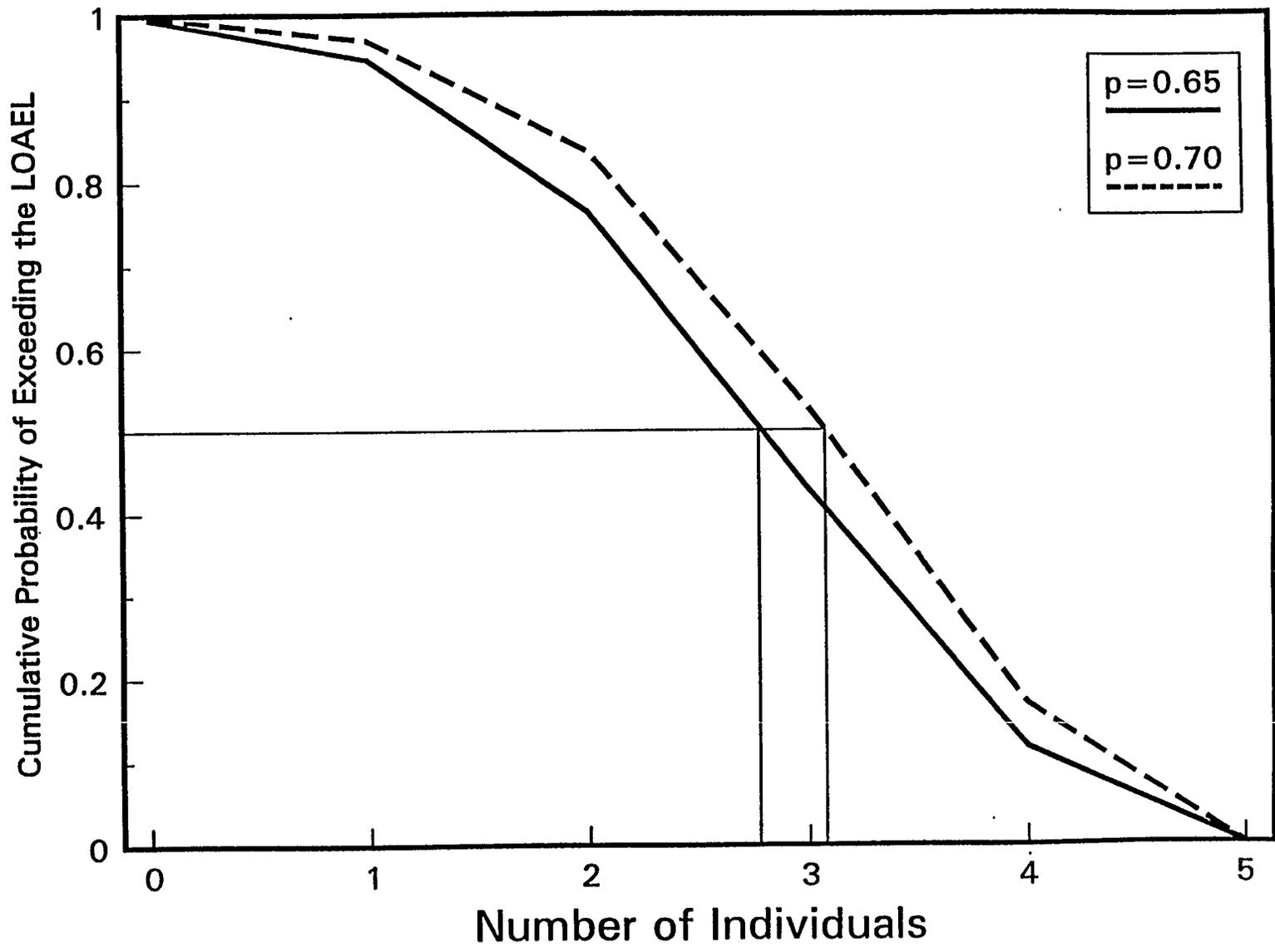


Fig. 7.13. Cumulative binomial probability of red fox experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level.

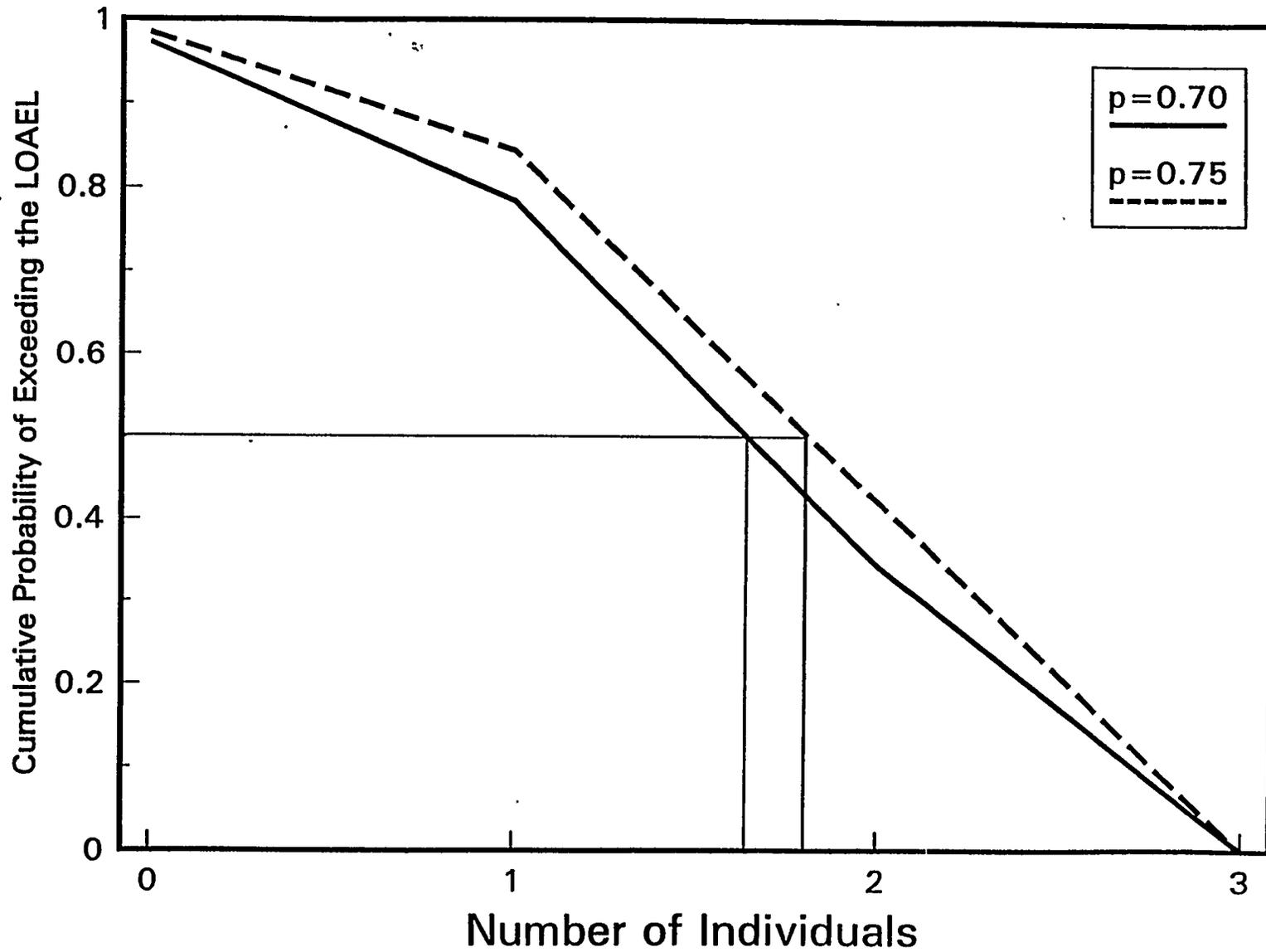


Fig. 7.14. Cumulative binomial probability of belted kingfisher experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level.

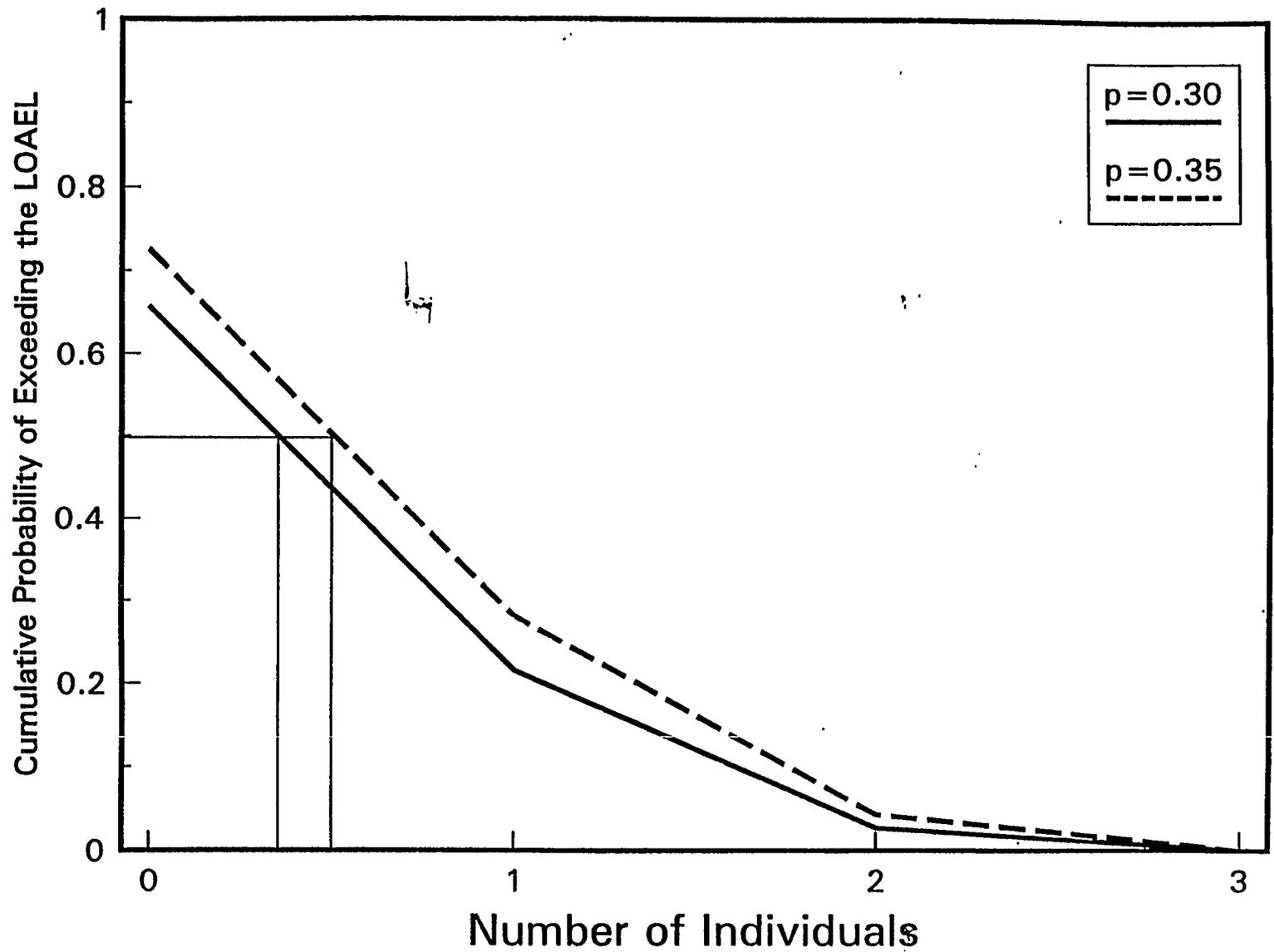


Fig. 7.15. Cumulative binomial probability of wild turkey experiencing exposure to mercury in WAG 2 in excess of the lowest observed adverse effects level.

7.1.3.2 Effects of retained contaminants

Cadmium. The mammalian NOAELs and LOAELs for cadmium were derived from a study of mice fed cadmium for multiple generations. (Schroeder and Mitchner 1971). Consumption of 2.518 mg/kg-d cadmium impaired reproduction and caused the exposed line to die out before the third generation. Only one dose level was tested. The study was considered to represent a chronic exposure; therefore, a subchronic-chronic correction factor was not employed. A NOAEL was estimated using a LOAEL-LOAEL correction factor of 0.1. The 0.25 mg/kg-d exposure was considered to be a chronic NOAEL; the 2.5 mg/kg-d exposure was considered to be a chronic LOAEL. Based on the results of Schroeder and Mitchner (1971), shrews experiencing exposure \geq LOAEL are likely to display impaired reproduction.

Chromium. The mammalian NOAEL for chromium was based on a study of rats fed Cr⁺⁶ in water for one year (Mackenzie et al. 1958). No adverse effects were observed at the highest dose of 3.28 mg/kg-d. The study was considered to represent a chronic exposure; therefore, a subchronic-chronic correction factor was not employed. The 3.28 mg/kg-d exposure was considered to be a chronic NOAEL. The mammalian LOAEL for chromium was based on a study of rats fed Cr⁺⁶ in water for 3 months (Steven et al. 1976). Mortality significantly increased among rats consuming 131.4 mg/kg-d. The study was considered to represent a subchronic exposure, therefore a subchronic-chronic correction factor was employed. The 13.14 mg/kg-d exposure was considered to be a chronic LOAEL. Based on the results of Steven et al. (1976), shrews experiencing exposure \geq LOAEL are likely to display increased mortality.

Mercury. For the purposes of this assessment, it is assumed that 100% of the mercury to which wildlife is exposed consists of methyl mercury.

Both the avian NOAEL and LOAEL are based upon a study of mallard ducks fed methyl mercury for three generations (Heinz 1979). The study was considered to represent a chronic exposure, and a subchronic-chronic correction factor was not employed. The only dose level administered, 0.064 mg/kg-d, caused hens to lay fewer eggs, lay more eggs outside of the nest box, and produce fewer ducklings. This dose level was considered to be an LOAEL. Because an experimental NOAEL was not established, the NOAEL was estimated using an LOAEL-NOAEL correction factor of 0.1. Based on the results of Heinz (1979), kingfisher, red-tailed hawks, and turkey experiencing exposure \geq LOAEL are likely to display impaired reproduction.

The fox and mink NOAELs and LOAELs for mercury were derived from a study of mink fed methyl mercury for 93 d (Wobeser et al. 1976). While consumption of 0.247 mg/kg-d methyl mercury resulted in significant mortality, weight loss, and behavioral impairment, no effects were observed at the 0.15 mg/kg-d exposure level. The 0.15 mg/kg-d exposure was considered to be an NOAEL and the 0.247 mg/kg-d exposure was considered to be an LOAEL. Because the study was subchronic in duration (<1 yr), a subchronic-chronic correction factor was applied (NOAEL=0.015, LOAEL=0.025). Based on the results of Wobeser et al. (1976), shrews, mice, fox, and mink experiencing exposure \geq LOAEL are likely to display increased mortality, weight loss, and behavioral impairment.

PCBs. The mammalian NOAEL and LOAEL for PCBs were derived from a study of mink fed Aroclor 1254 for 4.5 mo. (Aulerich and Ringer 1977). While consumption of 0.69 mg/kg-d of Aroclor 1254 reduced kit survivorship, no effects were observed at the 0.14 mg/kg-d exposure level. The 0.14 mg/kg-d exposure was considered to be a chronic NOAEL; the 0.69 mg/kg-d exposure was considered to be a chronic LOAEL. Based on the results of Aulerich and Ringer (1977), shrews experiencing exposure \geq LOAEL are likely to display reduced survivorship of young.

Selenium. The mammalian NOAELs and LOAELs for selenium were derived from a study of mice fed selenium for three generations. (Schroeder and Mitchner 1971). Consumption of 0.76 mg/kg-d selenium resulted in reduced reproductive success, and increased incidence of runts and failure to breed. Only one dose level was tested. The study was considered to represent a chronic exposure; therefore, a subchronic-chronic correction factor was not employed. An NOAEL was estimated using an LOAEL-LOAEL correction factor of 0.1. The 0.076 mg/kg-d exposure was considered to be a chronic NOAEL; the 0.76 mg/kg-d exposure was considered to be a chronic LOAEL. Based on the results of Schroeder and Mitchner (1971), shrews experiencing exposure \geq LOAEL are likely to display impaired reproduction.

7.1.3.3 Biological surveys

Mink Survey. Results of the mink survey (see Sect. 7.1.2.2.1) indicate that mink are present on the ORR and within WAG 2, have large home ranges, and do not avoid the industrial facilities on the ORR. The methods employed in the study do not allow numbers or density of mink to be determined. Concentrations of metals in hair of the single mink collected from WAG 2 were comparable to that from mink collected offsite.

Kingfisher Survey. Results of the kingfisher survey (see Sect. 7.1.2.2.2) indicate that contaminants are being accumulated by both juveniles and adult birds. While contaminants in eggshells and nestling feathers indicate exposure, there is insufficient information to evaluate the toxicological significance of this contamination.

The toxicological significance of the tissue concentrations in adult kingfisher was evaluated by comparison of burdens and effects levels reported in other bird species. This comparison suggests that it is unlikely that cadmium or lead in the kingfisher from WOC contribute significantly to risk. Leach et al. (1979) observed a 50% reduction in egg production among chickens consuming a diet containing 48 mg/kg cadmium. Cadmium concentrations in the livers and kidneys of these birds were 100 mg/kg and 40 mg/kg. Cadmium concentrations in healthy birds from unpolluted areas ranged from 0.1 to 32 mg/kg in liver and 0.3 to 137 mg/kg in kidney (Furness 1996). In comparison, cadmium concentrations in the kidney (1.53 mg/kg) and liver (0.9 mg/kg) of the kingfisher collected from the WOC watershed were significantly less than concentrations associated with reproductive impairment and at the low end of the ranges observed among healthy birds from unpolluted areas. Lead concentrations in the kidney (0.42 mg/kg) and liver (0.4 mg/kg) of the WOC kingfisher were approximately one order of magnitude lower than the minimal level at which overt toxicity is observed in birds (3 to 6 mg/kg) (Frenson 1996), suggesting that lead accumulation is unlikely to be contributing to risks to kingfishers.

In contrast to cadmium and lead, selenium and mercury burdens may present a hazard to WOC kingfishers. The concentration of selenium in the liver of Bird 3 (from WOC; 7.5 mg/kg) is less than the 10 mg/kg toxicity threshold recommend by Heinz (1996), but greater than the 3 mg/kg reproductive impairment threshold, suggesting the potential for reproductive impairment. Mercury concentrations of 49 to 125 mg/kg in kidney and 4.6 to 91 mg/kg in liver have been reported for free-living birds found dead or dying (Thompson 1996). Nephrotoxicity and kidney lesions occur in birds at mercury concentrations in kidney of 5 to 13 mg/kg (Nicholson and Osborn 1983). While mercury concentrations in the kidney (26.8 mg/kg) and liver (17.6 mg/kg) of the WOC kingfisher were generally lower than concentrations associated with mortality, the kidney concentration exceeds nephrotoxic levels, suggesting that mercury accumulation may be causing kidney damage to White Oak Creek kingfishers.

7.1.3.4 Weight of evidence

Short-tailed Shrews. One line of evidence, literature toxicity data, was available to evaluate potential risk to short-tailed shrews in WAG 2. Point estimates of exposure indicated that 18 contaminants exceeded NOAELs with 16 also exceeding LOAELs (Table C.12). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that Aroclor-1260, cadmium, chromium, mercury, and selenium present significant risks to the shrew population in WAG 2.

White-footed Mice. One line of evidence, literature toxicity data, was available to evaluate potential risk to white-footed mice in WAG 2. Point estimates of exposure indicated that ten contaminants exceeded NOAELs with five also exceeding LOAELs (Table C.13). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the mouse population in WAG 2.

White-tailed Deer. One line of evidence, literature toxicity data, was available to evaluate potential risk to white-tailed deer in WAG 2. Point estimates of exposure indicated that seven contaminants exceeded NOAELs with two also exceeding LOAELs (Table C.14). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that no contaminant presents a significant risk to the deer population in WAG 2.

Red Fox. One line of evidence, literature toxicity data, was available to evaluate potential risk to red fox in WAG 2. Point estimates of exposure indicated that 12 contaminants exceeded NOAELs with 3 also exceeding LOAELs (Table C.15). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the fox population in WAG 2.

Red-tailed Hawk. One line of evidence, literature toxicity data, was available to evaluate potential risk to red-tailed hawk in WAG 2. Point estimates of exposure indicated that three contaminants exceeded NOAELs with one also exceeding LOAELs (Table C.16). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the hawk population in WAG 2.

Wild Turkey. One line of evidence, literature toxicity data, was available to evaluate potential risk to wild turkey in WAG 2. Point estimates of exposure indicated that three contaminants exceeded NOAELs with two also exceeding LOAELs (Table C.17). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the turkey population in WAG 2.

Mink. Two lines of evidence, biological survey data and literature toxicity data, were available to evaluate potential risks to mink within WAG 2. The biological survey data indicate that mink are present within WAG 2; but due to the sampling methods employed, estimates of the abundance of the mink population cannot be made from these data. Residue analysis indicates that mink in WAG 2 have contaminant concentrations in hair similar to that in mink from offsite.

Point estimates of exposure indicated that four contaminants exceeded NOAELs with two also exceeding LOAELs (Tables C.1 and C.18). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the mink population in WAG 2.

Belted Kingfisher. Two lines of evidence, biological survey data and literature toxicity data, were available to evaluate potential risks to belted kingfisher within WAG 2. The biological survey data indicate that kingfisher are present and may be reproducing within WAG 2; but due to the sampling methods employed, estimates of the abundance of the kingfisher population cannot be made from these data. Limited residue analysis indicates that kingfisher in WAG 2 have selenium and mercury burdens in liver and kidney that may impair reproduction (selenium) or be nephrotoxic (mercury).

Point estimates of exposure indicated that mercury exceeded NOAELs and LOAELs, while PCBs exceeded NOAELs only (Table C.2). Monte Carlo simulation of exposure and comparison of these estimates to NOAELs and LOAELs (Table C.19) and calculation of binomial probability distributions (Table C.27) suggest that only mercury presents a significant risk to the kingfisher population in WAG 2.

The limited mercury body burden data corroborate the results of exposure assessment for mercury; therefore, the weight of evidence suggests that mercury presents risks to kingfisher within WAG 2. Because fish from WOC were not analyzed for selenium, the literature toxicity line of evidence is unavailable. The weight of evidence for selenium, therefore, is based solely on the body burden data, which suggest that selenium may present a risk to kingfisher within WAG 2.

7.1.4 Uncertainties Concerning Risks to Wildlife

Limited Sample Sizes. Because contaminant body burden data for belted kingfisher in WOC watershed are limited to one observation, conclusions based upon this data point should be viewed with caution. This single observation may not be representative of body burdens present in other kingfisher from WOC.

Bioavailability of Contaminants. Bioavailability of contaminants was assumed to be comparable between soil and water from WAG 2 and the diets used in the literature toxicity tests. Because bioavailability may not be comparable, exposure estimates based upon the contaminant concentrations may either under- or overestimate the actual contaminant exposure experienced.

Extrapolation from Published Toxicity Data. While published toxicity studies are available for mink, there are no published data for the other endpoints. To estimate toxicity of contaminants at the site, it was necessary to extrapolate from studies performed on test species (i.e., mallard ducks, ring-necked pheasant, and rats). While it was assumed that toxicity could be estimated as a function of body size, the accuracy of the estimate is not known. For example, hawks or kingfisher may be more or less sensitive to contaminants than ducks or pheasants due to factors other than metabolic rate.

Additional extrapolation uncertainty exists for those contaminants for which data consisted of only LOAELs or tests were subchronic in duration. For either case, an uncertainty factor of 10 was employed to estimate NOAELs or chronic data. The uncertainty factor of 10 may either over- or underestimate the actual LOAEL-NOAEL or subchronic-chronic relationship.

Toxicity of PCBs to piscivorous wildlife was evaluated using toxicity data from studies on Aroclor 1254. Because toxicity of PCB congeners can vary dramatically, the applicability of data for Aroclor 1254 is unknown.

Variable Food Consumption. While food consumption by wildlife was assumed to be similar to that reported for the same or related species in other locations, the validity of this assumption cannot be determined. Food consumption by wildlife in WAG 2 may be greater or less than that reported in the literature, resulting in either an increase or decrease in contaminant exposure.

Single Contaminant Tests vs Exposure to Multiple Contaminants in the Field. While wildlife in WAG 2 are exposed to multiple contaminants concurrently, published toxicological values only consider effects experienced by exposures to single contaminants. Because some contaminants to which wildlife are exposed can interact antagonistically, single contaminant studies may overestimate their toxic potential. Similarly, for those contaminants that interact additively or synergistically, single contaminant studies may underestimate their toxic potential.

Inorganic Forms or Species Present in the Environment. Toxicity of metal species varies dramatically depending upon the valence state or form (organic or inorganic) of the metal. For example, arsenic (III) and methyl mercury are more toxic than arsenic (V) and inorganic mercury, respectively. The available data on the contaminant concentrations in media do not report which species or form of contaminant was observed. Because benchmarks used for comparison represented the more toxic species/forms of the metals (particularly for arsenic and mercury), if the less toxic species/form of the metal were actually present in media from WAG 2, potential toxicity at the sites may be overestimated.

Uptake Factors. Soil to biota uptake factors specific to WAG 2 were unavailable. Therefore, it was assumed that the uptake factors developed as part of the Bear Creek assessment were applicable. Due to the differing geologies and histories between WAG 2 and Bear Creek, the Bear Creek uptake factors may over- or underestimate the actual biota concentrations in WAG 2.

Contaminant Concentrations in Unanalyzed Food Types. Uptake factors were not available for all food types consumed by the endpoint species. It was assumed that the uptake factors for food types for which we had data were representative of that for those without data. Due to the different life histories of among food types, contaminant burdens are likely to differ from the measured data. Therefore, an assumption of comparability among food types may either over- or underestimate exposure.

Monte Carlo Simulation. In performing Monte Carlo simulations, distributions must be assigned to parameters. Because wildlife are mobile, the mean of the contaminant concentration is likely to best represent their exposure. In this report, the contaminant concentrations in fish were assumed to be normally distributed.

7.2 RISKS TO PLANTS AND EARTHWORMS FROM EXPOSURE TO RADIONUCLIDES

7.2.1 Exposure Assessment

Native biota may receive external radiation exposure from radionuclides in air, water, soil, sediment, and other biota, such as vegetation or earthworms. Organisms also receive internal radiation

exposure from radionuclides ingested via food and water and from radionuclides absorbed through the skin and respiratory organs (IAEA 1976, Blaylock and Trabalka 1978). Evaluation of the resulting radiation doses received by biota requires quantitative information on the radionuclides to which they are exposed. In all cases the radiation source must be known in terms of the quantity of each specific radionuclide and the corresponding energy released per disintegration. Special corrections for factors that may affect exposures are applied when appropriate.

Radiation dose rates (mrad/d) were calculated for plants, earthworms, and representative terrestrial and semiaquatic wildlife species in the WOC watershed using methodology adapted from Blaylock et al. (1993), Baker and Soldat (1992), and DOE (1995b). The methodology was modified to address site-specific conditions and receptors. Current condition dose rates from internal exposures via ingestion of food and soil and inhalation of dust were evaluated, as were dose rates from external exposures via soil.

The representative terrestrial and semiaquatic wildlife selected as endpoints for the radiological assessment were the same as those for the chemical data assessment: short-tailed shrew, white-footed mouse, wild turkey, red fox, white-tailed deer, red-tailed hawk, mink, and belted kingfisher (see Sect. 7.1). Life history parameters used in the radiological assessment were identical to those used for the chemical data assessment (Table C.4-C.11). In addition, it was necessary to develop species-specific values for fraction of time spent above and below ground, underwater, or at the water's surface. A mink was assumed to spend 50% of its time above ground, 20% below ground (in burrows), 20% swimming at the water's surface, and, conservatively, 10% of its time immersed in water. Kingfishers were assumed to split their time equally between perching above water or soil and roosting in burrows. The short-tailed shrew, white-footed mouse, and red fox were all assumed to spend 75% of their time above ground and 25% below the soil surface in dens or burrows. White-tailed deer and wild turkeys spend 100% of their time above ground and were assigned a value of 1. Red-tailed hawks also spend 100% of their time above ground, but much of this is flying or perched high in trees; thus, it was assumed that hawks would only be exposed to external radiation 10% of the time.

7.2.1.1 Exposure models

Terrestrial receptors may receive both external and internal doses of radiation. Internal exposures are a result of ingestion of contaminated food, soil, or water or inhalation of contaminated soil. Receptors on the ground surface receive external exposures from contaminated surface soil via direct radiation. Both above-ground and below-ground exposures are possible, depending on the habits of the receptor. Baker and Soldat (1992) provide general equations for estimating dose rates to wildlife. An adaptation of the Blaylock et al. (1993) methodology was used in estimating radiation dose rates in the *Remedial Investigation for Waste Area Grouping 5 at Oak Ridge National Laboratory* (DOE 1995a). The Blaylock et al. methodology was further modified for the current assessment to account for the availability of site-specific uptake data. The general methodology and the equations specific to each exposure route used in estimation of dose rates for WOC wildlife are described in this subsection. Equations used in this assessment estimate the daily dose rates from current conditions. Dose rates from alpha, beta, and gamma emissions (only beta and gamma for external exposures) were calculated for each radionuclide, including the dose rates from all short-lived daughter products. Dose rates from each radionuclide were then summed over all exposure routes and all radionuclides to arrive at the overall dose rate received for each receptor at each site. Data on radionuclide activities in White Oak Creek water were not available in time for evaluation in this report. The models for estimating dose rates from exposure to radionuclides in water are provided in this section, and the analysis will be performed when the data become available. Mink and kingfisher are the only wildlife receptors likely to experience significant water-related exposures.

External Exposures: Direct radiation from soil. The equation for estimating external dose rates (mrad/d) for terrestrial receptors exposed to contaminated soil uses dose coefficients was published by Eckerman and Ryman (1993). Dose rate reduction factors are used to account for the fraction of time the receptor spends above or below ground. Dose coefficients assume the source region is a smooth plane (Eckerman and Ryman 1993), but this is rarely the case in a terrestrial habitat. A representative average dose reduction factor for ground roughness is 0.7 (Eckerman and Ryman 1993). The equation for dose from external exposures for an organism above ground is written as follows:

Above soil exposures

$$D_{above\,grd} = F_{above} F_{ruf} \sum C_{soil,i} DF_{grd,i} CFb , \quad (6)$$

where

- $D_{above\,grd}$ = external dose rate to receptor from above ground exposures to contaminated soil (mrad/d),
- F_{above} = dose rate reduction factor accounting for the fraction of time the receptor spends above ground (unitless),
- F_{ruf} = dose rate reduction factor accounting for ground roughness (unitless). [representative average of 0.7 (Eckerman and Ryman 1993) used for this assessment],
- $C_{soil,i}$ = activity of radionuclide I in surface soil (pCi/g),
- $DF_{grd,i}$ = dose coefficient for radionuclide I in surface soil (Table III.5, Eckerman and Ryman 1993) (Sv/s per Bq/m³),
- CFb = conversion factor to change Sv/s per Bq/m³ to mrad g/pCi d equals 5.12×10^{14} .

Dose from alpha radiation is not a concern for external sources as alpha radiation lacks penetrating power. The effective dose coefficients from Eckerman and Ryman (1993) incorporate both beta and gamma emissions. Radionuclide-specific parameters are provided in Table C.31. The lower of the 95% UCL and the maximum detected concentration in surface soil were used in estimating the dose rate from external exposures for each radionuclide.

Below-ground exposures are calculated similarly. However, the ground roughness dose reduction factor is not used for below-ground exposure, and the exposure fraction is adjusted to reflect the fraction of time the receptor spends below ground. This mode of exposure is treated as immersion in a continuous soil medium. To account for exposures from above and below, a factor of 2 is included. Because wild turkey, white-tailed deer, and red-tailed hawks do not go below ground, they do not receive a dose via this exposure route. The equation for below-ground external exposures is written as follows:

Below ground exposures—terrestrial wildlife

$$D_{below\,grd} = 2 F_{below} \sum C_{soil,i} DF_{grd,i} CFb , \quad (7)$$

where

- $D_{below\,grd}$ = external dose rate to receptor in burrow from contaminated soil (mrad/d),
- 2 = dose addition factor to account for receptor immersed in soil (unitless),

F_{below} = dose rate reduction factor accounting for the fraction of time the receptor spends below ground (unitless).

All other parameters are as defined for above-ground exposures.

External exposures for plants and earthworms are calculated using similar equations. Earthworms are assumed to spend 100% of their time below ground. Therefore, the F_{below} term in the below ground equation is set to 1. In addition, the dose coefficient is replaced with the energy for emission of beta or gamma particles for each radionuclide; and the CFb conversion factor is replaced with a conversion factor appropriate for converting MeV to g mrad/pCi d (0.0512).

Below-ground—Earthworms and Plants

$$D_{\text{belowgrd}} = 2 \sum C_{\text{soil},i} \epsilon_i \text{ CFa} , \quad (8)$$

where

D_{belowgrd} = external dose rate to plant or earthworm from contaminated soil (mrad/d),
 2 = dose addition factor to account for receptor immersed in soil (unitless),
 ϵ_i = energy for β or λ emissions by nuclide I (MeV/nt),
 Cfa = conversion factor to go from MeV/nt to g mrad/pCi d (5.12×10^{-2}).

All other parameters are as defined for above-ground exposures.

Plants receive external exposures to both above- and below-ground parts. The below-ground model is identical to that for earthworms. While the above-ground parts of plants receive a lower dose rate from soil than below-ground parts, the models used in this assessment estimate only the dose rate for below-ground parts.

External Exposures: Direct radiation from water. The equation for estimating external dose rates (mrad/d) for terrestrial receptors exposed to radionuclides in water uses dose coefficients published by Eckerman and Ryman (1993) and is similar to the equation used for soil. Dose rate reduction factors are used to account for the fraction of time the receptor spends underwater or at the water's surface. The equation for dose from external exposures for an organism immersed in water is written as follows:

Underwater exposures

$$D_{\text{underwater}} = F_{\text{underwater}} \sum C_{\text{water},i} \frac{1}{WD} DF_{\text{water},i} \text{ CFw} , \quad (9)$$

where

$D_{\text{underwater}}$ = external dose rate to receptor from underwater exposures to contaminated soil (mrad/d)
 $F_{\text{underwater}}$ = dose rate reduction factor accounting for the fraction of time the receptor spends underwater (unitless)
 $C_{\text{water},i}$ = activity of radionuclide I in water (pCi/L)
 WD = density of water (1000 g/L)
 $Df_{\text{water},i}$ = dose coefficient for radionuclide I for organism immersed in water (Eckerman and Ryman 1993) ($\text{Sv m}^3/\text{Bq s}$)

CFw = conversion factor to change Sv m³/Bq s to mrad g/pCi d equals 3.20×10^{14} .

Dose from alpha radiation is not a concern for external sources as alpha radiation lacks penetrating power. The effective dose coefficients from Eckerman and Ryman (1993) incorporate both beta and gamma emissions. Radionuclide-specific parameters are provided in Table C.31. The mink is the only representative wildlife species likely to spend time under water.

Exposures at the water surface are calculated similarly. However, the exposure fraction is adjusted to reflect the fraction of time the receptor spends at the water surface. Because organisms at the water's surface are exposed from below instead of from above and below, a dose reduction factor of 0.5 is applied. Mink were assumed to spend 20% of their time swimming at the water surface, and kingfishers were assumed to spend 33% of their time perched close to the water surface. Other receptors were not expected to spend significant time in water, so external water exposures were not addressed. The equation for external exposures at the water surface is written:

Water surface exposures

$$D_{\text{surface}} = F_{\text{surface}} \cdot 0.5 \sum C_{\text{water},i} \frac{1}{WD} DF_{\text{water},i} CFw, \quad (10)$$

where

- D_{surface} = external dose rate to receptor from exposures at the water surface (mrad/d),
 F_{surface} = dose rate reduction factor accounting for the fraction of time the receptor spends at the water surface (unitless),
 0.5 = dose reduction factor to account for the difference between being immersed in water and being at the water surface (unitless).

All other parameters are the same as those defined for underwater exposures.

Internal Exposures: Ingestion. Wildlife receptors may receive internal radiation doses after ingesting contaminated prey, soil, or water or after inhaling contaminated dust. Blaylock et al. (1993) provide an equation for estimating the internal dose to fish contaminated with radionuclides. This equation can be modified to address plants, earthworms, and consumers eating a variety of prey types, ingesting soil, and drinking water.

The equation is

$$D = \sum QF C_{\text{tissue}} E_i Cfa, \quad (11)$$

where

- D = internal dose rate received after ingestion of contaminated prey, soil, and water, (mrad/d) or uptake of radionuclides from soil (for plants and invertebrates),
 QF = quality factor to account for the greater biological effectiveness of α particles (20 for α ; 1 for β and λ emissions; unitless),
 C_{tissue} = activity (pCi/g) of radionuclide I in tissue of organism,
 E_i = energy for α , β , or λ emissions by nuclide I (MeV/nt),
 Cfa = Conversion factor to go from MeV/nt to g mrad/pCi d. (5.12×10^{-2}).

The primary modifications involve estimating the concentration in the consumer species and because absorbed energy fractions are generally unavailable for terrestrial wildlife, conservatively assuming that absorbed energies equaled 1 for all radionuclides and all receptors. Estimation of tissue concentrations was conducted using soil-to-tissue ratios derived from data collected in the Bear Creek watershed for three plant types (browse, canopy vegetation, and herbaceous vegetation) and small rodents; soil-to-tissue ratios for earthworms were derived from data collected at WAG 2, WAG 5, and the Bear Creek watershed. These data are likely to be more relevant to the WOC watershed than are more general literature-derived bioaccumulation factors. When site-specific uptake factors were unavailable for specific radionuclides, values were derived from those for related isotopes with measured values or from measured values of the elemental form of the radionuclide.

Dose from internal exposures was calculated for alpha, beta, and gamma energies of each radionuclide. Energies were obtained from Eckerman and Ryman (1993) and are provided in Table C.31. Because different types of radiation differ in their relative biological effectiveness per unit of absorbed dose, a quality factor derived from data on humans is normally applied (NCRP 1987). A quality factor of 1 is used for beta and gamma radiation and 20 for alpha radiation (Blaylock et al. 1993).

The lower of the 95% UCL and the maximum detected concentration in surface soil was used in exposure calculations. Radiation energies (MeV) were obtained from Eckerman and Ryman (1993) for all radionuclides and their short-lived daughter products.

When measured soil-to-tissue uptake factors were available, tissue concentrations for plants, earthworms, short-tailed shrews, and white-footed mice were estimated by multiplying the appropriate uptake factor by soil activity levels. When site-specific data were lacking, it was necessary to use transfer factors from available literature. For plants and earthworms the literature-derived values were soil-to-tissue transfer factors, so estimation of tissue concentrations was performed as described previously. However, for mammals and birds, literature values used were bioaccumulation factors defined as the ratio of activity level in receptor tissue divided by the activity level in the receptor's food. Therefore, when site-specific data were lacking, tissue concentrations were estimated as follows:

$$C_{\text{tissue}} = BAF_i (C_{ij} P_j + C_{\text{soil}} P_{\text{soil}} + C_{\text{water}} \frac{1}{WD}), \quad (12)$$

where

C_{tissue}	=	concentration of radionuclide I in receptor tissue (pCi/g),
BAF_i	=	bioaccumulation factor for radionuclide I . (pCi/g tissue over pCi/g food),
C_{ij}	=	activity (pCi/g) of radionuclide I in prey type j ,
P_j	=	proportion of prey type j in receptor's diet (unitless),
C_{soil}	=	activity (pCi/g) of radionuclide I in surface soil,
P_{soil}	=	proportion of soil in receptor diet. (unitless),
C_{water}	=	activity (pCi/L) of radionuclide I in surface water,
WD	=	density of water (1000 g/L).

It was assumed that uptake of radionuclides from ingested food, water, and soil was similar. For predators which consume small mammals, a small mammal tissue concentration is needed to estimate the dose rate to the predator. In order to simplify the exposure models, the white-footed mouse was used as the generic prey species. Therefore, small tissue concentrations used as input to the red fox, red-tailed hawk, and mink dose rate models were based on white-footed mouse tissue concentrations

calculated as described previously. Uptake factors used in the radiological assessment are provided in Table C.32.

Concentrations in fish will be estimated by multiplying water concentrations by fish bioconcentration factors obtained from Blaylock et al. (1993) when data on radionuclide activity in water become available.

Internal Exposures: Inhalation. Wildlife species using burrows receive an additional internal dose from inhalation of dust originating from contaminated soil. Intake of radionuclide *I* by inhalation is estimated as follows (DOE 1995b):

$$D_{inh} = QF F_{below} \sum C_{soil,i} A \frac{1}{AD} \epsilon_i CFa, \quad (13)$$

where

- D_{inh} = internal dose rate from inhalation of contaminated soil (mrad/d),
- F_{exp} = dose reduction factor for fraction of time receptor spends below ground (unitless),
- A = mass of respirable dust per volume of air breathed (0.1 g/m^3 ; DOE 1995b),
- AD = air density (1200 g/m^3 ; Eckerman and Ryman 1993),
- ϵ_i = α , β , or γ radiation energies for radionuclide *I* (MeV/nt),
- CFa = conversion factor to go from MeV/nt to mrad g/pCi d (5.12×10^{-2}).

Healy (1980) suggests that 0.0001 g/m^3 would be a conservative value when addressing human exposures to dust. Because burrowing animals are likely to spend a greater portion of their time in a confined space (burrow) than humans and are physically closer to the soil surface, an air mass loading of 0.1 g/m^3 was selected as a conservative estimate of the mass of respirable dust (*A*) to which these animals may be exposed.

Total internal exposures were obtained by adding ingestion and inhalation dose rates over all radionuclides, including all short-lived daughter products. In addition, the dose rate to a kingfisher collected near ORNL Building 4505 was estimated based on a measured body burden of 91.27 pCi/g Cs-137 (Table C.25).

7.2.2 Effects Levels for Radionuclides

The discharge of radioactive waste into the environment results in long-term, low dose exposure to organisms. In most cases, acute mortality can be discounted. Any potential increase in morbidity and mortality that might result from the exposure to chronic irradiation above background is unlikely to be detected because of natural fluctuations in the size of populations. The International Atomic Energy Agency (IAEA) and the Atomic Energy Act (1992) recommend limiting the dose for terrestrial organisms to 100 mrad/d (IAEA 1992). Species-specific effects data were not available, so 100 mrad/d was selected as the threshold dose for all the representative wildlife receptors. This level of exposure to the maximally exposed individual is thought to be protective of the overall receptor population (IAEA 1992, DOE 1995a).

7.2.3 Risk Characterization for Terrestrial Receptors Exposed to Radionuclides

Potential risks to plants, invertebrates, and selected terrestrial and semiaquatic wildlife receptors from exposures to radionuclides were estimated for each of four reaches along WOC by comparing

estimated dose rates to the recommended dose rate limit of 100 mrad/d. Dose rates were summed for all radionuclides over all exposure routes to obtain the overall dose in mrad/d received by each receptor at each site. Tables C.33 through C.36 present overall exposures from soil for each receptor by site and identify the radionuclides contributing the majority of the dose rate. Water-related dose rates were not estimated for this assessment.

Possible radiation effects are anticipated for plants, earthworms, and wildlife receptors frequenting all four reaches of WOC as the overall dose rates in each reach exceeded the effects threshold of 100 mrad/d. Dose rates were generally highest for all receptors at the IHP, lower in LWC and MWC WOC, and high in LMB. Watershed-wide population level effects are likely considering herbaceous plant, earthworm, short-tailed shrew, and white-footed mouse dose rates exceeded the dose rate limit in all reaches. The primary contributors to dose rates in the IHP, MWC, and LWC were $^{239,240}\text{Pu}$, ^{241}Am , and ^{137}Cs ; and ^{90}Sr , $^{239,240}\text{Pu}$, ^{241}Am , and ^{137}Cs were the primary contributors in LMB.

Intermediate Holding Pond (IHP). Potential radiation effects are expected for terrestrial plant, invertebrate, and wildlife receptors at the IHP as dose rates exceeded the effects threshold of 100 mrad/d for all receptors except the white-tailed deer and belted kingfisher (Table C.33). The kingfisher dose rate estimate was based only on external exposures to radionuclides in soil; adding likely internal exposures from ingestion of water and fish may result in a dose rate exceeding 100 mrad/d. Plants, earthworms, shrews, and mice exceeded the limit by more than a factor of 10 (ranging from 21.77 for mice to 43.4 for plants). Dose rates for turkey, fox, hawk, and mink were lower (1.5 to 3.26 times the effects threshold).

Plutonium-239/240, Americium-241, and Cesium-137 were the primary contributors to estimated dose rates for plants, earthworms, shrews, and mice (Table C.33). Plutonium-239/240 accounted for greater than 40% of the dose for these receptors. Cesium-137 was the only significant contributor to dose rates for turkey, deer, fox, hawk, mink, and kingfisher, accounting for greater than 91% of the dose rate for these receptors.

Middle White Oak Creek (MWC). Potential radiation effects are expected for terrestrial plant, invertebrate, and wildlife receptors in the vicinity of MWC as dose rates exceeded the effects threshold of 100 mrad/d for plants, earthworms, shrews, and mice (Table C.34). Dose rates for turkey, deer, fox, hawk, mink, and kingfisher were all well below the effects threshold (ranging from 3 to 17 mrad/d). Plants, earthworms, shrews, and mice exceeded the limit by more than a factor of 7 (ranging from 7.2 times the threshold for mice to 14 times for plants).

Plutonium-239/240 and Americium-241 were the primary contributors to estimated dose rates for plants, earthworms, shrews, and mice (Table C.34). Plutonium-239/240 accounted for greater than 62% of the dose for these receptors. Cesium-137 was the only significant contributor to dose rates for turkey, deer, fox, hawk, mink, and kingfisher, accounting for greater than 53% of the relatively low dose rates for these receptors.

Lower White Oak Creek (LWC). Potential radiation effects are expected for terrestrial plant, invertebrate, and wildlife receptors in the vicinity of LWC as dose rates exceeded the effects threshold of 100 mrad/d for plants, earthworms, shrews, and mice (Table C.35). Dose rates for turkey, deer, fox, hawk, mink, and kingfisher were all below the effects threshold (ranging from 10 to 49 mrad/d). Plants, earthworms, shrews, and mice exceeded the limit by more than a factor of 7 (ranging from 7.2 times the threshold for mice to 13.5 times for plants).

Plutonium-239/240 and Americium-241 were the primary contributors to estimated dose rates for plants, earthworms, shrews, and mice (Table C.35). Plutonium-239/240 accounted for greater than 51% of the dose for these receptors. Cesium-137 was the only significant contributor to dose rates for turkey, deer, fox, hawk, mink, and kingfisher, accounting for greater than 84% of the dose rates for these receptors.

Lower Melton Branch (LMB). Potential radiation effects are expected for terrestrial plant, invertebrate, and wildlife receptors at the LMB as dose rates exceeded the effects threshold of 100 mrad/d for all receptors except the belted kingfisher (Table C.36). The kingfisher dose rate estimate was based only on external exposures to radionuclides in soil; adding likely internal exposures from ingestion of water and fish may result in a dose rate exceeding 100 mrad/d. Plants, earthworms, fox, and hawk exceeded the limit by more than a factor of 10 (ranging from 10.87 for the hawk to 30.5 for earthworms). Dose rates for shrews, mice, turkeys, deer, and mink were lower (2.75 to 6.22 times the effects threshold).

Plutonium-239/240 (66.6%) and Cesium-137 (17.1%) were the primary contributors to the estimated dose rate for plants. Strontium-90 and Curium-244 accounted for over 80% of the estimated dose rate for shrews and mice (Table C.36). Strontium-90 was the only significant contributor to dose rates for earthworms, turkey, deer, fox, hawk, and mink, accounting for greater than 92% of the dose rate for these receptors.

Kingfisher Tissue Data. In addition to evaluation of potential radionuclide exposures in the four reaches of WOC, the internal dose rate for a single kingfisher collected near Building 4505 at Oak Ridge National Laboratory, upstream of WOC, was estimated based on a measured body burden of 91.27 pCi/g of Cesium-137 (Table C.25) (Baron and Ashwood 1996). The model for internal ingestion exposures described in Sec. 7.2.1.1 was used to estimate a dose rate of 3.8 mrad/d for this kingfisher, a rate well below the effects threshold of 100 mrad/d. Baron and Ashwood suggest that the high levels of Cesium-137 in this kingfisher indicate its foraging territory includes WOC or nearby surface impoundments. If the Cs-137 body burden found in the kingfisher from the Building 4505 area is representative of body burdens in kingfishers along WOC, the overall dose rates for kingfishers along the creek may be below 100 mrad/d. Cs-137 was the primary contributor to external exposures from radionuclides in soil for kingfishers along all four reaches of WOC, and the highest dose rate (at the IHP) was estimated at 64 mrad/d (Tables C.33-C.36).

7.7.4 Uncertainties in the Radiological Risk Assessment

There are a number of areas of uncertainty in the estimation of risks to wildlife from exposure to radionuclides at WOC. It is believed that the methodology used in this assessment overestimates the dose rates that would be received. Whereas some of the information needed to implement the methodology is well known, much is unknown or unspecified statistically. A conservative but reasonable approach to model assumptions and radiological exposure scenarios was adopted for this assessment in order to avoid underestimating risks to biota.

- It was conservatively assumed that energy absorption by the wildlife receptors evaluated in this assessment was 100%. While larger organisms are likely to absorb a greater fraction of energy than smaller organisms, it is unlikely that actual absorption would be as high as 100%. Thus, radiation dose rates may be lower than those reported here.
- The dose coefficients obtained from Eckerman and Ryman (1993) used to estimate dose rates from external exposures were developed for application in determining dose rates to humans. These dose coefficients were applied directly for wildlife receptors, but the actual dose

coefficients for wildlife, given differences in size, behavior, and general morphology, may be greater or lesser than those developed for humans. However, similar values have not been developed specifically for wildlife receptors.

- Uptake factors from soil to earthworms were not available for many radionuclides. If uptake factors were unavailable, the higher of the plant or mammal uptake factors was used for earthworms. It is unknown whether actual earthworm uptake factors for these radionuclides would be higher or lower than those used, but use of the larger of the plant and mammal values was a conservative approach.
- Mammal soil-to-tissue uptake factors were calculated from measured data on small rodents. While these clearly apply directly to white-footed mice, they were also used for the other small mammalian receptor, the short-tailed shrew. Short-tailed shrew diets differ somewhat from that of white-footed mice as shrews eat a higher proportion of invertebrates. However, it was assumed that site-specific data on mice were more applicable to shrews than literature-derived bioaccumulation factors.
- Uptake factors were not available for birds such as the red-tailed hawk. Radionuclide activities in red-tailed hawk tissues were estimated using mammal uptake factors assuming that uptake would be similar for hawks and mammals. It is unknown whether actual bird uptake factors would be higher or lower than those used here.
- Literature-derived bioaccumulation factors were used to estimate radionuclide activities in mammal and bird tissues. While these values are believed to be representative of bioaccumulation likely to be observed on site, actual values may be higher or lower than used in this assessment.
- Dietary information for the wildlife receptors evaluated here was obtained from available literature. Diets of given species in the WOC watershed may vary somewhat from those reported in the literature from other areas.
- Species-specific time budget information is lacking, so it was necessary to make a number of assumptions regarding the fraction of time each receptor spends above or below ground, underwater, or at the water's surface. The fractions used in this assessment were selected based on general knowledge of species behavior patterns, but actual fractions may be higher or lower.
- The air mass loading factor of 0.1 g/m^3 used in estimating exposures from inhalation of radionuclide-contaminated dust was selected as a conservative value. Healy (1980) suggested 0.0001 g/m^3 would be a conservative value for estimating human exposures from inhalation of dust.

8. SUMMARY AND CONCLUSIONS

This screening assessment is intended to indicate whether there are credible hazards to ecological endpoints in the WOC watershed due to contamination and whether there are data gaps that need to be filled. The following are conclusions concerning the hazards:

- Radionuclides in WAG 2 do not pose a hazard to fish or aquatic invertebrates. Strontium-90 in three seeps exceeds the screening benchmarks for aquatic life, but these seeps do not support communities of aquatic macroorganisms.
- Radionuclides in WAG 2 pose a hazard to terrestrial plants, wildlife, and soil invertebrates in all four reaches of WOC and in lower MB. Exposures were highest in the IHP reach.
- The unfiltered water in all reaches exceeded water quality criteria and other toxicological benchmarks for aquatic life.
- Water from WOC adjacent to and below ORNL was toxic in a fish embryo-larval test but not the standard subchronic tests.
- The fish community in WOC has low species richness, but this may be due to lack of recovery from past toxicity. However, the riffle invertebrate community, which is exposed primarily to chemicals in water like the fish and which has flying stages which should allow recovery, also has low species richness.
- Sediments from the IHP, MWC, and LMB are contaminated to levels that have been associated with toxic effects at other sites. Contaminants include metals, PAHs, and PCBs.
- The species richness of benthic invertebrates from WAG 2 sediments is lower than in background WOC sediments but not other reference streams. The exception is WOL, which has very low species richness; but no comparable reference lakes were characterized.
- Chromium and mercury levels in IHP and LWC soils and mercury in MWC and LMB soils exceeded levels that were reported to be toxic to earthworms.
- Multiple metals were found in IHP, MWC, LWC, and LMB soils at concentrations that have been reported to be toxic to plants.
- Metals in seep waters in the W4T and WS reaches are at concentrations that have been reported to be toxic to plants.
- Hazards to wildlife were found in all WOC reaches. However, the hazards were greatest for mercury among chemicals, for IHP among reaches, and for shrews among species.
- Kingfishers from WOC show elevated levels of contaminants, and the one adult found had tissue levels of mercury and selenium that are indicative of toxicity. Mink from WOC do not appear to be contaminated.

The following are major uncertainties that could be addressed by additional data collection:

- Background concentrations need to be better characterized for the seeps, surface waters, sediments, and floodplain soils.
- The contamination of soil and biota on the burial grounds other than WAGs 5 and 6 are unknown.
- The bioavailable concentrations of metals in WOC watershed are unknown.
- The toxicity of soil and sediment in the watershed are unknown.
- The toxicity of water is unknown for some reaches and has been irregular in others (Table 3.4).
- The fish communities have not been quantitatively characterized for the smaller tributaries (Table 3.4).
- The chemical composition of fish is unknown for some tributaries and reaches (Table 3.4).
- Because of the high Se levels in a kingfisher, Se levels should be characterized in fish.

In sum, plausible hazards exist in all reaches of WAG 2 and to all ecological endpoints. Radionuclides, organic chemicals, and metals are all implicated. However, none of the risk estimates or observations of the state of the watershed suggest that there is a need for an accelerated response. That is, no threatened or endangered species are at risk and no wetlands or other highly valued populations or ecosystems are experiencing catastrophic effects. Some parts of the watershed are uncharacterized and should be surveyed and sampled before the RI is completed. These results imply that a more complete data set should be assembled and used to prepare a definitive ecological risk assessment for the WOC watershed.

9. REFERENCES

- Ables, E.D. 1969. "Home range studies of red foxes (*Vulpes vulpes*)."*J. Mamm.* 50:108-120.
- Alexander, G.R. 1977. "Food of vertebrate predators on trout waters in north central lower Michigan."*Mich. Acad.* 10:181-195.
- Ambrose, A.M., P.S. Larson, J.F. Borzelleca, and G.R. Hennigar, Jr. 1976. "Long-term toxicological assessment of nickel in rats and dogs."*J. Food Sci. Tech.* 13:181-187.
- Andersen, D.E., and O.J. Rongstad. 1989. "Home-range estimates of red-tailed hawks based on random and systematic relocations."*J. Wildl. Manage.* 53:802-807.
- Arnold, T.W., and E.K. Fritzell. 1987. "Food habits of prairie mink during the waterfowl breeding season."*Can. J. Zool.* 65:2322-2234.
- Ashwood, T.L. (ed.) 1993. *Seventh annual report on the ORNL Biological Monitoring and Abatement Program*. ORNL/TM-00000. Oak Ridge National Laboratory, Oak Ridge, TN.
- Ashwood, T.L. (ed.) 1994. *Eighth annual report on the ORNL Biological Monitoring and Abatement Program*. ORNL/TM-12767. Oak Ridge National Laboratory, Oak Ridge, TN.
- Aulerich, R. J., and R. K. Ringer. 1977. "Current status of PCB toxicity to mink and effect on their reproduction." *Arch. Environ. Contam. Toxicol.* 6:279-292.
- Aulerich, R.J., R.K. Ringer, M.R. Bleavins, and A. Napolitano. 1982. "Effects of supplemental dietary copper on growth, reproductive performance and kit survival of standard dark mink and the acute toxicity of copper to mink." *J. Animal Sci.* 55: 337-343. (Cited in ATSDR 1990).
- Azar, A., H.J. Trochimowicz, and M.E. Maxwell. 1973. "Review of lead studies in animals carried out at Haskell Laboratory: two-year feeding study and response to hemorrhage study." In *Environmental Health Aspects of Lead: Proceedings, International Symposium*, D. Barth et al., (eds.). Commission of European Communities. pp. 199-210.
- Barber, H.L. 1984. Eastern mixed forest. pp. 345-354. In Halls, L.K., (ed.) *White-tailed Deer: Ecology and Management*. Harrisburg, PA: Stackpole Books.
- Baes, C. F., III, R. D. Sharp, A. L. Sjoreen, and R. W. Shor. 1984. *A Review and Analysis of Parameters for Assessing Transport of Environmentally Released Radionuclides through Agriculture*. ORNL-5786. Health and Safety Research Division, Oak Ridge National Laboratory, Oak Ridge, TN. 150p.
- Baker, D.A., and J.K. Soldat. 1992. *Methods for Estimating Doses to Organisms from Radioactive Materials Released into the Aquatic Environment*. PNL-8150. Pacific Northwest Laboratory, Richland, WA.
- Baron, L. A., and T. L. Ashwood. 1996. Monitoring bioaccumulation of contaminants in the belted kingfisher (*Ceryle alcyon*). Environmental Monitoring and Assessment. In Review.

- Barrett, G.W., and K.L. Stueck. 1976. "Caloric ingestion rate and assimilation efficiency of the short-tailed shrew, *Blarina brevicauda*." *Ohio J. Sci.* 76: 25-26.
- Barsotti, D.A., R.J. Marlar, and J.R. Allen. 1976. "Reproductive dysfunction in Rhesus monkeys exposed to low levels of polychlorinated biphenyls (Aroclor 1248)." *Fd. Cosmet. Toxicol.* 14: 99-103.
- Batzli, G.O. 1977. "Population dynamics of the white-footed mouse in flood plain and upland forests." *Am. Midl. Nat.* 97: 18-32.
- Bent, A.C. 1940. Life histories of North American cuckoos, goat suckers, hummingbirds, and their allies. Washington, D.C.: U.S. Government Printing Office; Smithsonian Inst. U.S. Nat. Mus., Bull. 176.
- Beyer, W.N., E. Conner, and S. Gerould. 1994. "Survey of soil ingestion by wildlife." *J. Wildl. Mgmt.* 58:375-382.
- Blaylock, B.G., and J.R. Trabalka. 1978. "Evaluating the effects of ionizing radiation on aquatic organisms." pp. 103-152 in J.T. Lett and H. Alder, eds. *Advances in Radiation Biology*. Academic Press, New York.
- Blaylock, B. G., M. L. Frank, F. O. Hoffman, L. A. Hook, G. W. Suter, and J. A. Watts. 1992. *Screening of Contaminants in Waste Area Grouping 2 at Oak Ridge National Laboratory, Oak Ridge, Tennessee*. ORNL/ER-62. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Blaylock, B.G., M.L. Frank, and B.R. O'Neal. 1993. *Methodology for Estimating Radiation Dose Rates to Freshwater Biota Exposed to Radionuclides in the Environment*. ES/ER/TM-78. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Bleavins, M.R., and R.J. Aulerich. 1981. "Feed consumption and food passage time in mink (*Mustela vison*) and European ferrets (*Mustela putorius furo*)." *Lab. Anim. Sci.* 31:268-269.
- Borders, D. M., D. J. Pridmore, J. J. Beauchamp, D. E. McConnell, S. T. Puruker, and T. M. Scanlon. 1996. *Waste Area Grouping 2 Phase I Remedial Investigation Report: Tributary Data Assessment*. ORNL/ER-362. Energy Systems Restoration Program, Oak Ridge, Tenn.
- Borzelleca, J.F., L.W. Condie, Jr., and J.L. Egle, Jr. 1988. "Short-term toxicity (one-and ten-day gavage) of barium chloride in male and female rats." *J. American College of Toxicology.* 7:675-685.
- Brooks, R.P., and W.J. Davis. 1987. "Habitat selection by breeding belted kingfishers (*Ceryle alcyon*)." *Am. Midl. Nat.* 117:63-70.
- Brown, L., and D. Amadon. 1968. *Eagles, Hawks, and Falcons of the World*, Vol. 1. McGraw-Hill. New York
- Buben, J.A., and E.J. O'Flaherty. 1985. "Delineation of the role of metabolism in the hepatotoxicity of trichloroethylene and perchloroethylene: a dose-effect study." *Toxicol. Appl. Pharmacol.* 78:105-122.

- Buckner, C.H. 1964. "Metabolism, food capacity, and feeding behavior in four species of shrews." *Can. J. Zool.* 42:259-279.
- Buckner, C.H. 1966. "Populations and ecological relationships of shrews in Tamarack bogs of southeastern Manitoba." *J. Mamm.* 74:181-194.
- Burgess, S.A., and J.R. Bider. 1980. "Effects of stream habitat improvements on invertebrates, trout populations, and mink activity." *J. Wildl. Manage.* 44:871-880.
- Burt, W.H., and R.P. Grossenheider. 1976. *A Field Guide to the Mammals of America North of Mexico*. 3rd ed., Houghton Mifflin Co., Boston.
- Cain, B.W., and E.A. Pafford. 1981. "Effects of dietary nickel on survival and growth of mallard ducklings." *Arch. Environm. Contam. Toxicol.* 10:737-745.
- Calder, W.A., and E.J. Braun. 1983. "Scaling of osmotic regulation in mammals and birds." *Am. J. Physiol.* 224:Rr601-R606.
- Carriere, D., K. Fischer, D. Peakall, and P. Angehrn. 1986. "Effects of dietary aluminum in combination with reduced calcium and phosphorus on the ring dove (*Streptopelia risoria*)." *Water, Air, and Soil Poll.* 30:757-764.
- Chew, R.M. 1951. "The water exchanges of some small mammals." *Ecol. Monogr.* 21(3):215-224.
- Craighead, J.J., and F.C. Craighead. 1969, *Hawks, Owls, and Wildlife*. Dover Publ. Co. New York.
- Curl, E. A., P. J. Edwards, C. Elliott, and J. P. Leahey. 1987. "The conjugation and accumulation of metabolites of cypermethrin by earthworms." *Pesticide Sci.* 20:207-222.
- Dahlgren, R.B., R.L. Linder and C.W. Carlson. 1972. "Polychlorinated biphenyls: their effects on penned pheasants." *Environmental Health Perspectives* 1:89-101.
- Davis, W.J. 1982. "Territory size in *Megasceryle alcyon* along a stream habitat." *The Auk.* 99:353-362.
- Davis A., R. Barale, G. Brun, et al. 1987. "Evaluation of the genetic and embryotoxic effects of bis(tri-n-butyltin)oxide (TBTO), a broad spectrum pesticide, in multiple in vivo and in vitro short-term tests." *Muta. Res.* 188:65-95.
- DeGraaf, R.M., G.M. Witman, J.W. Lanier, B.J. Hill, and J.M. Keniston. 1981. *Forest Habitat for Birds of the Northeast*. U.S.D.A. Forest Service. Northeast Forest Experiment Station and Eastern Region.
- DOE (U.S. Department of Energy). 1995a. *Remedial Investigation Report on Waste Area Grouping 5 at Oak Ridge National Laboratory, Oak Ridge, Tennessee*. DOE/OR/01-1326. U.S. Department of Energy, Oak Ridge, Tenn.
- DOE (U.S. Department of Energy). 1995. *Remedial Investigation Report on Waste Area Grouping 5 at Oak Ridge National Laboratory, Oak Ridge, Tennessee, Vol. 4, Appendix C-Risk Assessment*, DOE/OR/01-1326&D1/V4. U.S. Department of Energy, Oak Ridge, Tenn.

- DOE (U.S. Department of Energy). 1995b. *Remedial Investigation/Feasibility Study for the Clinch River/Poplar Creek Operable Unit*. DOE/OR/01-1393. U.S. Department of Energy, Oak Ridge, Tenn.
- Domingo, J.L., J.L. Paternain, J.M. Llobet, and J. Corbella. 1986. "Effects of vanadium on reproduction, gestation, parturition and lactation in rats upon oral administration." *Life Sci.* 39:819-824.
- Dowdy, S., and S. Wearden. 1983. *Statistics for Research*. John Wiley and Sons. New York.
- Dunning, J.B. 1984. *Body Weights of 686 Species of North American Birds*. West. Bird Banding Assoc. Monogr. No. 1. Eldon Publ. Co. Cave Creek, Ariz.
- Eckerman, K.F., and J.C. Ryman. 1993. *External Exposure to Radionuclides in Air, Water, and Soil*. EPA 402-R-93-081 Federal Guidance Report No. 12. Office of Radiation and Indoor Air, U.S. Environmental Protection Agency, Washington, D.C.
- Edens, F.W., E. Benton, S.J. Bursian, and G.W. Morgan. 1976. "Effect of Dietary Lead on Reproductive Performance in Japanese Quail, *Coturnix coturnix japonica*." *Toxicol. Appl. Pharmacol.* 38:307-314.
- EPA (U. S. Environmental Protection Agency). 1986c. *90-day Gavage Study in Albino Rats Using Acetone*. Office of Solid Waste, Washington, D.C.
- EPA (U. S. Environmental Protection Agency). 1993a. *Wildlife Exposure Factors Handbook*. Vol. I. EPA/600/R-93/187a. Office of Research and Development, Washington, D.C.
- EPA (U. S. Environmental Protection Agency). 1993b. *Wildlife Exposure Factors Handbook*. Vol. II. EPA/600/R-93/187b. Office of Research and Development, Washington, D.C.
- EPA. (U. S. Environmental Protection Agency). 1993d. *Wildlife Criteria Portions of the Proposed Quality Guidance for the Great Lakes System*. EPA/822/R-93/006. Office of Science and Technology, Washington, D.C.
- EPA. (U. S. Environmental Protection Agency). 1995. *Great Lakes Water Quality Initiative Technical Support Document for Wildlife Criteria*. EPA/820/B-95/009. U.S. Environmental Protection Agency, Washington D.C.
- Feron, V.J., C.F.M. Hendriksen, A.J. Speek, et al. 1981. "Lifespan oral toxicity study of vinyl chloride in rats." *Food Cosmet. Toxicol.* 13:633-638.
- Ford, C. J., J. E. Nyquist, S. T. Puruker, and R. F. Winterfield. 1996. *Characterization and Inventory of Contaminants in WAG 2 Floodplain Soils of White Oak Creek*. ORNL/TM-13208. Energy Systems Restoration Program, Oak Ridge, TN.
- Formigli, L., R. Scelsi, P. Poggi, C. Gregotti, A. DiNucci, E. Sabbioni, L. Gottardi, and L. Manzo. 1986. "Thallium induced testicular toxicity in the rat." *Environ. Res.* 40:531-539.

- Franson, J. C. 1996. "Interpretation of tissue lead residues in birds other than waterfowl." pp. 265-279. In W. N. Beyer, G. H. Heinz and A. W. Redmon-Norwood (eds.), *Environmental Contaminants in Wildlife*. Lewis Publishers, New York.
- Furness, R. W. 1996. "Cadmium in birds. pp.389-404. In W. N. Beyer, G. H. Heinz and A. W. Redmon-Norwood (eds.), *Environmental Contaminants in Wildlife*. Lewis Publishers, New York.
- George, S.B., J.R. Choate, and H.H. Genoways. 1986. "*Blarina brevicauda*." *Mamm. Species*. 261:1-9.
- Gerell, R. 1970. "Home ranges and movements of the mink *Mustela vison* Schreber in southern Sweden." *Oikos* 21:160-173.
- Getz, L.L. 1989. "A 14-year study of *Blarina brevicauda* populations in east-central Illinois." *J. Mamm.* 70: 58-66.
- Green, D.A. and J.S. Millar. 1987. "Changes in gut dimensions and capacity of *Peromyscus maniculatus* relative to diet quality and energy needs." *Can. J. Zool.* 65:2159-2162.
- Hamilton, W.J., Jr. 1940. "The summer food of minks and raccoons on the Montezuma marsh." *New York. J. Wildl. Manage.* 4:80-84.
- Hammonds, J.S., F.O. Hoffman, and S.M. Bartell. 1994. *An Introductory Guide to Uncertainty Analysis in Environmental and Health Risk Assessment*. ES/ER/TM-35/R1. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Haseltine, S.D., L. Sileo, D.J. Hoffman, and B.D. Mulhern. 1985. "Effects of chromium on reproduction and growth in black ducks."
- Healy, J.W. 1980. "Review of resuspension models." pp. 209-235. In W. C. Hanson (ed.) *Transuranic Elements in the Environment*. U.S. Department of Energy, Washington, D.C.
- Heinz, G.H. 1979. "Methyl mercury: reproduction and behavioral effects on three generations of mallard ducks." *J. Wildl. Mgmt.* 43:394-401.
- Heinz, G. H. 1996. "Selenium in birds." p. 447-458. In W. N. Beyer, G. H. Heinz, and A. W. Redmon-Norwood (eds.), *Environmental Contaminants in Wildlife*. Lewis Publishers, New York.
- Heinz, G.H. , D.J. Hoffman, A.J. Krynitsky, and D.M.G. Weller. 1987. "Reproduction in mallards fed selenium." *Environ. Toxicol. Chem.* 6:423-433.
- Hicks, D. 1996. *Waste Area Grouping 2 Phase I Remedial Investigation Seep Task Data Report: Contaminant Source Area Assessment*. ORNL/ER-363. Energy Systems Restoration Program, Oak Ridge, TN.
- Hockman, J. G., and J. A. Chapman. 1983. "Comparative feeding habits of red foxes (*Vulpes vulpes*) and gray foxes (*Urocyon cinereoargenteus*) in Maryland." *Amer. Midl. Natur.* 110(2): 276-285.
- IAEA (International Atomic Energy Agency). 1976. *Effects of Ionizing Radiation on Aquatic Organisms and Ecosystems*. IAEA Technical Report Series 172. Vienna, Austria.

- IAEA (International Atomic Energy Agency). 1982. *Generic Models and Parameters for Assessing the Environmental Transfer of Radionuclides from Routine Releases: Exposures of Critical Groups*. IAEA Report No 57, IAEA, Vienna, Austria.
- IAEA (International Atomic Energy Agency). 1992. *Effects of Ionizing Radiation on Plants and Animals at Levels Implied by Current Radiation Protection Standards*. IAEA Technical Report Series 332. Vienna, Austria.
- IAEA (International Atomic Energy Agency). 1994. *Handbook of Parameter Values for the Prediction of Radionuclide Transfer in Temperate Environments*. IAEA Technical Report Series 364, Vienna, Austria.
- ICRP (International Commission on Radiological Protection). 1983. *Radionuclide Transformations: Energy and Intensity of Emissions*. ICRP Report No 38.
- Janes, S.W. 1984. "Influences of territory composition and interspecific competition on red-tailed hawk reproductive success." *Ecology*. 65: 862-870.
- Johnson, D., Jr., A.L. Mehring, Jr., and H.W. Titus. 1960. "Tolerance of chickens for barium." *Proc. Soc. Exp. Biol. Med.* 104:436-438.
- Korschgen, L.J. 1958. "December food habits of mink in Missouri." *J. Mamm.* 39:521-527.
- Korschgen, L. J. 1967. Feeding habits and foods. pp. 137-198. In the wild turkey and its management. O. H. Hewitt (ed.). The Wildlife Society, Inc., Bethesda, Md.
- Lackey, J.A., D.G. Huckaby, and B.G. Ormiston. 1985. "Peromyscus leucopus." *Mamm. Species* 247:1-10.
- Lamb, J.C., IV, R.E. Chapin, J. Teague, A.D. Lawton, and J.R. Reel. 1987. "Reproductive effects of four phthalic acid esters in the mouse." *Toxicol. Appl. Pharmacol.* 88:255-269.
- Landrum, C.L., T.L. Ashwood, and D.K. Cox. 1993. *Belted Kingfishers as Ecological Monitors of Contaminations: A Review*. ORNL/M-2533. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Lane, R. W., B. L. Riddle, and J. F. Borzelleca. 1982. "Effects of 1,2-dichloroethane and 1,1,1-trichloroethane in drinking water on reproduction and development in mice." *Toxicol. Appl. Pharmacol.* 63:409-421.
- Laskey, J.W., G.L. Rehnberg, J.F. Hein, and S.D. Carter. 1982. "Effects of chronic manganese (Mn_3O_4) exposure on selected reproductive parameters in rats." *J. Toxicol. Environ. Health.* 9:677-687.
- Laskey, J.W., and F.W. Edens. 1985. "Effects of chronic high-level manganese exposure on male behavior in the Japanese quail (*Coturnix coturnix japonica*)." *Poult. Sci.* 64:579-584.
- Leach, R. M., K. W.-L. Wang, and D. E. Baker. 1979. "Cadmium and the food chain: the effect of dietary cadmium on tissue composition in chicks and laying hens." *J. Nutr.* 109:437-443.

- Lepore, P.D., and R.F. Miller, 1965. "Embryonic viability as influenced by excess molybdenum in chicken breeder diets." *Proc. Soc. Exp. Biol. Med.* 118:155-157
- Mackenzie, K.M. and D.M. Angevine. 1981. "Infertility in mice exposed in utero to benzo[a]pyrene." *Biol. Reprod.* 24:183-191.
- Mackenzie, R.D., R.U. Byerrum, C.F. Decker, C.A. Hoppert, and R.F. Langham. 1958. "Chronic toxicity studies, II. Hexavalent and trivalent chromium administered in drinking water to rats." *Am. Med. Assoc. Arch. Ind. Health.* 18:232-234.
- Marathe, M.R., and G.P. Thomas. 1986. "Embryotoxicity and teratogenicity of lithium carbonate in Wistar rat." *Toxicol. Lett.* 34:115-120.
- Marchinton, R.L., and D.H. Hirth. 1984. "Behavior." pp. 129-168. In L. K. Halls (ed.) *White-tailed Deer: Ecology and Management*. Stackpole Books, Harrisburg, Penn.
- Mautz, W.W., H. Silver, J.B. Holter, H.H. Hayes, and W.E. Urban. 1976. "Digestibility and related nutritional data for seven northern deer browse species." *J. Wildl. Manage.* 40(4):630-638.
- Mehring, A.L., Jr., J.H. Brumbaugh, A.J. Sutherland, and H.W. Titus. 1960. "The tolerance of growing chickens for dietary copper." *Poult. Sci.* 39:713-719.
- Miller, D.H., and L.L. Getz. 1977. "Factors influencing local distribution and species diversity of forest small mammals in New England." *Can. J. Zool.* 55: 806-814.
- Mineau, P., B.T. Collins, and A. Baril. 1996. "On the use of scaling factors to improve interspecies extrapolation of acute toxicity in birds." *Regulatory Toxicology and Pharmacology*. In press.
- Mitchell, J.L. 1961. "Mink movements and population on a Montana river." *J. Wildl. Manage.* 25:48-54.
- NCA (National Coffee Association). 1982. *24-month chronic toxicity and oncogenicity study of methylene chloride in rats*. Final report. Hazelton Laboratories, Inc., Vienna, Va.
- NCRP (National Council on Radiation Protection and Measurements). 1987. *Recommendations on Limits for Exposure to Ionizing Radiation*. NCRP Report No. 91, National Council on Radiation Protection and Measurements, Bethesda, Md.
- NCRP (National Council on Radiation Protection and Measurements). 1989. *Screening Techniques for Determining Compliance with Environmental Standards: Releases of Radionuclides to the Atmosphere*. NCRP Commentary No. 3.
- National Geographic Society. 1987. *Field Guide to the Birds of North America*. 2nd Edition.
- Nawrot, P.S., and R.E. Staples. 1979. "Embryofetal toxicity and teratogenicity of benzene and toluene in the mouse." *Teratology.* 19:41A.
- NCRP (National Council on Radiation Protection and Measurements). 1991. *Effects of Ionizing Radiation on Aquatic Organisms*. NCRP Report No 109. National Council on Radiation Protection and Measurements, Bethesda, Md.

- Nicholson, J. K., and D. Osborn. 1983. "Kidney lesions in pelagic seabirds with high tissue levels of cadmium and mercury." *J. Zool. (Lond.)* 200:99-118.
- Ondreicka, R., E. Ginter, and J. Kortus. 1966. "Chronic toxicity of aluminum in rats and mice and its effects on phosphorus metabolism." *Brit. J. Indust. Med.* 23:305-313.
- Opresko, D.M., B.E. Sample, and G.W. Suter, II. 1995. *Toxicological Benchmarks for Wildlife: 1995 Revision*. ES/ER/TM-86/R2. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Oswald, C., P.Fonken, D. Atkinson, and M. Palladino. 1993. "Lactational water balance and recycling in white-footed mice, red-backed voles, and gerbils." *J. Mammal.* 74:963-970.
- Palmer, A.K., A.E. Street, F.J.C. Roe, A.N. Worden, and N.J. Van Abbe. 1979. "Safety evaluation of toothpaste containing chloroform, II. Long term studies in rats." *J. Environ. Pathol. Toxicol.* 2:821-833.
- Pattee, O.H. 1984. "Eggshell thickness and reproduction in American kestrels exposed to chronic dietary lead." *Arch Environ. Contam. Toxicol.* 13:29-34.
- Peakall, D.B. 1974. "Effects of di-N-butylphthalate and di-2-ethylhexylphthalate on the eggs of ring doves." *Bull. Environ. Contam. Toxicol.* 12:698-702.
- Perry, H.M., E.F. Perry, M.N. Erlanger, and S.J. Kopp. 1983. "Cardiovascular effects of chronic barium ingestion." In Proceedings of the 17th Annual Conference on Trace Substances in Environmental Health, Vol. 17. U. of Missouri Press, Columbia, Mo..
- Peterson, M. A., R. R. Petri, G. R. Southworth. 1994. "Bioaccumulation studies." In T. L. Ashwood (ed.), *Eighth Annual Report on the ORNL Biological Monitoring and Abatement Program*. Draft ORNL/TM-12767. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Sample, B.E., L.A. Baron, and B.L. Jackson. 1995. *Preliminary Assessment of the ecological risks to wide-ranging wildlife species on the Oak Ridge Reservation*. DOE/OR/01-1407&D1. Oak Ridge, Tenn.
- Sargeant, A.B. 1972. "Red fox spatial characteristics in relation to waterfowl predation." *J. Wildl. Manage.* 36:225-236.
- Sargeant, A.B. 1978. "Red fox prey demands and implications to prairie duck production." *J. Wildl. Manage.* 42(3):520-527.
- Schlatterer, B., T.M.M. Coenen, E. Ebert, R. Grau, V. Hilbig, and R. Munk. 1993. "Effects of Bis(tri-*n*-butyltin)oxide in Japanese quail exposed during egg laying period: an interlaboratory comparison study." *Arch. Environ. Contam. Toxicol.* 24:440-448.
- Schlesinger, W.H. and G.L. Potter. 1974. "Lead, copper, and cadmium concentrations in small mammals in the Hubbard Brook experimental forest." *Oikos.* 25:148-152.
- Schlicker, S.A., and D.H. Cox. 1968. "Maternal dietary zinc and development; zinc, iron, and copper content of the rat fetus." *J. Nutr.* 95:287-294.

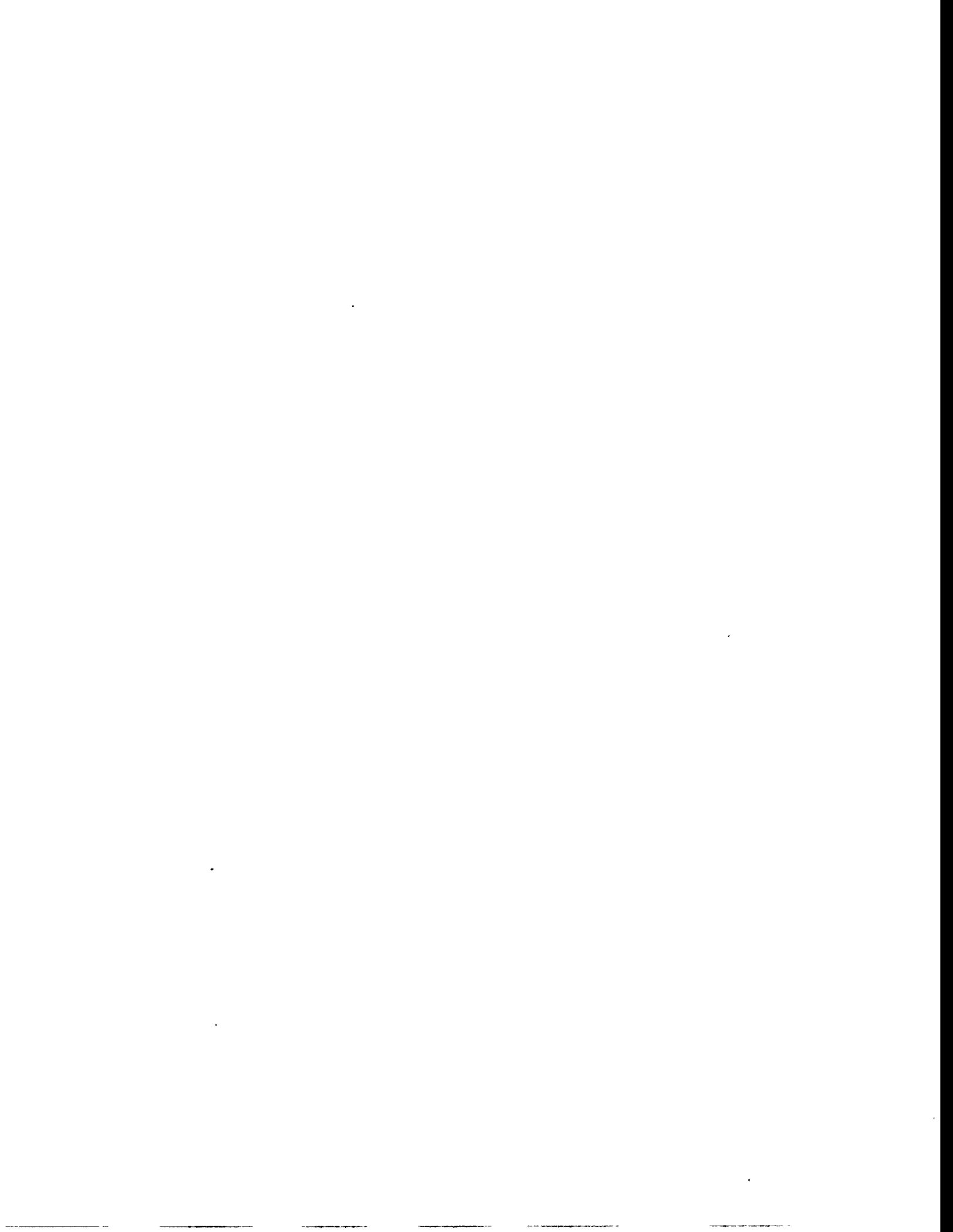
- Schroeder, H.A., M. Mitchener, J.J. Balassa, M. Kanisawa, and A.P. Nason. 1968. "Zirconium, niobium, antimony, and fluorine in mice: effects on growth, survival and tissue levels." *J. Nutr.* 95:95-101.
- Schroeder, H. A., and M. Mitchener. 1971. "Toxic effects on the reproduction of mice and rats." *Arch. Environ. Health* 23:102-106.
- Schroeder, H. A., and M. Mitchener. 1975. "Life term studies in rats: effects of aluminum, barium, beryllium, and tungsten." *J. Nutr.* 105:421-427.
- Sealander, J.A. 1943. "Winter food habits of mink in southern Michigan." *J. Wildl. Manage.* 7: 411-417.
- Skoryna, S.C. 1981. "Effects of oral supplementation with stable strontium." *Can. Med. Assoc. J.* 125:703-712.
- Smith W.P. 1991. "*Odocoileus virginianus*." *Mammalian Species.* 388:1-13.
- Smith, G.J., and V.P. Anders. 1989. "Toxic effects of boron on mallard reproduction." *Environ. Toxicol. Chem.* 8:943-950.
- Stahl, J.L., J.L. Greger, and M.E. Cook. 1990. "Breeding-hen and progeny performance when hens are fed excessive dietary zinc." *Poult. Sci.* 69: 259-263.
- Steven, J.D., L.J. Davies, E.K. Stanley, R.A. Abbott, M. Ihnat, L. Bidstrup, and J.F. Jaworski. 1976. "Effects of chromium in the Canadian environment." NRCC No. 151017.
- Stevens, R.T., 1995. Heavy Metals and PCBs in Mink (*Mustela vison*) and Muskrat (*Ondatra zibethcus*) from the U.S. Department of Energy's Oak Ridge Reservation. unpubl. Masters Thesis. Tennessee Technological University. Cookeville, Tenn.
- Storer, N.L., and T.S. Nelson. 1968. "The effect of various aluminum compounds on chick performance." *Poult. Sci.* 47:244-247.
- Storm, G.L., R.D. Andrews, R. L. Phillips, R.A. Bishop, D.B. Siniff, and J.R. Tester. 1976. "Morphology, reproduction, dispersal, and mortality of midwestern red fox populations." *Wildl. Monogr.*
- Suter, G. W., B. E. Sample, D. S. Jones, and T. L. Ashwood. 1995. *Approach and Strategy for Performing Ecological Risk Assessments for the U.S. Department of Energy's Oak Ridge Reservation: 1995 Revision.* ES/ER/TM-33/R2, Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Talmage, S.S., and B.T. Walton. 1993. "Food chain transfer and potential renal toxicity to small mammals at a contaminated terrestrial field site." *Ecotoxicology.* 2:243-256
- Tewe, O.O., and J.H. Maner. 1981 "Long-term and carry-over effect of dietary inorganic cyanide (KCN) in the life cycle performance and metabolism of rats." *Toxicol. Appl. Pharmacol.* 58:1-7.

- Thompson, D. R. 1996. "Mercury in birds and terrestrial mammals." pp. 341-356. In W. N. Beyer, G. H. Heinz, and A. W. Redmon-Norwood (eds.), *Environmental Contaminants in Wildlife*. Lewis Publishers, New York.
- Torgerson, O., and W.R. Porath. 1984. "Midwest oak/hickory forest." pp. 411-426. In *White-Tailed Deer: Ecology and Management*.
- U.S. Fish and Wildlife Service. 1964. "Pesticide-wildlife studies, 1963: a review of Fish and Wildlife Service investigations during the calendar year." FWS Circular 199.
- U.S. Fish and Wildlife Service. 1969. "Bureau of sport fisheries and wildlife." Publication 74.
- van Gestel, C. A. M. 1992. "The influence of soil characteristics on the toxicity of chemicals for earthworms: a review." pp. 44-54. In *Ecotoxicology of Earthworms*. P. W. Grieg-Smith et al. (eds). Intercept Ltd.
- van Zyll de Jong, C.G. 1983. *Handbook of Canadian Mammals*. National Museums of Canada.
- Verschuuren, H.G., R. Kroes, E.M. Den Tonkelaar, J.M. Berkvens, P.W. Helleman, A.G. Rauws, P.L. Schuller, G.J. Van Esch. 1976c. "Toxicity of methylmercury in rats I. reproduction study." *Toxicology* 6:97-106.
- Vogtsberger, L.M. and G.W. Barrett. 1973. "Bioenergetics of captive red foxes." *J. Wildl. Manage.* 37(4): 495-500.
- Washington-Allen, R.A., T.L. Ashwood, S.W. Christensen. 1995. *Terrestrial Mapping of the Oak Ridge Reservation: Phase I*. ES/ER/TM-152. Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Weir, R.J., and R.S. Fisher. 1972. "Toxicological studies on borax and boric acid." *Toxicol. Appl. Pharmacol.* 23:351-364.
- Whitaker, J.O., Jr., and M.G. Ferraro. 1963. "Summer food of 220 short-tailed shrews from Ithica, New York." *J. Mamm.* 44:419.
- Whitaker, J.O., Jr. 1966. "Food of *Mus musculus*, *Peromyscus maniculatus bairdi*, and *Peromyscus leucopus* in Vigo County, Indiana." *J. Mamm.* 47:473-486.
- White, D.H. and M.P. Dieter. 1978. "Effects of dietary vanadium in mallard ducks." *J. Toxicol. Environ. Health.* 4:43-50.
- White, D.H., and M.T. Finley. 1978. "Uptake and retention of dietary cadmium in mallard ducks." *Environ. Res.* 17:53-59.
- Will, M. E., and G. W. Suter, II. 1995a. *Toxicological Benchmarks for Potential Contaminants of Concern for Effects on Soil and Litter Invertebrates and Heterotrophic Process*. ES/ER/TM-126/R2, Oak Ridge National Laboratory, Oak Ridge, Tenn.
- Will, M. E., and G. W. Suter, II. 1995b. *Toxicological Benchmarks for Potential Contaminants of Concern for Effects on Terrestrial Plants*. ES/ER/TM-85/R2, Oak Ridge National Laboratory, Oak Ridge, Tenn..

- Wills, J.H., G.E. Groblewski, and F. Coulston. 1981. "Chronic and multigeneration toxicities of small concentrations of cadmium in the diet rats." *Ecotoxicol. Environ. Safety*. 5:452-464.
- Wobeser, G., N. O. Nielsen, and B. Schiefer. 1976. "Mercury and mink. II. Experimental methylmercury intoxication." *Can. J. Comp. Med.* 40:34-45.
- Wolff, J.O., R.D. Dueser, and K. S. Berry. 1985. "Food habits of sympatric *Peromyscus leucopus* and *Peromyscus maniculatus*" *J. Mamm.* 66:795-798.
- Wolff, J.O. 1985. "The effects of density, food, and interspecific interference on home range size in *Peromyscus leucopus* and *Peromyscus maniculatus*." *Can. J. Zool.* 63:2657-2662.

Appendix A

**BENTHIC MACROINVERTEBRATE COMMUNITY STUDY
OF WASTE AREA GROUPING 2 DEPOSITION AREAS**



**BENTHIC MACROINVERTEBRATE COMMUNITY STUDY
OF WASTE AREA GROUPING 2 DEPOSITION AREAS**

November 17, 1995

Mark R. Smith and John G. Smith
Biological Monitoring and Abatement Program
Environmental Sciences Division
Oak Ridge National Laboratory
Jaycor

1. INTRODUCTION

Benthic macroinvertebrates are organisms visible to the unaided eye that inhabit the bottom substrates of aquatic systems, and include organisms such as insects, mollusks, crustaceans, annelids, bryozoans, and planarians (Platts et al. 1983). Because these organisms live in close association with the substrate and have relatively long life spans, analyses of benthic macroinvertebrate community composition can aid in assessing the role of sediment contaminants in overall system "health." Benthic macroinvertebrates have been collected from riffle areas in streams of the White Oak Creek (WOC) watershed since 1986 for the Biological Monitoring and Abatement Program (BMAP) at Oak Ridge National Laboratory (ORNL). However, deposition areas (e.g., pools, White Oak Lake) have not been sampled. It is in these areas that contaminants entering the watershed may accumulate, and therefore have a greater impact.

The objectives of this study were to evaluate the ecological condition of the benthic macroinvertebrate communities in major areas of sediment deposition in streams of the WOC watershed in Waste Area Grouping (WAG) 2, and to evaluate whether significant ecological damage would result from maintenance operations.

This study focused on major deposition areas in WOC watershed including White Oak Lake (WOL) and two weirs each on WOC and Melton Branch (MEB).

2. DESCRIPTION OF STUDY AREAS

Samples were collected from six sites within WAG 2 (Fig. 1) and four sites outside WAG 2 that served as references (Figs. 1 and 2). Sampling sites in WAG 2 included two each in WOL, WOC, and MEB. In WOL, the sampling sites were located approximately 100 m and 300 m upstream of WOL Dam, and were referred to as WOLLS and WOLUS, respectively. In WOC, one site was located at the X-14 weir just upstream of the confluence with MEB, and the other was located at the weir just downstream of Melton Valley Drive; these sites were referred to as WOCLS and WOCUS, respectively. In MEB, one site was located in the deposition area just upstream of the X-13 weir (MEBLS), and the other was located approximately 200 m downstream of the HFIR discharge tributary (MEBUS).

Artificial impoundments were selected as reference sites based on similarities in physical structure (substrate type, water depth, and width) (Table 1). Reference sites consisted of two locations within the main ORNL complex on streams just north of Bethel Valley Road; First Creek and WOC (Fig. 1). The two other reference sites were located on Clear Creek, located just southeast of Norris Dam in Norris, Tennessee (Fig. 2). The First Creek site, located ~ 28 m upstream of Bethel Valley Road, was a small impoundment behind a spill dam; this site was referred to as First Creek Pool (FCP). A weir on WOC (WCK68), located approximately 6.8 kilometers above the mouth of the stream was also used as a reference site. This site was located just downstream of a reference site used in the ORNL BMAP. The two remaining reference sites (CCKLD and CCKUD) were above spill dams on Clear Creek (Fig. 2), a relatively undisturbed stream with little development from its headwaters to the mouth of the stream.

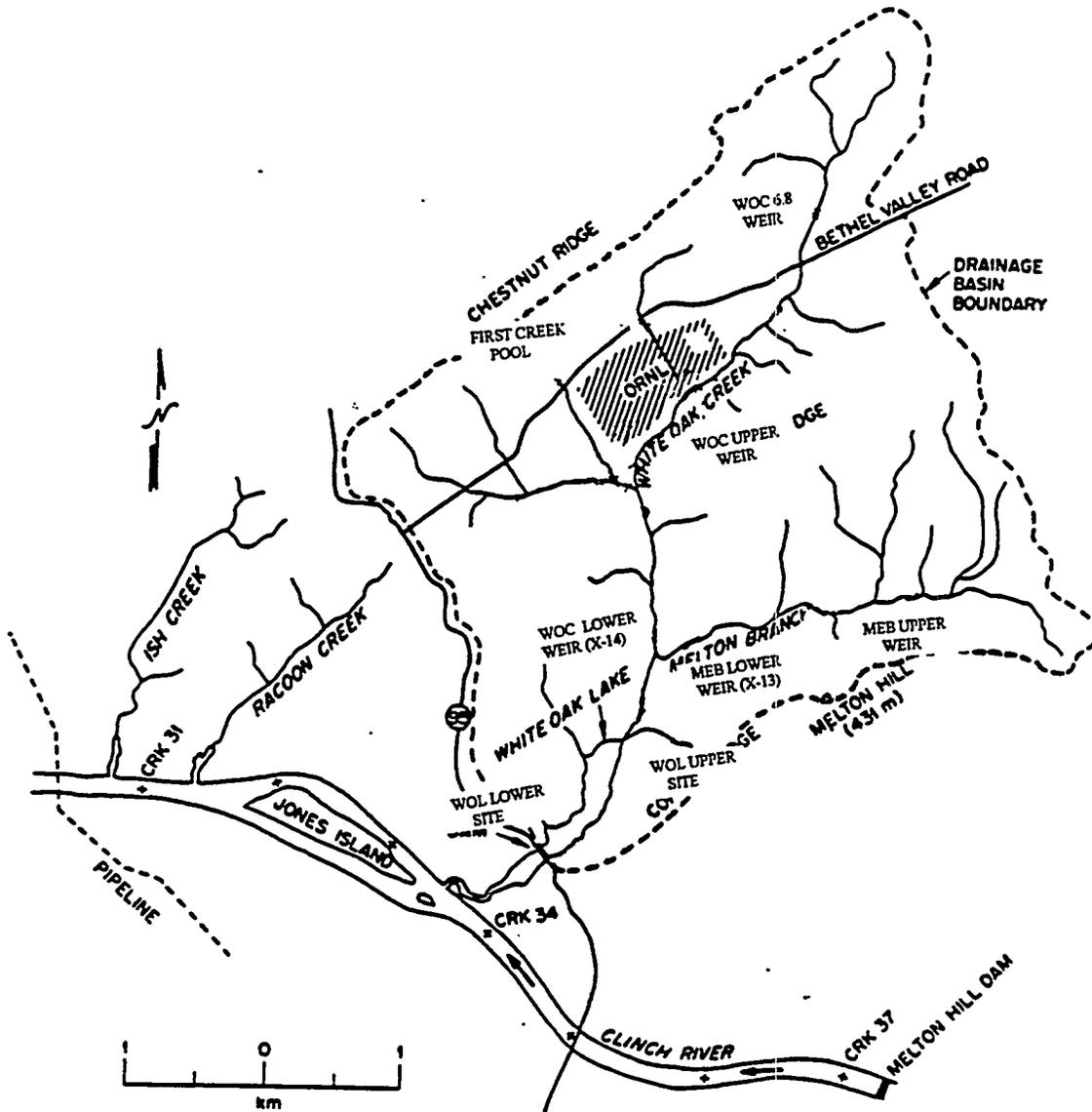


Fig. A.1. Location of benthic macroinvertebrate core sampling sites on the Oak Ridge Reservation.

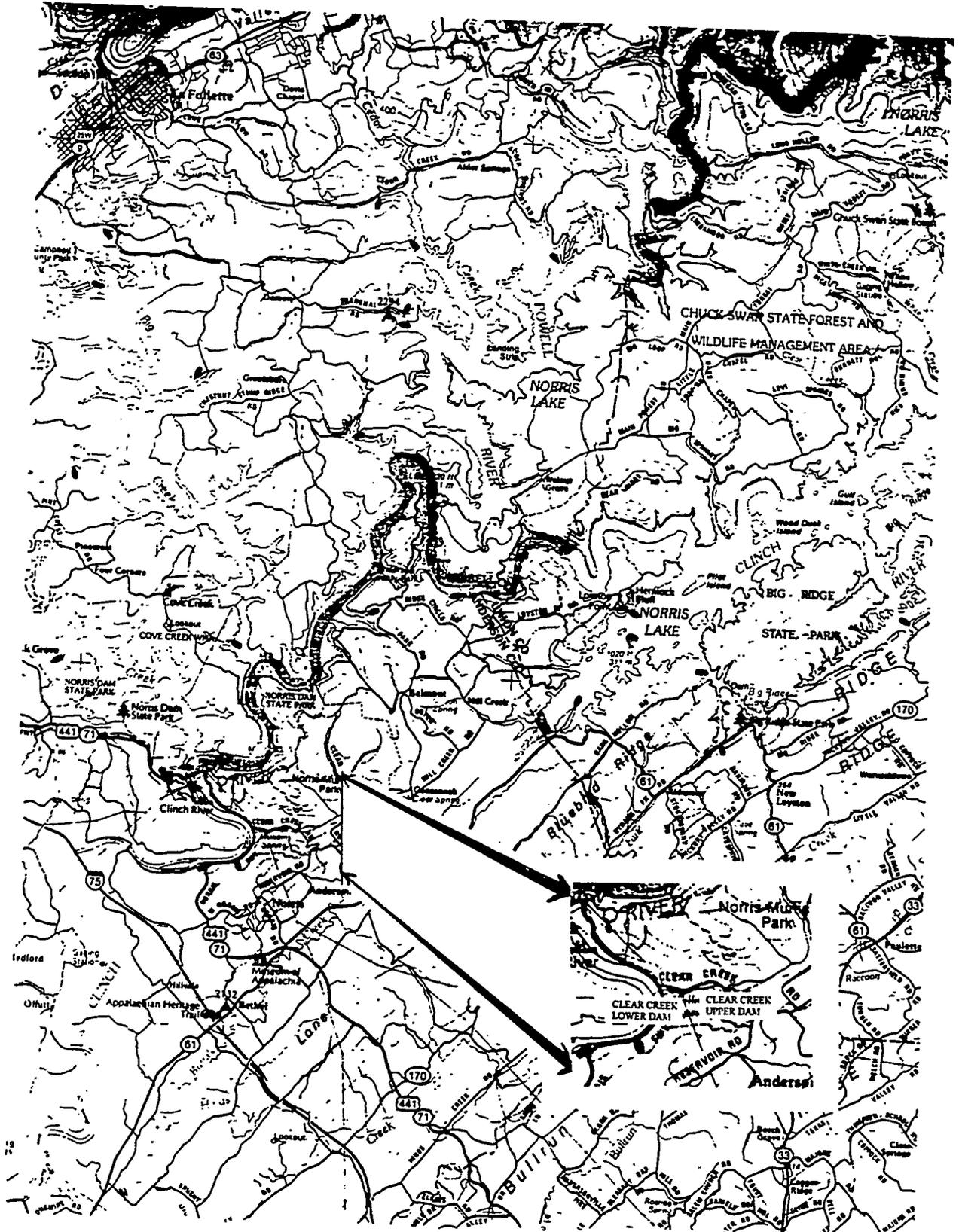


Fig. A.2. Location of reference benthic macroinvertebrate core sampling sites on Clear Creek, Anderson County, Tennessee.

Table A.1. Summary of physical characteristics of the WAG 2 study sites and reference sites (June 1995)

Site	Average depth (cm)	Substrate type	Transect width (m)
WAG 2 Sites			
WOC ^a lower weir (WOCLS)	18.2	Silt	16.0
WOC ^a upper weir (WOCUS)	36.6	Silt	14.0
Melton Branch lower weir (MEBLS)	51.8	Silt	6.0
Melton Branch upper weir (MEBUS)	41.4	Gravel/sand/silt	4.3
WOL ^b lower site (WOLLS)	100.2	Silt	170.0
WOL ^b upper site (WOLUS)	41.4	Silt	180.0
Reference sites			
First Creek pool (FCP)	32.2	Silt/detritus	19.0
WOC ^a 6.8 weir (WCK68)	22.2	Gravel/sand/silt	4.0
Clear Creek lower dam (CCKLD)	78.8	Gravel/sand/silt	19.0
Clear Creek upper dam (CCKUD)	74.6	Silt/detritus	20.2

^aWOC= White Oak Creek; ^bWOL= White Oak Lake.

3. MATERIALS AND METHODS

3.1 SAMPLE COLLECTION

Before collection of invertebrate samples, various chemical and physical aspects (D.O., conductivity, pH, temperature, site width, and water depth) were measured at each site, in accordance with Standard Operating Procedures (SOP-3) of the BMAP Benthic Macroinvertebrate Monitoring Project Quality Assurance Plan (Q.P.) (Smith 1992). Five replicate macroinvertebrate samples were collected from each site with a hand-held coring device. The coring device had an internal diameter of 5 cm, and was inserted into the substrate to a depth of 20.5 cm. An aluminum skirt (27 x 27 cm) attached to the corer ensured consistent sampling depths. Following procedures outlined in section 7, SOP-5 of the Benthic Macroinvertebrate Q.P., sample replicates were taken at evenly spaced intervals along a transect running perpendicular to the shoreline at each site. After the corer was extracted from the substrate, the contents were emptied into a Surber net (363- μ m mesh), and substrate composition was determined by visual inspection, using methods described in SOP-2.

After identifying substrate composition, the Surber net containing the sample was lowered into the water and gently agitated to flush out the fine sediments. The contents of the net were then transferred into an appropriately labeled sample jar, and preserved in 80% ethanol. Prior to processing, samples were stored and maintained in a secure storage facility at ORNL, in accordance with approved quality assurance procedures (SOP-9, Smith 1992).

3.2 LABORATORY PROCEDURES

Appropriate procedures outlined in the BMAP Benthic Macroinvertebrate Monitoring Project Sample Processing QA Plan (Wojtowicz and Smith 1992) were followed for processing samples. Briefly, sample processing consisted of washing each sample in a 250 μ m sieve and sorting the remaining organisms to the lowest practical taxon. Chironomidae (midge) larvae were mounted on microscope slides for identification under compound microscopes, while the remaining organisms were identified using stereo microscopes. A reference collection was created containing two or three specimens of each taxon encountered.

The procedure normally followed for evaluating sorting efficiency (SOP- 4 in Wojtowicz and Smith 1992) was modified for processing WAG 2 samples because it was felt that the modified procedure was more rigorous. In the original procedure, half the sample material sorted by an individual is randomly selected for resorting by a different individual. If the prescribed sorting efficiency of 80% is not met, all of the future material sorted by the individual that failed is checked until that individual can consistently achieve the prescribed efficiency. In the procedure used for WAG 2 samples, if any of an individual's randomly checked samples (i.e., of the 10% checked) do not meet the prescribed sorting efficiency of 80%, then all of that individuals sorted material is resorted. If an individual always sorts samples at or above the prescribed efficiency level, sorting proceeds more rapidly. The results of sorting efficiency quality control checks for the WAG 2 samples are given in WAG 2 project files in a summary report provided by the laboratory that processed the samples. No sorting failures were reported for the WAG 2 samples.

3.3 DATA ANALYSIS

Data analysis was performed with the aid of Statistical Analysis System software and procedures (SAS 1985a, 1985b). Statistical analyses consisted of both descriptive and parametric techniques. Descriptive techniques included determinations of mean values for density, total richness, EPT richness (the total number of Ephemeroptera, Plecoptera, and Trichoptera taxa), and Chironomidae taxa richness for each site. A one-way analysis of the variance (ANOVA) with site as the main effect was performed on density, total richness, and Chironomidae taxa richness. Site differences were separated with a Tukey's Test, with the level of significance set as $p \leq 0.05$. Because major differences in substrate were observed among some sites while samples were collected (Table 1), the analysis was repeated on two groups based on substrate type (i.e., silt and gravel). However, this analysis gave no indication that substrate was a major factor contributing to any differences. Therefore, no further detailed consideration was given to this factor in the results. Prior to performing the ANOVAs, values for each response variable were appropriately transformed (i.e. $\log_{10}(X+1)$ for density values, and square root of X for total and Chironomidae taxa richness values, where X = the individual values for density, taxonomic richness, and Chironomidae richness; Elliot 1977).

Because Ephemeroptera, Plecoptera, and Trichoptera (EPT) taxa are generally considered sensitive to environmental disturbance (Lenat 1988), BMAP benthic macroinvertebrate studies effectively use this metric to evaluate the biological condition of stream sites (e.g., Smith 1993). However, due to the very low numbers encountered at all sites (no site had a mean EPT richness above two), it was felt this metric would not be useful for characterizing WAG 2 sites or effectively separating any site differences, and therefore not considered further in the analysis.

4. DATA ASSESSMENT

The sampling and analysis plan developed and approved by project management for the benthic macroinvertebrate task of the WAG 2 Remedial Investigation (RI) resulted from issues and data deficiencies identified through the data quality objectives (DQO) process (Watkins and Herbes 1994). All goals established in the DQO process for this task were met.

4.1 SAMPLE CHAIN-OF-CUSTODY

Sample chain-of-custody was maintained and documented from collection through their final disposition in a secure area at ORNL after processing. Copies of the sample chain-of-custody sheets are in WAG 2 project files.

4.2 COMPLETENESS

Having developed the most extensive and suitable benthic macroinvertebrate sampling plan possible within budget constraints, anything less than 100% completion of the plan outlined in the RI Work Plan (Watkins and Herbes 1994) was considered unacceptable. All 50 samples collected for the project were processed, and the results from these samples are included in this report.

4.3 PRECISION

An acceptable sampling precision for benthic macroinvertebrate studies is often controlled by budget constraints (Resh and McElravy 1993). A high level of precision (i.e., $< \pm 20\%$ of the mean for benthic macroinvertebrate samples) can require an astronomical number of sampling units (e.g., > 100 sampling units per site). It has been found however, that estimates of $\pm 40\%$ of the true mean can generally be achieved with six sampling units (Resh and McElravy 1993). Thus, depending upon sample variation, five sample units as used in this study should allow differences in site means of two to three fold to be detected.

In the laboratory, no sorting efficiency failures occurred in the 10% of samples (i.e., 5 of 50 samples) subjected to quality control checks (see summary report in WAG 2 project files).

4.4 DATA REVIEW

The final data set was obtained in electronic form (Lotus_®) and in hard copy from the laboratory that processed the samples. A copy of the data as submitted by the laboratory is provided in WAG 2 project files, as part of their final report submitted to the benthic macroinvertebrate task PI. The accuracy of the data was initially validated by comparing an output of the data from the electronic copy with the copy provided in the subcontractor's report. Procedures available with the Statistical Analysis System (SAS 1985a; SAS 1985b) allowed the data to be subjected to various sorts (i.e., rearrangements) to identify inconsistencies and misspellings. Inconsistencies and misspellings were corrected, and appropriate changes were made to make the data set compatible with other data collected by the task Principal Investigator since 1984 from other projects associated with the Department of Energy's Oak Ridge Operations; these changes had no affect on data accuracy or precision. Some observations were eliminated because they included taxa not considered benthic. The observations eliminated included those with the invertebrate taxa Corixidae, Entomobryidae, and Gerridae. These three taxonomic groups live on the surface of the water or in the water column and are sometimes collected accidentally with benthic sampling devices. A copy of the final data set used to conduct all analyses is provided in WAG 2 project files.

5. RESULTS

5.1 TAXONOMIC COMPOSITION

A checklist of benthic macroinvertebrate taxa collected at each site is presented in WAG 2 project files. There were a total of 105 taxa identified, with 60 of these being in the Chironomidae (Diptera) family. Taxonomic composition varied with site, with only one major taxonomic group being present at all sites (Oligochaeta). Although most taxa identified to the genus level were collected from less than 50% of the sites, *Dubiraphia* sp. (Elmidae: Coleoptera), and five Chironomidae genera (*Chironomus*, *Cryptotendipes*, *Cryptochironomus*, *Stictochironomus*, and *Tanytarsus*) were present in at least 60% of the sites.

Plecoptera were collected from three sites only, all of which were reference sites (CCKUD, FCP, and WCK68). Conversely, Megaloptera were collected only at three study sites (WOCLS, WOLLS, and MEBUS).

5.2 ABUNDANCE

5.2.1 Total density

Mean densities (number of organisms/0.1 m²) for each site are presented in Fig. 3. Statistically significant differences in densities were not detected among any sites (Tables 2 and 3). Although large differences in densities appeared to exist among some of these sites (e.g., WOCLS had 954.8 organisms/0.1 m², compared to 132.3 organisms/0.1 m² at WOLUS), high variation among samples may have limited the ability of the test to detect statistically significant differences.

5.2.2 Relative abundance

Oligochaetes and chironomids were the predominant taxa at all sites, and except for FCP, WOCLS, and WOCUS, the Chironomidae accounted for over 60% of the total density (Fig. 4). Oligochaetes and chironomids co-dominated at FCP and both WOC sites. The relative abundance of the other taxa, which included Coleoptera (beetles), Odonata (dragonflies), Gastropods (snails), Ephemeroptera (mayflies), Plecoptera (stoneflies), and Trichoptera (caddisflies), ranged from 1.54% at WOLUS to 27.6% at FCP.

Because of the predominance of the Chironomidae at most sites (i.e., >35% of total density at all sites), a closer examination was made of the relative abundances of the major chironomid taxa to further characterize the study sites. Three of the four major taxonomic groups examined comprised > 5% of the total community density at five of the ten sites (Fig. 4). At the extremes in relative abundance, only two of the taxonomic groups exceeded 5% of the total community density at MEBLS, WOLUS, and FCP, while at CCKUD and WCK68, all four major taxonomic groups exceeded 5% of the total density. There were no distinct or consistent differences between the reference sites and the WAG 2 study sites in predominant taxa which made the WAG 2 sites uniquely different from the reference sites. For example, the Chironomini comprised half or more of the chironomid composition at the two Melton Branch sites, CCKUD, and WCK68, while at the two WOL sites and FCP, the Tanypodinae comprised over half of the total chironomid density.

5.3 RICHNESS

5.3.1 Total richness

The reference sites WCK68 and CCKUD had significantly higher total richness values than both WOL sites, while WOCUS had significantly higher richness values than WOLUS (Tables 2 and 3). The WOL sites did not have significantly lower richness values than the remaining sites, including the reference sites FCP and CCKLD.

5.3.2 Chironomidae taxa richness

Chironomidae taxon represented 60 of the 105 taxa identified. Statistical analyses were performed on four major taxa of Chironomidae (Tanypodinae, Tanytarsini, Orthoclaadiinae, and Chironomini) to ascertain if differences may have existed between any of the sites investigated. The pattern of differences among the sites in the richness of the Chironomidae closely resembled the pattern exhibited by total richness (Fig. 3). Chironomid richness was highest at CCKUD and WCK68, and lowest at the two WOL sites, with about a 3-fold difference existing between these high and low

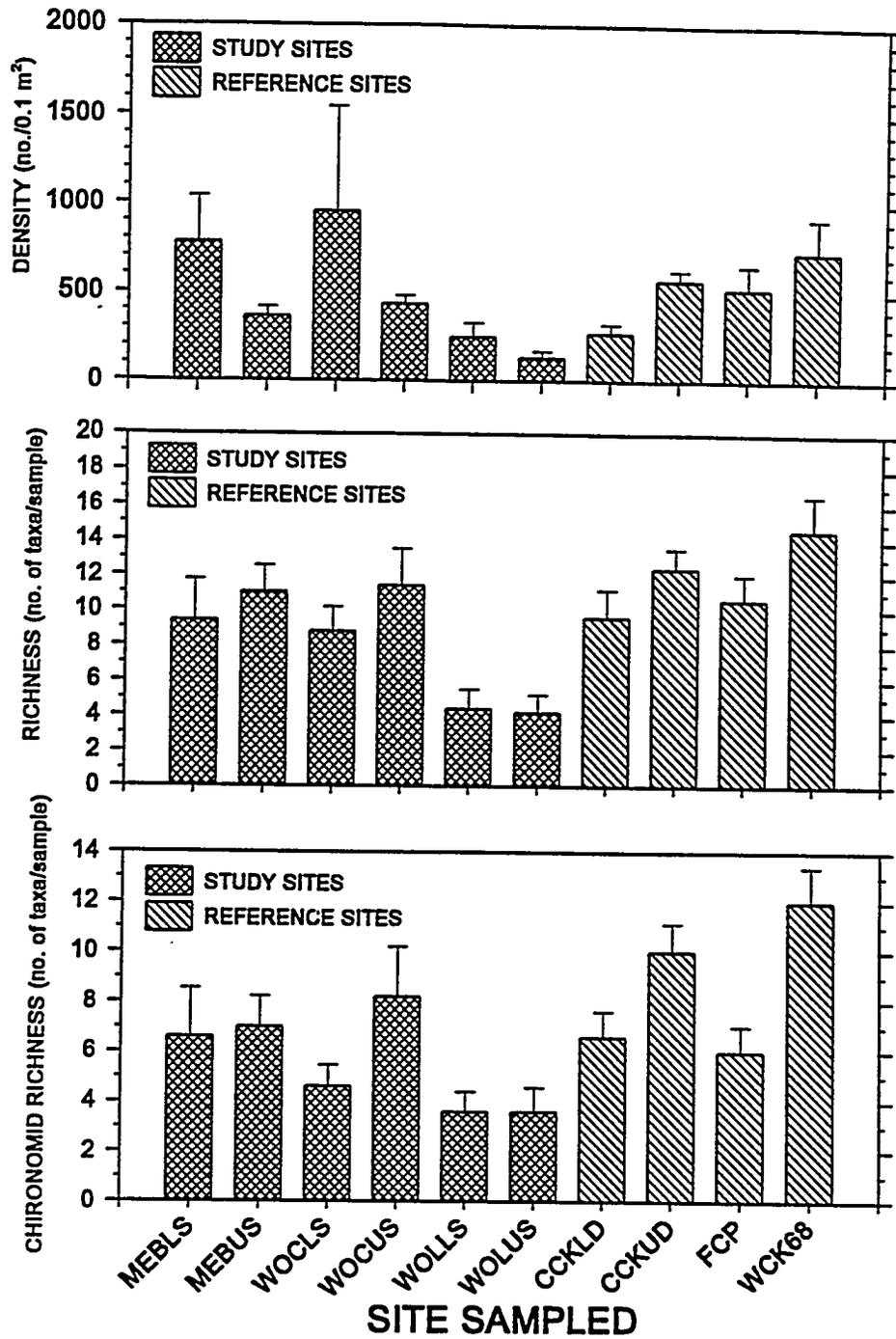


Fig. A.3. Mean total density, mean total richness, and mean richness of major Chironomidae taxa of the benthic macroinvertebrate communities in WAG 2 study sites and reference sites (June 1995).

Table A.2. F-values and *p*-values for one-way analysis of variance (ANOVA) using the variables density, total richness, and Chironomidae taxa richness for benthic macroinvertebrate communities in WAG 2 and associated reference sites (June 1995)

Variable ^a	<i>f</i> -value	<i>p</i> -value
Density	1.15	0.3499
Total Richness	4.39	0.0005
Chironomidae Richness	3.63	0.0022

^aDegrees of freedom = 9 and 40 for numerator and denominator respectively.

Table A.3. Results of site comparisons with Tukey test of values for density, total richness, and Chironomidae taxa richness of the benthic macroinvertebrate communities in WAG 2 and associated reference sites, June 1995.^a

DENSITY

ALL

WOCLS MEBLS WCK68 WOLLS WOLUS CCKLD MEBUS FCP WOCUS CCKUD

TOTAL RICHNESS

ALL

WCK68 CCKUD WOCUS MEBUS FCP CCKLD MEBLS WOCLS WOLLS WOLUS

CHIRONOMIDAE RICHNESS

ALL

WCK68 CCKUD WOCUS MEBUS CCKLD MEBLS FCP WOCLS WOLLS WOLUS

^aSites not connected by the same line are significantly different ($p < 0.05$), based on Tukey's Studentized range test (HSD). Sites are arranged in order of highest to lowest values from left to right. $n = 5$.

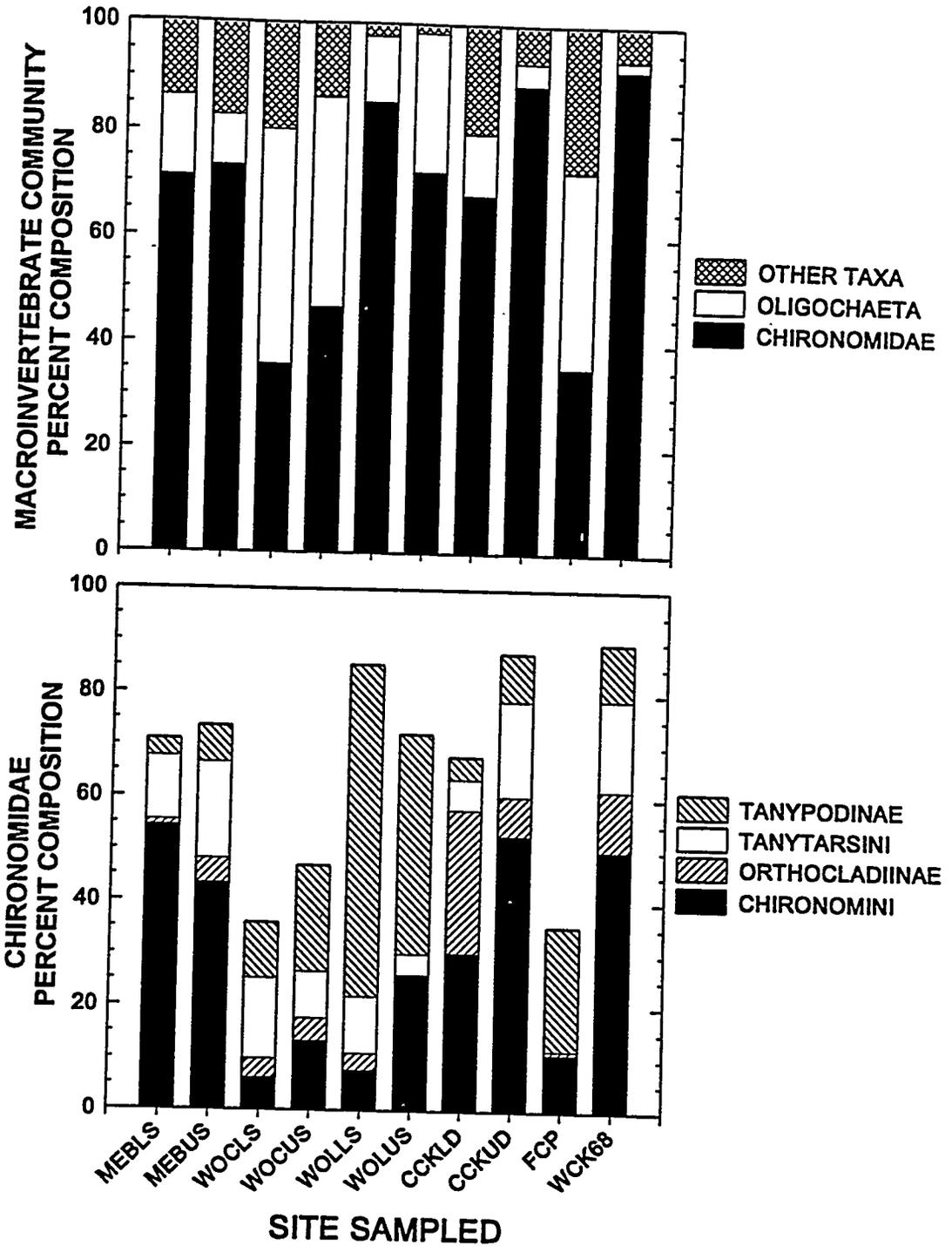


Fig. A.4. Relative abundance of selected benthic macroinvertebrate and of the major Chironomidae taxa at study sites in WAG 2 and associated reference sites, June 1995. The Chironominae were included as a separate taxonomic group because the individuals left in this group could not be further identified; further identification would have placed them within the Tanytarsini or Chironomini.

values. However, only these site differences were statistically significant (Tables 2 and 3). Values for the other six sites were statistically indistinguishable from either those for CCKUD, WCK68, or the two WOL sites. This implies that chironomid richness at all WAG 2 study sites fell within the range that existed at the reference sites used in this study.

6. DISCUSSION

Several uncertainties were apparent within the benthic macroinvertebrate task. The small number of samples taken per site (five) allow only for the detection of large differences (i.e., $\geq 50\%$). This was further complicated by the diverse habitat present at these sites. The resulting variability among sampling units limited the capacity to detect differences among sites. In addition, the single sampling event could not take into account the seasonal variation present in benthic macroinvertebrate communities, thus the possibility of failing to detect important site differences is increased. However, results reported in this study do provide an indication of benthic macroinvertebrate community composition at the various sites when the samples were taken. Identification of substrate composition did not include other substrate factors such as organic content. Because a majority of the organisms collected rely on organic materials in the substrate as a primary food source, organic content of the substrate at the various sites may have also influenced community composition.

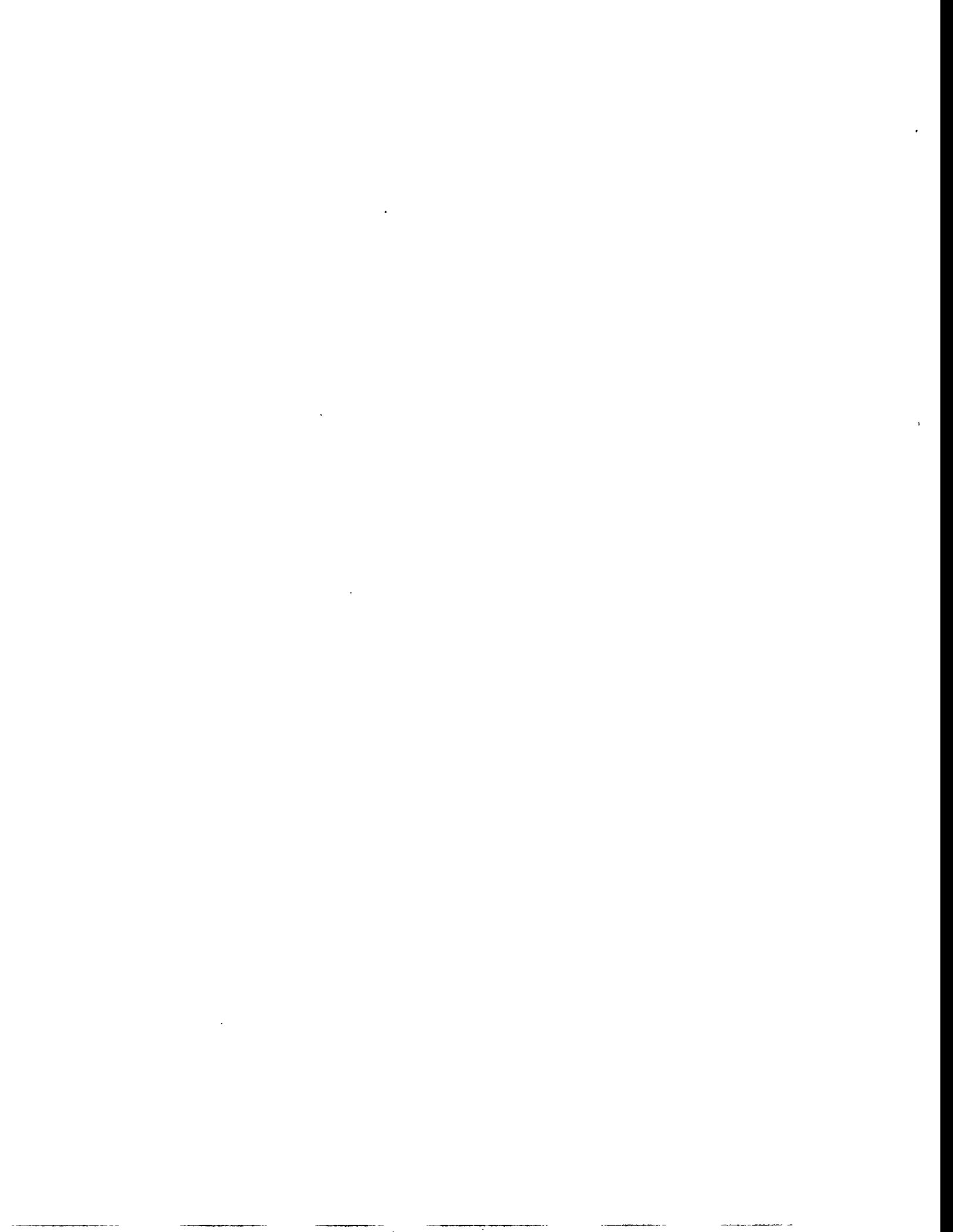
There were no detectable statistically significant differences in densities among the WAG 2 and reference sites, possibly because sites with the highest densities also tended to have high variation among replicates. Relative abundance of the three categories of macroinvertebrates at each site indicated that with the exception of FCP, WOCUS, and WOCLS where oligochaetes co-dominated with the Chironomidae, chironomids were the most numerous organisms regardless of substrate type. This is not unexpected, since oligochaetes and Chironomidae use hemoglobin to aid in the uptake of oxygen, thus enabling them to inhabit areas where low oxygen levels can occur, such as areas of sediment deposition (Hynes 1974; Brinkhurst and Gelder 1991).

The two sites within WOL had the lowest values for all parameters investigated. However, these differences were only statistically significant from those of just two reference sites for total richness and Chironomidae taxa richness. White Oak Lake sites were clearly the most depauperate, although it is not possible to determine whether contaminants, habitat, or other perturbations are responsible for the condition of the benthic macroinvertebrate community. White Oak Lake impounds WOC for a substantial distance upstream of the two sampling sites, whereas at the other sites, impoundments resulting from the weirs were generally less than 50 m long. No reference sites were found of equivalent size, thus comparisons in benthic community composition with reference sites used in this study may not be entirely appropriate. The remaining sites within WAG 2 were generally similar to the reference sites.

The objectives of this study were to evaluate the ecological condition of the benthic macroinvertebrate community in sediment deposition areas of WAG 2, and to determine if significant ecological damage would result from routine maintenance operations. These results suggest that since no major differences existed between the MEB, WOC, and the reference sites, it appears that maintenance scheduled for the four WAG 2 weir sites will not impact any fragile or unique benthic macroinvertebrate communities, although short-term, severe impacts would occur to those communities currently existing. The presence of an apparently depauperate community in WOL also suggests that maintenance scheduled for this body of water should result in no major impacts to either a fragile or unique community.

7. LITERATURE CITED

- Brinkhurst, R. O., and S. R. Gelder. 1991. "Annelida: Oligochaeta and Brachiobdellida." pp 401-435. IN: J.H. Thorp and A. P. Covich (eds.), *Ecology and Classification of North American Freshwater Invertebrates*. Academic Press, Inc. San Diego, California.
- Elliot, J. M. 1977. *Some Methods for the Statistical Analysis of Samples of Benthic Invertebrates*. Scientific Publication No. 25. Freshwater Biological Association, Ambleside, England.
- Hynes, H.B.N. 1974. *The Biology of Polluted Waters*. University of Toronto Press. Toronto, Canada.
- Lenat, D. R. 1988. Water quality assessment of streams using a qualitative collection method for benthic macroinvertebrates. *J. North Am. Benthol. Soc.* 7:222-233.
- Platts, W. W., W. F. Meghan, and G. W. Minshall. 1983. Methods for evaluating stream, riparian, and biotic conditions. U.S. Forest Service General Technical Report INT138. Intermountain Forest and Range Experimental Station, Ogden, UT.
- Resh, V. H., and E. P. McElravy. 1993. "Contemporary Quantitative Approaches to Biomonitoring Using Benthic Macroinvertebrates." pp. 159-194. IN: D. M. Rosenberg and V. H. Resh (eds.), *Freshwater Biomonitoring and Benthic Macroinvertebrates*. Chapman & Hall, New York.
- SAS (SAS Institute, Inc.). 1985a. *SAS User's Guide: Basics, Version 5 Edition*. SAS Institute, Inc., Cary, North Carolina.
- SAS (SAS Institute, Inc.). 1985b. *SAS User's Guide: Statistics, Version 5 Edition*. SAS Institute, Inc., Cary, North Carolina.
- Smith, J. G. 1992. *Biological Monitoring and Abatement Program (BMAP) Benthic Macroinvertebrate Monitoring Project Sample Collection and Storage QA Plan*. QAP-X-90-ES-068. Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Watkins, D. R., and S. E. Herbes (eds.). 1994. *FY 1995 Remedial Investigation Work Plan for Waste Area Grouping 2 at Oak Ridge National Laboratory, Oak Ridge, Tennessee*. DOE/OR/01-1292&D1, Oak Ridge National Laboratory, Oak Ridge, Tennessee.
- Wojtowicz, J. A., and J. G. Smith. 1992. *Biological Monitoring and Abatement Program (BMAP) Benthic Macroinvertebrate Monitoring Project Sample Processing QA Plan*. QAP-X-91-ES-068. Oak Ridge National Laboratory, Oak Ridge, Tennessee.



Appendix B

**DATA TABLES FOR RISKS TO FISH, INVERTEBRATES,
AND PLANTS**



Table B.1. Screening of aqueous concentrations of chemicals detected in streams against screening benchmarks for aquatic life (Suter and Mabrey 1996). All concentrations are mg/L.

Chemical	Reach	Samples	Detects	P.L. Mean	95% UCB	Quotient	Benchmark Value	Benchmark
1,1-Dichloroethane	W4T	3	1	0.0010	0.001	0.0001	14.6800	LCV_ALLO
	W4T	3	1	0.0010	0.001	0.0001	14.6800	LCV_FISH
	W4T	3	1	0.0010	0.001	0.0001	8.2190	LTV_FISH
	W4T	3	1	0.0010	0.001	0.0012	0.8340	S_ACU_V
	W4T	3	1	0.0010	0.001	0.0215	0.0466	S_CHR_V
ALUMINUM	1C	9	7	0.3308	0.503	1.0941	0.4600	LCV_ALLO
	1WC	2	2	0.1485	0.613	1.3317	0.4600	LCV_ALLO
	BWC	2	1	0.1960	0.196	0.4261	0.4600	LCV_ALLO
	HRT	8	5	0.1249	0.178	0.3869	0.4600	LCV_ALLO
	IHP	12	7	0.1666	0.306	0.6643	0.4600	LCV_ALLO
	LMB	29	16	0.1149	0.151	0.3281	0.4600	LCV_ALLO
	LWC	2	1	0.1230	0.123	0.2674	0.4600	LCV_ALLO
	MWC	18	14	0.1122	0.131	0.2851	0.4600	LCV_ALLO
	NWT	2	2	0.1210	0.481	1.0454	0.4600	LCV_ALLO
	RAC	2	2	0.1480	0.843	1.8315	0.4600	LCV_ALLO
	UMB	9	5	0.2312	0.306	0.6648	0.4600	LCV_ALLO
	W4T	16	11	0.2607	0.491	1.0673	0.4600	LCV_ALLO
	WOL	13	13	0.8752	1.102	2.3958	0.4600	LCV_ALLO
	WS	10	8	0.3284	0.455	0.9900	0.4600	LCV_ALLO
	1C	9	7	0.3308	0.503	1.0941	0.4600	LCV_ALLO
	1WC	2	2	0.1485	0.613	1.3317	0.4600	LCV_AQPL
	BWC	2	1	0.1960	0.196	0.4261	0.4600	LCV_AQPL
	HRT	8	5	0.1249	0.178	0.3869	0.4600	LCV_AQPL
	IHP	12	7	0.1666	0.306	0.6643	0.4600	LCV_AQPL
	LMB	29	16	0.1149	0.151	0.3281	0.4600	LCV_AQPL
	LWC	2	1	0.1230	0.123	0.2674	0.4600	LCV_AQPL
	MWC	18	14	0.1122	0.131	0.2851	0.4600	LCV_AQPL
	NWT	2	2	0.1210	0.481	1.0454	0.4600	LCV_AQPL
	RAC	2	2	0.1480	0.843	1.8315	0.4600	LCV_AQPL
	UMB	9	5	0.2312	0.306	0.6648	0.4600	LCV_AQPL
	W4T	16	11	0.2607	0.491	1.0673	0.4600	LCV_AQPL
	WOL	13	13	0.8752	1.102	2.3958	0.4600	LCV_AQPL
	WS	10	8	0.3284	0.455	0.9900	0.4600	LCV_AQPL
	1C	9	7	0.3308	0.503	0.2649	1.9000	LCV_DAPH
	1WC	2	2	0.1485	0.613	0.3224	1.9000	LCV_DAPH
	BWC	2	1	0.1960	0.196	0.1032	1.9000	LCV_DAPH
	HRT	8	5	0.1249	0.178	0.0937	1.9000	LCV_DAPH
	IHP	12	7	0.1666	0.306	0.1608	1.9000	LCV_DAPH
	LMB	29	16	0.1149	0.151	0.0794	1.9000	LCV_DAPH
	LWC	2	1	0.1230	0.123	0.0647	1.9000	LCV_DAPH
	MWC	18	14	0.1122	0.131	0.0690	1.9000	LCV_DAPH
	NWT	2	2	0.1210	0.481	0.2531	1.9000	LCV_DAPH
	RAC	2	2	0.1480	0.843	0.4434	1.9000	LCV_DAPH
	UMB	9	5	0.2312	0.306	0.1609	1.9000	LCV_DAPH
	W4T	16	11	0.2607	0.491	0.2584	1.9000	LCV_DAPH
	WOL	13	13	0.8752	1.102	0.5800	1.9000	LCV_DAPH
	WS	10	8	0.3284	0.455	0.2397	1.9000	LCV_DAPH
	1C	9	7	0.3308	0.503	0.1531	3.2880	LCV_FISH
	1WC	2	2	0.1485	0.613	0.1863	3.2880	LCV_FISH
	BWC	2	1	0.1960	0.196	0.0596	3.2880	LCV_FISH
	HRT	8	5	0.1249	0.178	0.0541	3.2880	LCV_FISH
	IHP	12	7	0.1666	0.306	0.0929	3.2880	LCV_FISH
LMB	29	16	0.1149	0.151	0.0459	3.2880	LCV_FISH	
LWC	2	1	0.1230	0.123	0.0374	3.2880	LCV_FISH	
MWC	18	14	0.1122	0.131	0.0399	3.2880	LCV_FISH	
NWT	2	2	0.1210	0.481	0.1463	3.2880	LCV_FISH	
RAC	2	2	0.1480	0.843	0.2562	3.2880	LCV_FISH	
UMB	9	5	0.2312	0.306	0.0930	3.2880	LCV_FISH	
W4T	16	11	0.2607	0.491	0.1493	3.2880	LCV_FISH	
WOL	13	13	0.8752	1.102	0.3352	3.2880	LCV_FISH	
WS	10	8	0.3284	0.455	0.1385	3.2880	LCV_FISH	
1C	9	7	0.3308	0.503	0.9320	0.5400	LTV_DAPH	
1WC	2	2	0.1485	0.613	1.1344	0.5400	LTV_DAPH	

B-4

BWC	2	1	0.1960	0.196	0.3630	0.5400	LTV_DAPH
HRT	8	5	0.1249	0.178	0.3296	0.5400	LTV_DAPH
IHP	12	7	0.1666	0.306	0.5658	0.5400	LTV_DAPH
LMB	29	16	0.1149	0.151	0.2795	0.5400	LTV_DAPH
LWC	2	1	0.1230	0.123	0.2278	0.5400	LTV_DAPH
MWC	18	14	0.1122	0.131	0.2429	0.5400	LTV_DAPH
NWT	2	2	0.1210	0.481	0.8905	0.5400	LTV_DAPH
RAC	2	2	0.1480	0.843	1.5602	0.5400	LTV_DAPH
UMB	9	5	0.2312	0.306	0.5663	0.5400	LTV_DAPH
W4T	16	11	0.2607	0.491	0.9092	0.5400	LTV_DAPH
WOL	13	13	0.8752	1.102	2.0408	0.5400	LTV_DAPH
WS	10	8	0.3284	0.455	0.8433	0.5400	LTV_DAPH
1C	9	7	0.3308	0.503	0.1071	4.7000	LTV_FISH
1WC	2	2	0.1485	0.613	0.1303	4.7000	LTV_FISH
BWC	2	1	0.1960	0.196	0.0417	4.7000	LTV_FISH
HRT	8	5	0.1249	0.178	0.0379	4.7000	LTV_FISH
IHP	12	7	0.1666	0.306	0.0650	4.7000	LTV_FISH
LMB	29	16	0.1149	0.151	0.0321	4.7000	LTV_FISH
LWC	2	1	0.1230	0.123	0.0262	4.7000	LTV_FISH
MWC	18	14	0.1122	0.131	0.0279	4.7000	LTV_FISH
NWT	2	2	0.1210	0.481	0.1023	4.7000	LTV_FISH
RAC	2	2	0.1480	0.843	0.1793	4.7000	LTV_FISH
UMB	9	5	0.2312	0.306	0.0651	4.7000	LTV_FISH
W4T	16	11	0.2607	0.491	0.1045	4.7000	LTV_FISH
WOL	13	13	0.8752	1.102	0.2345	4.7000	LTV_FISH
WS	10	8	0.3284	0.455	0.0969	4.7000	LTV_FISH
1C	9	7	0.3308	0.503	0.6710	0.7500	NAWQ_ACU
1WC	2	2	0.1485	0.613	0.8167	0.7500	NAWQ_ACU
BWC	2	1	0.1960	0.196	0.2613	0.7500	NAWQ_ACU
HRT	8	5	0.1249	0.178	0.2373	0.7500	NAWQ_ACU
IHP	12	7	0.1666	0.306	0.4074	0.7500	NAWQ_ACU
LMB	29	16	0.1149	0.151	0.2012	0.7500	NAWQ_ACU
LWC	2	1	0.1230	0.123	0.1640	0.7500	NAWQ_ACU
MWC	18	14	0.1122	0.131	0.1749	0.7500	NAWQ_ACU
NWT	2	2	0.1210	0.481	0.6412	0.7500	NAWQ_ACU
RAC	2	2	0.1480	0.843	1.1234	0.7500	NAWQ_ACU
UMB	9	5	0.2312	0.306	0.4077	0.7500	NAWQ_ACU
W4T	16	11	0.2607	0.491	0.6546	0.7500	NAWQ_ACU
WOL	13	13	0.8752	1.102	1.4694	0.7500	NAWQ_ACU
WS	10	8	0.3284	0.455	0.6072	0.7500	NAWQ_ACU
1C	9	7	0.3308	0.503	5.7848	0.0870	NAWQ_CHR
1WC	2	2	0.1485	0.613	7.0409	0.0870	NAWQ_CHR
BWC	2	1	0.1960	0.196	2.2529	0.0870	NAWQ_CHR
HRT	8	5	0.1249	0.178	2.0455	0.0870	NAWQ_CHR
IHP	12	7	0.1666	0.306	3.5121	0.0870	NAWQ_CHR
LMB	29	16	0.1149	0.151	1.7346	0.0870	NAWQ_CHR
LWC	2	1	0.1230	0.123	1.4138	0.0870	NAWQ_CHR
MWC	18	14	0.1122	0.131	1.5074	0.0870	NAWQ_CHR
NWT	2	2	0.1210	0.481	5.5274	0.0870	NAWQ_CHR
RAC	2	2	0.1480	0.843	9.6841	0.0870	NAWQ_CHR
UMB	9	5	0.2312	0.306	3.5149	0.0870	NAWQ_CHR
W4T	16	11	0.2607	0.491	5.6431	0.0870	NAWQ_CHR
WOL	13	13	0.8752	1.102	12.6673	0.0870	NAWQ_CHR
WS	10	8	0.3284	0.455	5.2342	0.0870	NAWQ_CHR
ANTIMONY							
HRT	8	1	0.0190	0.019	0.0311	0.6100	LCV_ALLO
LMB	29	1	0.0190	0.019	0.0311	0.6100	LCV_ALLO
MWC	18	2	0.0384	0.039	0.0644	0.6100	LCV_ALLO
HRT	8	1	0.0190	0.019	0.0311	0.6100	LCV_AQPL
LMB	29	1	0.0190	0.019	0.0311	0.6100	LCV_AQPL
MWC	18	2	0.0384	0.039	0.0644	0.6100	LCV_AQPL
HRT	8	1	0.0190	0.019	0.0035	5.4000	LCV_DAPH
LMB	29	1	0.0190	0.019	0.0035	5.4000	LCV_DAPH
MWC	18	2	0.0384	0.039	0.0073	5.4000	LCV_DAPH
HRT	8	1	0.0190	0.019	0.0119	1.6000	LCV_FISH
LMB	29	1	0.0190	0.019	0.0119	1.6000	LCV_FISH
MWC	18	2	0.0384	0.039	0.0246	1.6000	LCV_FISH
HRT	8	1	0.0190	0.019	0.0100	1.9000	LTV_DAPH
LMB	29	1	0.0190	0.019	0.0100	1.9000	LTV_DAPH
MWC	18	2	0.0384	0.039	0.0207	1.9000	LTV_DAPH
HRT	8	1	0.0190	0.019	0.0082	2.3100	LTV_FISH

B-5

	LMB	29	1	0.0190	0.019	0.0082	2.3100	LTV_FISH
	MWC	18	2	0.0384	0.039	0.0170	2.3100	LTV_FISH
	HRT	8	1	0.0190	0.019	0.0193	0.9850	S_ACU_V
	LMB	29	1	0.0190	0.019	0.0193	0.9850	S_ACU_V
	MWC	18	2	0.0384	0.039	0.0399	0.9850	S_ACU_V
	HRT	8	1	0.0190	0.019	0.1827	0.1040	S_CHR_V
	LMB	29	1	0.0190	0.019	0.1827	0.1040	S_CHR_V
	MWC	18	2	0.0384	0.039	0.3778	0.1040	S_CHR_V
ARSENIC	HRT	7	1	0.0010	0.001	0.0011	0.9140	LCV_DAPH
	LMB	26	6	0.0013	0.002	0.0017	0.9140	LCV_DAPH
	MWC	16	2	0.0010	0.001	0.0011	0.9140	LCV_DAPH
	UMB	8	2	0.0020	0.002	0.0022	0.9140	LCV_DAPH
	W4T	9	1	0.0010	0.001	0.0011	0.9140	LCV_DAPH
	WOL	12	1	0.0010	0.001	0.0011	0.9140	LCV_DAPH
	WS	9	1	0.0010	0.001	0.0011	0.9140	LCV_DAPH
	HRT	7	1	0.0010	0.001	0.0003	2.9620	LCV_FISH
	LMB	26	6	0.0013	0.002	0.0005	2.9620	LCV_FISH
	MWC	16	2	0.0010	0.001	0.0003	2.9620	LCV_FISH
	UMB	8	2	0.0020	0.002	0.0007	2.9620	LCV_FISH
	W4T	9	1	0.0010	0.001	0.0003	2.9620	LCV_FISH
	WOL	12	1	0.0010	0.001	0.0003	2.9620	LCV_FISH
	WS	9	1	0.0010	0.001	0.0003	2.9620	LCV_FISH
	HRT	7	1	0.0010	0.001	0.0016	0.6330	LTV_DAPH
	LMB	26	6	0.0013	0.002	0.0025	0.6330	LTV_DAPH
	MWC	16	2	0.0010	0.001	0.0016	0.6330	LTV_DAPH
	UMB	8	2	0.0020	0.002	0.0032	0.6330	LTV_DAPH
	W4T	9	1	0.0010	0.001	0.0016	0.6330	LTV_DAPH
	WOL	12	1	0.0010	0.001	0.0016	0.6330	LTV_DAPH
	WS	9	1	0.0010	0.001	0.0016	0.6330	LTV_DAPH
	HRT	7	1	0.0010	0.001	0.0005	2.1300	LTV_FISH
	LMB	26	6	0.0013	0.002	0.0007	2.1300	LTV_FISH
	MWC	16	2	0.0010	0.001	0.0005	2.1300	LTV_FISH
	UMB	8	2	0.0020	0.002	0.0009	2.1300	LTV_FISH
	W4T	9	1	0.0010	0.001	0.0005	2.1300	LTV_FISH
	WOL	12	1	0.0010	0.001	0.0005	2.1300	LTV_FISH
	WS	9	1	0.0010	0.001	0.0005	2.1300	LTV_FISH
	HRT	7	1	0.0010	0.001	0.0028	0.3600	NAWQ_ACU
	LMB	26	6	0.0013	0.002	0.0044	0.3600	NAWQ_ACU
	MWC	16	2	0.0010	0.001	0.0028	0.3600	NAWQ_ACU
	UMB	8	2	0.0020	0.002	0.0056	0.3600	NAWQ_ACU
	W4T	9	1	0.0010	0.001	0.0028	0.3600	NAWQ_ACU
	WOL	12	1	0.0010	0.001	0.0028	0.3600	NAWQ_ACU
	WS	9	1	0.0010	0.001	0.0028	0.3600	NAWQ_ACU
	HRT	7	1	0.0010	0.001	0.0053	0.1900	NAWQ_CHR
	LMB	26	6	0.0013	0.002	0.0083	0.1900	NAWQ_CHR
	MWC	16	2	0.0010	0.001	0.0053	0.1900	NAWQ_CHR
	UMB	8	2	0.0020	0.002	0.0105	0.1900	NAWQ_CHR
	W4T	9	1	0.0010	0.001	0.0053	0.1900	NAWQ_CHR
	WOL	12	1	0.0010	0.001	0.0053	0.1900	NAWQ_CHR
	WS	9	1	0.0010	0.001	0.0053	0.1900	NAWQ_CHR
Acetone	HRT	1	1	0.0010	0.001	0.0000	507.6400	LCV_ALLO
	IHP	1	1	0.0030	0.003	0.0000	507.6400	LCV_ALLO
	MWC	1	1	0.0050	0.005	0.0000	507.6400	LCV_ALLO
	UMB	1	1	0.0030	0.003	0.0000	507.6400	LCV_ALLO
	W4T	3	1	0.0020	0.002	0.0000	507.6400	LCV_ALLO
	WOL	1	1	0.0030	0.003	0.0000	507.6400	LCV_ALLO
	HRT	1	1	0.0010	0.001	0.0000	3114.1820	LCV_DAPH
	IHP	1	1	0.0030	0.003	0.0000	3114.1820	LCV_DAPH
	MWC	1	1	0.0050	0.005	0.0000	3114.1820	LCV_DAPH
	UMB	1	1	0.0030	0.003	0.0000	3114.1820	LCV_DAPH
	W4T	3	1	0.0020	0.002	0.0000	3114.1820	LCV_DAPH
	WOL	1	1	0.0030	0.003	0.0000	3114.1820	LCV_DAPH
	HRT	1	1	0.0010	0.001	0.0000	507.6400	LCV_FISH
	IHP	1	1	0.0030	0.003	0.0000	507.6400	LCV_FISH
	MWC	1	1	0.0050	0.005	0.0000	507.6400	LCV_FISH
	UMB	1	1	0.0030	0.003	0.0000	507.6400	LCV_FISH
	W4T	3	1	0.0020	0.002	0.0000	507.6400	LCV_FISH
	WOL	1	1	0.0030	0.003	0.0000	507.6400	LCV_FISH
	HRT	1	1	0.0010	0.001	0.0000	161.8670	LTV_FISH

B-6

IHP	1	1	0.0030	0.003	0.0000	161.8670	LTV_FISH
MWC	1	1	0.0050	0.005	0.0000	161.8670	LTV_FISH
UMB	1	1	0.0030	0.003	0.0000	161.8670	LTV_FISH
W4T	3	1	0.0020	0.002	0.0000	161.8670	LTV_FISH
WOL	1	1	0.0030	0.003	0.0000	161.8670	LTV_FISH
HRT	1	1	0.0010	0.001	0.0000	200.0000	S_ACU_V
IHP	1	1	0.0030	0.003	0.0000	200.0000	S_ACU_V
MWC	1	1	0.0050	0.005	0.0000	200.0000	S_ACU_V
UMB	1	1	0.0030	0.003	0.0000	200.0000	S_ACU_V
W4T	3	1	0.0020	0.002	0.0000	200.0000	S_ACU_V
WOL	1	1	0.0030	0.003	0.0000	200.0000	S_ACU_V
HRT	1	1	0.0010	0.001	0.0001	11.2000	S_CHR_V
IHP	1	1	0.0030	0.003	0.0003	11.2000	S_CHR_V
MWC	1	1	0.0050	0.005	0.0004	11.2000	S_CHR_V
UMB	1	1	0.0030	0.003	0.0003	11.2000	S_CHR_V
W4T	3	1	0.0020	0.002	0.0002	11.2000	S_CHR_V
WOL	1	1	0.0030	0.003	0.0003	11.2000	S_CHR_V

BARIUM

1C	9	8	0.0412	0.046	0.0079	5.8000	LCV_ALLO
1WC	2	2	0.0340	0.040	0.0070	5.8000	LCV_ALLO
BWC	2	2	0.0640	0.260	0.0448	5.8000	LCV_ALLO
HRT	8	8	0.0580	0.063	0.0109	5.8000	LCV_ALLO
IHP	12	11	0.0349	0.037	0.0063	5.8000	LCV_ALLO
LMB	29	29	0.0511	0.057	0.0098	5.8000	LCV_ALLO
LWC	2	2	0.0380	0.070	0.0120	5.8000	LCV_ALLO
MWC	18	16	0.0348	0.036	0.0063	5.8000	LCV_ALLO
NWT	2	1	0.0440	0.044	0.0076	5.8000	LCV_ALLO
RAC	2	1	0.0290	0.029	0.0050	5.8000	LCV_ALLO
UMB	9	9	0.0526	0.065	0.0113	5.8000	LCV_ALLO
W4T	16	16	0.1646	0.204	0.0351	5.8000	LCV_ALLO
WOL	13	12	0.0462	0.051	0.0087	5.8000	LCV_ALLO
WS	10	10	0.0619	0.075	0.0130	5.8000	LCV_ALLO
1C	9	8	0.0412	0.046	0.0079	5.8000	LCV_DAPH
1WC	2	2	0.0340	0.040	0.0070	5.8000	LCV_DAPH
BWC	2	2	0.0640	0.260	0.0448	5.8000	LCV_DAPH
HRT	8	8	0.0580	0.063	0.0109	5.8000	LCV_DAPH
IHP	12	11	0.0349	0.037	0.0063	5.8000	LCV_DAPH
LMB	29	29	0.0511	0.057	0.0098	5.8000	LCV_DAPH
LWC	2	2	0.0380	0.070	0.0120	5.8000	LCV_DAPH
MWC	18	16	0.0348	0.036	0.0063	5.8000	LCV_DAPH
NWT	2	1	0.0440	0.044	0.0076	5.8000	LCV_DAPH
RAC	2	1	0.0290	0.029	0.0050	5.8000	LCV_DAPH
UMB	9	9	0.0526	0.065	0.0113	5.8000	LCV_DAPH
W4T	16	16	0.1646	0.204	0.0351	5.8000	LCV_DAPH
WOL	13	12	0.0462	0.051	0.0087	5.8000	LCV_DAPH
WS	10	10	0.0619	0.075	0.0130	5.8000	LCV_DAPH
1C	9	8	0.0412	0.046	0.6609	0.0691	S_ACU_V
1WC	2	2	0.0340	0.040	0.5834	0.0691	S_ACU_V
BWC	2	2	0.0640	0.260	3.7587	0.0691	S_ACU_V
HRT	8	8	0.0580	0.063	0.9145	0.0691	S_ACU_V
IHP	12	11	0.0349	0.037	0.5297	0.0691	S_ACU_V
LMB	29	29	0.0511	0.057	0.8251	0.0691	S_ACU_V
LWC	2	2	0.0380	0.070	1.0068	0.0691	S_ACU_V
MWC	18	16	0.0348	0.036	0.5256	0.0691	S_ACU_V
NWT	2	1	0.0440	0.044	0.6368	0.0691	S_ACU_V
RAC	2	1	0.0290	0.029	0.4197	0.0691	S_ACU_V
UMB	9	9	0.0526	0.065	0.9445	0.0691	S_ACU_V
W4T	16	16	0.1646	0.204	2.9450	0.0691	S_ACU_V
WOL	13	12	0.0462	0.051	0.7334	0.0691	S_ACU_V
WS	10	10	0.0619	0.075	1.0878	0.0691	S_ACU_V
1C	9	8	0.0412	0.046	12.0185	0.0038	S_CHR_V
1WC	2	2	0.0340	0.040	10.6089	0.0038	S_CHR_V
BWC	2	2	0.0640	0.260	68.3490	0.0038	S_CHR_V
HRT	8	8	0.0580	0.063	16.6286	0.0038	S_CHR_V
IHP	12	11	0.0349	0.037	9.6327	0.0038	S_CHR_V
LMB	29	29	0.0511	0.057	15.0031	0.0038	S_CHR_V
LWC	2	2	0.0380	0.070	18.3076	0.0038	S_CHR_V
MWC	18	16	0.0348	0.036	9.5571	0.0038	S_CHR_V
NWT	2	1	0.0440	0.044	11.5789	0.0038	S_CHR_V
RAC	2	1	0.0290	0.029	7.6316	0.0038	S_CHR_V
UMB	9	9	0.0526	0.065	17.1754	0.0038	S_CHR_V

B-7

	W4T	16	16	0.1646	0.204	53.5530	0.0038	S_CHR_V
	WOL	13	12	0.0462	0.051	13.3365	0.0038	S_CHR_V
	WS	10	10	0.0619	0.075	19.7810	0.0038	S_CHR_V
BERYLLIUM	1C	9	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	HRT	8	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	IHP	12	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	LMB	29	5	0.0001	0.000	0.0373	0.0053	LCV_ALLO
	MWC	18	2	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	UMB	9	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	W4T	16	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	WOL	13	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	WS	10	1	0.0000	0.000	0.0000	0.0053	LCV_ALLO
	1C	9	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	HRT	8	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	IHP	12	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	LMB	29	5	0.0001	0.000	0.0000	100.0000	LCV_AQPL
	MWC	18	2	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	UMB	9	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	W4T	16	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	WOL	13	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	WS	10	1	0.0000	0.000	0.0000	100.0000	LCV_AQPL
	1C	9	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	HRT	8	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	IHP	12	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	LMB	29	5	0.0001	0.000	0.0373	0.0053	LCV_DAPH
	MWC	18	2	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	UMB	9	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	W4T	16	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	WOL	13	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	WS	10	1	0.0000	0.000	0.0000	0.0053	LCV_DAPH
	1C	9	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	HRT	8	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	IHP	12	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	LMB	29	5	0.0001	0.000	0.0035	0.0570	LCV_FISH
	MWC	18	2	0.0000	0.000	0.0000	0.0570	LCV_FISH
	UMB	9	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	W4T	16	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	WOL	13	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	WS	10	1	0.0000	0.000	0.0000	0.0570	LCV_FISH
	1C	9	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	HRT	8	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	IHP	12	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	LMB	29	5	0.0001	0.000	0.0521	0.0038	LTV_DAPH
	MWC	18	2	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	UMB	9	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	W4T	16	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	WOL	13	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	WS	10	1	0.0000	0.000	0.0000	0.0038	LTV_DAPH
	1C	9	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	HRT	8	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	IHP	12	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	LMB	29	5	0.0001	0.000	0.0013	0.1480	LTV_FISH
	MWC	18	2	0.0000	0.000	0.0000	0.1480	LTV_FISH
	UMB	9	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	W4T	16	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	WOL	13	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	WS	10	1	0.0000	0.000	0.0000	0.1480	LTV_FISH
	1C	9	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	HRT	8	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	IHP	12	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	LMB	29	5	0.0001	0.000	0.0007	0.2710	S_ACU_V
	MWC	18	2	0.0000	0.000	0.0000	0.2710	S_ACU_V
	UMB	9	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	W4T	16	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	WOL	13	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	WS	10	1	0.0000	0.000	0.0000	0.2710	S_ACU_V
	1C	9	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	HRT	8	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	IHP	12	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	LMB	29	5	0.0001	0.000	0.0389	0.0051	S_CHR_V

B-8

	MWC	18	2	0.0000	0.000	0.0000	0.0051	S_CHR_V
	UMB	9	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	W4T	16	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	WOL	13	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
	WS	10	1	0.0000	0.000	0.0000	0.0051	S_CHR_V
BORON	1C	9	3	0.0218	0.036	0.0041	8.8300	LCV_ALLO
	HRT	8	2	0.0354	0.044	0.0050	8.8300	LCV_ALLO
	IHP	12	3	0.0354	0.046	0.0052	8.8300	LCV_ALLO
	LMB	29	14	0.0498	0.054	0.0061	8.8300	LCV_ALLO
	MWC	18	7	0.0325	0.040	0.0046	8.8300	LCV_ALLO
	UMB	9	2	0.0394	0.055	0.0062	8.8300	LCV_ALLO
	W4T	16	7	0.1750	0.226	0.0255	8.8300	LCV_ALLO
	WOL	13	5	0.1029	0.207	0.0234	8.8300	LCV_ALLO
	WS	10	5	0.1099	0.240	0.0272	8.8300	LCV_ALLO
	1C	9	3	0.0218	0.036	0.0041	8.8300	LCV_DAPH
	HRT	8	2	0.0354	0.044	0.0050	8.8300	LCV_DAPH
	IHP	12	3	0.0354	0.046	0.0052	8.8300	LCV_DAPH
	LMB	29	14	0.0498	0.054	0.0061	8.8300	LCV_DAPH
	MWC	18	7	0.0325	0.040	0.0046	8.8300	LCV_DAPH
	UMB	9	2	0.0394	0.055	0.0062	8.8300	LCV_DAPH
	W4T	16	7	0.1750	0.226	0.0255	8.8300	LCV_DAPH
	WOL	13	5	0.1029	0.207	0.0234	8.8300	LCV_DAPH
	WS	10	5	0.1099	0.240	0.0272	8.8300	LCV_DAPH
	1C	9	3	0.0218	0.036	5.1193	0.0070	LTV_DAPH
	HRT	8	2	0.0354	0.044	6.2762	0.0070	LTV_DAPH
	IHP	12	3	0.0354	0.046	6.5284	0.0070	LTV_DAPH
	LMB	29	14	0.0498	0.054	7.7056	0.0070	LTV_DAPH
	MWC	18	7	0.0325	0.040	5.7857	0.0070	LTV_DAPH
	UMB	9	2	0.0394	0.055	7.8653	0.0070	LTV_DAPH
	W4T	16	7	0.1750	0.226	32.2145	0.0070	LTV_DAPH
	WOL	13	5	0.1029	0.207	29.5793	0.0070	LTV_DAPH
	WS	10	5	0.1099	0.240	34.2666	0.0070	LTV_DAPH
	1C	9	3	0.0218	0.036	0.0033	11.0000	S_ACU_V
	HRT	8	2	0.0354	0.044	0.0040	11.0000	S_ACU_V
	IHP	12	3	0.0354	0.046	0.0042	11.0000	S_ACU_V
	LMB	29	14	0.0498	0.054	0.0049	11.0000	S_ACU_V
	MWC	18	7	0.0325	0.040	0.0037	11.0000	S_ACU_V
	UMB	9	2	0.0394	0.055	0.0050	11.0000	S_ACU_V
	W4T	16	7	0.1750	0.226	0.0205	11.0000	S_ACU_V
	WOL	13	5	0.1029	0.207	0.0188	11.0000	S_ACU_V
	WS	10	5	0.1099	0.240	0.0218	11.0000	S_ACU_V
	1C	9	3	0.0218	0.036	0.0655	0.5470	S_CHR_V
	HRT	8	2	0.0354	0.044	0.0803	0.5470	S_CHR_V
	IHP	12	3	0.0354	0.046	0.0835	0.5470	S_CHR_V
	LMB	29	14	0.0498	0.054	0.0986	0.5470	S_CHR_V
	MWC	18	7	0.0325	0.040	0.0740	0.5470	S_CHR_V
	UMB	9	2	0.0394	0.055	0.1007	0.5470	S_CHR_V
	W4T	16	7	0.1750	0.226	0.4123	0.5470	S_CHR_V
	WOL	13	5	0.1029	0.207	0.3785	0.5470	S_CHR_V
	WS	10	5	0.1099	0.240	0.4385	0.5470	S_CHR_V
CADMIUM	LMB	29	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	MWC	17	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	W4T	12	2	0.0011	0.001	9.1811	0.0002	LCV_ALLO
	WOL	13	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	LMB	29	1	0.0000	0.000	0.0000	0.0020	LCV_AQPL
	MWC	17	1	0.0000	0.000	0.0000	0.0020	LCV_AQPL
	W4T	12	2	0.0011	0.001	0.6886	0.0020	LCV_AQPL
	WOL	13	1	0.0000	0.000	0.0000	0.0020	LCV_AQPL
	LMB	29	1	0.0000	0.000	0.0000	0.0002	LCV_DAPH
	MWC	17	1	0.0000	0.000	0.0000	0.0002	LCV_DAPH
	W4T	12	2	0.0011	0.001	9.1811	0.0002	LCV_DAPH
	WOL	13	1	0.0000	0.000	0.0000	0.0002	LCV_DAPH
	LMB	29	1	0.0000	0.000	0.0000	0.0017	LCV_FISH
	MWC	17	1	0.0000	0.000	0.0000	0.0017	LCV_FISH
	W4T	12	2	0.0011	0.001	0.8101	0.0017	LCV_FISH
	WOL	13	1	0.0000	0.000	0.0000	0.0017	LCV_FISH
	LMB	29	1	0.0000	0.000	0.0000	0.0008	LTV_DAPH
	MWC	17	1	0.0000	0.000	0.0000	0.0008	LTV_DAPH
	W4T	12	2	0.0011	0.001	1.8362	0.0008	LTV_DAPH

B-9

	WOL	13	1	0.0000	0.000	0.0000	0.0008	LTV_DAPH
	LMB	29	1	0.0000	0.000	0.0000	0.0018	LTV_FISH
	MWC	17	1	0.0000	0.000	0.0000	0.0018	LTV_FISH
	W4T	12	2	0.0011	0.001	0.7651	0.0018	LTV_FISH
	WOL	13	1	0.0000	0.000	0.0000	0.0018	LTV_FISH
	LMB	29	1	0.0000	0.000	0.0000	0.0039	NAWQ_ACU
	MWC	17	1	0.0000	0.000	0.0000	0.0039	NAWQ_ACU
	W4T	12	2	0.0011	0.001	0.3531	0.0039	NAWQ_ACU
	WOL	13	1	0.0000	0.000	0.0000	0.0039	NAWQ_ACU
	LMB	29	1	0.0000	0.000	0.0000	0.0011	NAWQ_CHR
	MWC	17	1	0.0000	0.000	0.0000	0.0011	NAWQ_CHR
	W4T	12	2	0.0011	0.001	1.2520	0.0011	NAWQ_CHR
	WOL	13	1	0.0000	0.000	0.0000	0.0011	NAWQ_CHR
CALCIUM	1C	9	9	43.8333	48.460	0.4178	116.0000	LCV_ALLO
	1WC	2	2	40.9000	47.845	0.4125	116.0000	LCV_ALLO
	BWC	2	2	23.5500	80.058	0.6902	116.0000	LCV_ALLO
	HRT	8	8	41.7500	48.219	0.4157	116.0000	LCV_ALLO
	IHP	12	12	44.3667	47.985	0.4137	116.0000	LCV_ALLO
	LMB	29	29	58.4483	66.339	0.5719	116.0000	LCV_ALLO
	LWC	2	2	44.2000	58.090	0.5008	116.0000	LCV_ALLO
	MWC	18	18	43.4833	46.474	0.4006	116.0000	LCV_ALLO
	NWT	2	2	48.8500	129.350	1.1151	116.0000	LCV_ALLO
	RAC	2	2	84.1000	137.767	1.1876	116.0000	LCV_ALLO
	UMB	9	9	63.6889	82.307	0.7095	116.0000	LCV_ALLO
	W4T	16	16	76.5938	90.049	0.7763	116.0000	LCV_ALLO
	WOL	13	13	44.1000	47.219	0.4071	116.0000	LCV_ALLO
	WS	10	10	37.0600	45.099	0.3888	116.0000	LCV_ALLO
	1C	9	9	43.8333	48.460	0.4178	116.0000	LCV_DAPH
	1WC	2	2	40.9000	47.845	0.4125	116.0000	LCV_DAPH
	BWC	2	2	23.5500	80.058	0.6902	116.0000	LCV_DAPH
	HRT	8	8	41.7500	48.219	0.4157	116.0000	LCV_DAPH
	IHP	12	12	44.3667	47.985	0.4137	116.0000	LCV_DAPH
	LMB	29	29	58.4483	66.339	0.5719	116.0000	LCV_DAPH
	LWC	2	2	44.2000	58.090	0.5008	116.0000	LCV_DAPH
	MWC	18	18	43.4833	46.474	0.4006	116.0000	LCV_DAPH
	NWT	2	2	48.8500	129.350	1.1151	116.0000	LCV_DAPH
	RAC	2	2	84.1000	137.767	1.1876	116.0000	LCV_DAPH
	UMB	9	9	63.6889	82.307	0.7095	116.0000	LCV_DAPH
	W4T	16	16	76.5938	90.049	0.7763	116.0000	LCV_DAPH
	WOL	13	13	44.1000	47.219	0.4071	116.0000	LCV_DAPH
	WS	10	10	37.0600	45.099	0.3888	116.0000	LCV_DAPH
CHROMIUM	HRT	8	1	0.0030	0.003	1.5000	0.0020	LCV_ALLO
	LMB	29	1	0.0070	0.007	3.5000	0.0020	LCV_ALLO
	UMB	9	2	0.0036	0.005	2.4665	0.0020	LCV_ALLO
	W4T	16	2	0.0040	0.004	2.0000	0.0020	LCV_ALLO
	WOL	13	7	0.0154	0.019	9.7495	0.0020	LCV_ALLO
	HRT	8	1	0.0030	0.003	1.5000	0.0020	LCV_AQPL
	LMB	29	1	0.0070	0.007	3.5000	0.0020	LCV_AQPL
	UMB	9	2	0.0036	0.005	2.4665	0.0020	LCV_AQPL
	W4T	16	2	0.0040	0.004	2.0000	0.0020	LCV_AQPL
	WOL	13	7	0.0154	0.019	9.7495	0.0020	LCV_AQPL
	HRT	8	1	0.0030	0.003	0.4892	0.0061	LCV_DAPH
	LMB	29	1	0.0070	0.007	1.1416	0.0061	LCV_DAPH
	UMB	9	2	0.0036	0.005	0.8045	0.0061	LCV_DAPH
	W4T	16	2	0.0040	0.004	0.6523	0.0061	LCV_DAPH
	WOL	13	7	0.0154	0.019	3.1799	0.0061	LCV_DAPH
	HRT	8	1	0.0030	0.003	0.0410	0.0732	LCV_FISH
	LMB	29	1	0.0070	0.007	0.0957	0.0732	LCV_FISH
	UMB	9	2	0.0036	0.005	0.0674	0.0732	LCV_FISH
	W4T	16	2	0.0040	0.004	0.0547	0.0732	LCV_FISH
	WOL	13	7	0.0154	0.019	0.2665	0.0732	LCV_FISH
	HRT	8	1	0.0030	0.003	6.0000	0.0005	LTV_DAPH
	LMB	29	1	0.0070	0.007	14.0000	0.0005	LTV_DAPH
	UMB	9	2	0.0036	0.005	9.8660	0.0005	LTV_DAPH
	W4T	16	2	0.0040	0.004	8.0000	0.0005	LTV_DAPH
	WOL	13	7	0.0154	0.019	38.9979	0.0005	LTV_DAPH
	HRT	8	1	0.0030	0.003	0.0588	0.0510	LTV_FISH
	LMB	29	1	0.0070	0.007	0.1373	0.0510	LTV_FISH
	UMB	9	2	0.0036	0.005	0.0967	0.0510	LTV_FISH

B-10

W4T	16	2	0.0040	0.004	0.0784	0.0510	LTV_FISH
WOL	13	7	0.0154	0.019	0.3823	0.0510	LTV_FISH
HRT	8	1	0.0030	0.003	0.1875	0.0160	NAWQ_ACU
LMB	29	1	0.0070	0.007	0.4375	0.0160	NAWQ_ACU
UMB	9	2	0.0036	0.005	0.3083	0.0160	NAWQ_ACU
W4T	16	2	0.0040	0.004	0.2500	0.0160	NAWQ_ACU
WOL	13	7	0.0154	0.019	1.2187	0.0160	NAWQ_ACU
HRT	8	1	0.0030	0.003	0.2727	0.0110	NAWQ_CHR
LMB	29	1	0.0070	0.007	0.6364	0.0110	NAWQ_CHR
UMB	9	2	0.0036	0.005	0.4485	0.0110	NAWQ_CHR
W4T	16	2	0.0040	0.004	0.3636	0.0110	NAWQ_CHR
WOL	13	7	0.0154	0.019	1.7726	0.0110	NAWQ_CHR

COBALT

HRT	8	1	0.0020	0.002	0.3922	0.0051	LCV_ALLO
UMB	9	1	0.0050	0.005	0.9804	0.0051	LCV_ALLO
W4T	16	1	0.0030	0.003	0.5882	0.0051	LCV_ALLO
WOL	13	1	0.0020	0.002	0.3922	0.0051	LCV_ALLO
WS	10	1	0.0110	0.011	2.1569	0.0051	LCV_ALLO
HRT	8	1	0.0020	0.002	0.3922	0.0051	LCV_DAPH
UMB	9	1	0.0050	0.005	0.9804	0.0051	LCV_DAPH
W4T	16	1	0.0030	0.003	0.5882	0.0051	LCV_DAPH
WOL	13	1	0.0020	0.002	0.3922	0.0051	LCV_DAPH
WS	10	1	0.0110	0.011	2.1569	0.0051	LCV_DAPH
HRT	8	1	0.0020	0.002	0.0069	0.2900	LCV_FISH
UMB	9	1	0.0050	0.005	0.0172	0.2900	LCV_FISH
W4T	16	1	0.0030	0.003	0.0103	0.2900	LCV_FISH
WOL	13	1	0.0020	0.002	0.0069	0.2900	LCV_FISH
WS	10	1	0.0110	0.011	0.0379	0.2900	LCV_FISH
HRT	8	1	0.0020	0.002	0.4545	0.0044	LTV_DAPH
UMB	9	1	0.0050	0.005	1.1364	0.0044	LTV_DAPH
W4T	16	1	0.0030	0.003	0.6818	0.0044	LTV_DAPH
WOL	13	1	0.0020	0.002	0.4545	0.0044	LTV_DAPH
WS	10	1	0.0110	0.011	2.5000	0.0044	LTV_DAPH
HRT	8	1	0.0020	0.002	0.0025	0.8100	LTV_FISH
UMB	9	1	0.0050	0.005	0.0062	0.8100	LTV_FISH
W4T	16	1	0.0030	0.003	0.0037	0.8100	LTV_FISH
WOL	13	1	0.0020	0.002	0.0025	0.8100	LTV_FISH
WS	10	1	0.0110	0.011	0.0136	0.8100	LTV_FISH
HRT	8	1	0.0020	0.002	0.0103	0.1950	S_ACU_V
UMB	9	1	0.0050	0.005	0.0256	0.1950	S_ACU_V
W4T	16	1	0.0030	0.003	0.0154	0.1950	S_ACU_V
WOL	13	1	0.0020	0.002	0.0103	0.1950	S_ACU_V
WS	10	1	0.0110	0.011	0.0564	0.1950	S_ACU_V
HRT	8	1	0.0020	0.002	0.6536	0.0031	S_CHR_V
UMB	9	1	0.0050	0.005	1.6340	0.0031	S_CHR_V
W4T	16	1	0.0030	0.003	0.9804	0.0031	S_CHR_V
WOL	13	1	0.0020	0.002	0.6536	0.0031	S_CHR_V
WS	10	1	0.0110	0.011	3.5948	0.0031	S_CHR_V

COPPER

1C	9	4	0.0024	0.004	16.1181	0.0002	LCV_ALLO
1WC	2	1	0.0070	0.007	30.4348	0.0002	LCV_ALLO
HRT	8	1	0.0020	0.002	8.6957	0.0002	LCV_ALLO
IHP	12	7	0.0059	0.008	34.9408	0.0002	LCV_ALLO
LMB	29	18	0.0050	0.006	27.3447	0.0002	LCV_ALLO
LWC	2	1	0.0130	0.013	56.5217	0.0002	LCV_ALLO
MWC	18	10	0.0053	0.007	30.1813	0.0002	LCV_ALLO
UMB	9	5	0.0091	0.015	64.5996	0.0002	LCV_ALLO
W4T	16	6	0.0099	0.014	60.9745	0.0002	LCV_ALLO
WOL	13	6	0.0063	0.008	35.1402	0.0002	LCV_ALLO
WS	10	3	0.0028	0.004	16.9332	0.0002	LCV_ALLO
1C	9	4	0.0024	0.004	3.7072	0.0010	LCV_AQPL
1WC	2	1	0.0070	0.007	7.0000	0.0010	LCV_AQPL
HRT	8	1	0.0020	0.002	2.0000	0.0010	LCV_AQPL
IHP	12	7	0.0059	0.008	8.0364	0.0010	LCV_AQPL
LMB	29	18	0.0050	0.006	6.2893	0.0010	LCV_AQPL
LWC	2	1	0.0130	0.013	13.0000	0.0010	LCV_AQPL
MWC	18	10	0.0053	0.007	6.9417	0.0010	LCV_AQPL
UMB	9	5	0.0091	0.015	14.8579	0.0010	LCV_AQPL
W4T	16	6	0.0099	0.014	14.0241	0.0010	LCV_AQPL
WOL	13	6	0.0063	0.008	8.0822	0.0010	LCV_AQPL
WS	10	3	0.0028	0.004	3.8946	0.0010	LCV_AQPL

B-11

1C	9	4	0.0024	0.004	16.1181	0.0002	LCV_DAPH
1WC	2	1	0.0070	0.007	30.4348	0.0002	LCV_DAPH
HRT	8	1	0.0020	0.002	8.6957	0.0002	LCV_DAPH
IHP	12	7	0.0059	0.008	34.9408	0.0002	LCV_DAPH
LMB	29	18	0.0050	0.006	27.3447	0.0002	LCV_DAPH
LWC	2	1	0.0130	0.013	56.5217	0.0002	LCV_DAPH
MWC	18	10	0.0053	0.007	30.1813	0.0002	LCV_DAPH
UMB	9	5	0.0091	0.015	64.5996	0.0002	LCV_DAPH
W4T	16	6	0.0099	0.014	60.9745	0.0002	LCV_DAPH
WOL	13	6	0.0063	0.008	35.1402	0.0002	LCV_DAPH
WS	10	3	0.0028	0.004	16.9332	0.0002	LCV_DAPH
1C	9	4	0.0024	0.004	0.9756	0.0038	LCV_FISH
1WC	2	1	0.0070	0.007	1.8421	0.0038	LCV_FISH
HRT	8	1	0.0020	0.002	0.5263	0.0038	LCV_FISH
IHP	12	7	0.0059	0.008	2.1148	0.0038	LCV_FISH
LMB	29	18	0.0050	0.006	1.6551	0.0038	LCV_FISH
LWC	2	1	0.0130	0.013	3.4211	0.0038	LCV_FISH
MWC	18	10	0.0053	0.007	1.8268	0.0038	LCV_FISH
UMB	9	5	0.0091	0.015	3.9100	0.0038	LCV_FISH
W4T	16	6	0.0099	0.014	3.6906	0.0038	LCV_FISH
WOL	13	6	0.0063	0.008	2.1269	0.0038	LCV_FISH
WS	10	3	0.0028	0.004	1.0249	0.0038	LCV_FISH
1C	9	4	0.0024	0.004	0.6111	0.0061	LCV_NDI
1WC	2	1	0.0070	0.007	1.1540	0.0061	LCV_NDI
HRT	8	1	0.0020	0.002	0.3297	0.0061	LCV_NDI
IHP	12	7	0.0059	0.008	1.3248	0.0061	LCV_NDI
LMB	29	18	0.0050	0.006	1.0368	0.0061	LCV_NDI
LWC	2	1	0.0130	0.013	2.1431	0.0061	LCV_NDI
MWC	18	10	0.0053	0.007	1.1444	0.0061	LCV_NDI
UMB	9	5	0.0091	0.015	2.4494	0.0061	LCV_NDI
W4T	16	6	0.0099	0.014	2.3119	0.0061	LCV_NDI
WOL	13	6	0.0063	0.008	1.3324	0.0061	LCV_NDI
WS	10	3	0.0028	0.004	0.6420	0.0061	LCV_NDI
1C	9	4	0.0024	0.004	18.0837	0.0002	LTV_DAPH
1WC	2	1	0.0070	0.007	34.1463	0.0002	LTV_DAPH
HRT	8	1	0.0020	0.002	9.7561	0.0002	LTV_DAPH
IHP	12	7	0.0059	0.008	39.2019	0.0002	LTV_DAPH
LMB	29	18	0.0050	0.006	30.6795	0.0002	LTV_DAPH
LWC	2	1	0.0130	0.013	63.4146	0.0002	LTV_DAPH
MWC	18	10	0.0053	0.007	33.8619	0.0002	LTV_DAPH
UMB	9	5	0.0091	0.015	72.4776	0.0002	LTV_DAPH
W4T	16	6	0.0099	0.014	68.4105	0.0002	LTV_DAPH
WOL	13	6	0.0063	0.008	39.4255	0.0002	LTV_DAPH
WS	10	3	0.0028	0.004	18.9982	0.0002	LTV_DAPH
1C	9	4	0.0024	0.004	0.7414	0.0050	LTV_FISH
1WC	2	1	0.0070	0.007	1.4000	0.0050	LTV_FISH
HRT	8	1	0.0020	0.002	0.4000	0.0050	LTV_FISH
IHP	12	7	0.0059	0.008	1.6073	0.0050	LTV_FISH
LMB	29	18	0.0050	0.006	1.2579	0.0050	LTV_FISH
LWC	2	1	0.0130	0.013	2.6000	0.0050	LTV_FISH
MWC	18	10	0.0053	0.007	1.3883	0.0050	LTV_FISH
UMB	9	5	0.0091	0.015	2.9716	0.0050	LTV_FISH
W4T	16	6	0.0099	0.014	2.8048	0.0050	LTV_FISH
WOL	13	6	0.0063	0.008	1.6164	0.0050	LTV_FISH
WS	10	3	0.0028	0.004	0.7789	0.0050	LTV_FISH
1C	9	4	0.0024	0.004	0.2060	0.0180	NAWQ_ACU
1WC	2	1	0.0070	0.007	0.3889	0.0180	NAWQ_ACU
HRT	8	1	0.0020	0.002	0.1111	0.0180	NAWQ_ACU
IHP	12	7	0.0059	0.008	0.4465	0.0180	NAWQ_ACU
LMB	29	18	0.0050	0.006	0.3494	0.0180	NAWQ_ACU
LWC	2	1	0.0130	0.013	0.7222	0.0180	NAWQ_ACU
MWC	18	10	0.0053	0.007	0.3856	0.0180	NAWQ_ACU
UMB	9	5	0.0091	0.015	0.8254	0.0180	NAWQ_ACU
W4T	16	6	0.0099	0.014	0.7791	0.0180	NAWQ_ACU
WOL	13	6	0.0063	0.008	0.4490	0.0180	NAWQ_ACU
WS	10	3	0.0028	0.004	0.2164	0.0180	NAWQ_ACU
1C	9	4	0.0024	0.004	0.3089	0.0120	NAWQ_CHR
1WC	2	1	0.0070	0.007	0.5833	0.0120	NAWQ_CHR
HRT	8	1	0.0020	0.002	0.1667	0.0120	NAWQ_CHR
IHP	12	7	0.0059	0.008	0.6697	0.0120	NAWQ_CHR
LMB	29	18	0.0050	0.006	0.5241	0.0120	NAWQ_CHR

B-12

LWC	2	1	0.0130	0.013	1.0833	0.0120	NAWQ_CHR
MWC	18	10	0.0053	0.007	0.5785	0.0120	NAWQ_CHR
UMB	9	5	0.0091	0.015	1.2382	0.0120	NAWQ_CHR
W4T	16	6	0.0099	0.014	1.1687	0.0120	NAWQ_CHR
WOL	13	6	0.0063	0.008	0.6735	0.0120	NAWQ_CHR
WS	10	3	0.0028	0.004	0.3246	0.0120	NAWQ_CHR

Carbon disulfide

1C	1	1	0.0070	0.007	0.0287	0.2440	LCV_ALLO
HRT	1	1	0.0050	0.005	0.0205	0.2440	LCV_ALLO
IHP	1	1	0.0050	0.005	0.0205	0.2440	LCV_ALLO
LMB	1	1	0.0110	0.011	0.0451	0.2440	LCV_ALLO
MWC	1	1	0.0130	0.013	0.0533	0.2440	LCV_ALLO
NWT	1	1	0.0040	0.004	0.0164	0.2440	LCV_ALLO
RAC	1	1	0.0160	0.016	0.0656	0.2440	LCV_ALLO
UMB	1	1	0.0110	0.011	0.0451	0.2440	LCV_ALLO
WOL	1	1	0.0030	0.003	0.0123	0.2440	LCV_ALLO
WS	1	1	0.0170	0.017	0.0697	0.2440	LCV_ALLO
1C	1	1	0.0070	0.007	0.0287	0.2440	LCV_DAPH
HRT	1	1	0.0050	0.005	0.0205	0.2440	LCV_DAPH
IHP	1	1	0.0050	0.005	0.0205	0.2440	LCV_DAPH
LMB	1	1	0.0110	0.011	0.0451	0.2440	LCV_DAPH
MWC	1	1	0.0130	0.013	0.0533	0.2440	LCV_DAPH
NWT	1	1	0.0040	0.004	0.0164	0.2440	LCV_DAPH
RAC	1	1	0.0160	0.016	0.0656	0.2440	LCV_DAPH
UMB	1	1	0.0110	0.011	0.0451	0.2440	LCV_DAPH
WOL	1	1	0.0030	0.003	0.0123	0.2440	LCV_DAPH
WS	1	1	0.0170	0.017	0.0697	0.2440	LCV_DAPH
1C	1	1	0.0070	0.007	0.0007	9.5380	LCV_FISH
HRT	1	1	0.0050	0.005	0.0005	9.5380	LCV_FISH
IHP	1	1	0.0050	0.005	0.0005	9.5380	LCV_FISH
LMB	1	1	0.0110	0.011	0.0012	9.5380	LCV_FISH
MWC	1	1	0.0130	0.013	0.0014	9.5380	LCV_FISH
NWT	1	1	0.0040	0.004	0.0004	9.5380	LCV_FISH
RAC	1	1	0.0160	0.016	0.0017	9.5380	LCV_FISH
UMB	1	1	0.0110	0.011	0.0012	9.5380	LCV_FISH
WOL	1	1	0.0030	0.003	0.0003	9.5380	LCV_FISH
WS	1	1	0.0170	0.017	0.0018	9.5380	LCV_FISH
1C	1	1	0.0070	0.007	0.0012	5.7190	LTV_FISH
HRT	1	1	0.0050	0.005	0.0009	5.7190	LTV_FISH
IHP	1	1	0.0050	0.005	0.0009	5.7190	LTV_FISH
LMB	1	1	0.0110	0.011	0.0019	5.7190	LTV_FISH
MWC	1	1	0.0130	0.013	0.0023	5.7190	LTV_FISH
NWT	1	1	0.0040	0.004	0.0007	5.7190	LTV_FISH
RAC	1	1	0.0160	0.016	0.0028	5.7190	LTV_FISH
UMB	1	1	0.0110	0.011	0.0019	5.7190	LTV_FISH
WOL	1	1	0.0030	0.003	0.0005	5.7190	LTV_FISH
WS	1	1	0.0170	0.017	0.0030	5.7190	LTV_FISH
1C	1	1	0.0070	0.007	0.0440	0.1590	S_ACU_V
HRT	1	1	0.0050	0.005	0.0314	0.1590	S_ACU_V
IHP	1	1	0.0050	0.005	0.0314	0.1590	S_ACU_V
LMB	1	1	0.0110	0.011	0.0692	0.1590	S_ACU_V
MWC	1	1	0.0130	0.013	0.0818	0.1590	S_ACU_V
NWT	1	1	0.0040	0.004	0.0252	0.1590	S_ACU_V
RAC	1	1	0.0160	0.016	0.1006	0.1590	S_ACU_V
UMB	1	1	0.0110	0.011	0.0692	0.1590	S_ACU_V
WOL	1	1	0.0030	0.003	0.0189	0.1590	S_ACU_V
WS	1	1	0.0170	0.017	0.1069	0.1590	S_ACU_V
1C	1	1	0.0070	0.007	0.7874	0.0089	S_CHR_V
HRT	1	1	0.0050	0.005	0.5624	0.0089	S_CHR_V
IHP	1	1	0.0050	0.005	0.5624	0.0089	S_CHR_V
LMB	1	1	0.0110	0.011	1.2373	0.0089	S_CHR_V
MWC	1	1	0.0130	0.013	1.4623	0.0089	S_CHR_V
NWT	1	1	0.0040	0.004	0.4499	0.0089	S_CHR_V
RAC	1	1	0.0160	0.016	1.7998	0.0089	S_CHR_V
UMB	1	1	0.0110	0.011	1.2373	0.0089	S_CHR_V
WOL	1	1	0.0030	0.003	0.3375	0.0089	S_CHR_V
WS	1	1	0.0170	0.017	1.9123	0.0089	S_CHR_V

Chloroform

1C	1	1	0.0000	0.000	0.0000	1.2400	LCV_ALLO
IHP	1	1	0.0020	0.002	0.0016	1.2400	LCV_ALLO
MWC	1	1	0.0010	0.001	0.0008	1.2400	LCV_ALLO

B-13

WOL	1	1	0.0000	0.000	0.0000	1.2400	LCV_ALLO
1C	1	1	0.0000	0.000	0.0000	4.4830	LCV_DAPH
IHP	1	1	0.0020	0.002	0.0004	4.4830	LCV_DAPH
MWC	1	1	0.0010	0.001	0.0002	4.4830	LCV_DAPH
WOL	1	1	0.0000	0.000	0.0000	1.2400	LCV_FISH
1C	1	1	0.0000	0.000	0.0000	1.2400	LCV_FISH
IHP	1	1	0.0020	0.002	0.0016	1.2400	LCV_FISH
MWC	1	1	0.0010	0.001	0.0008	1.2400	LCV_FISH
WOL	1	1	0.0000	0.000	0.0000	1.2400	LCV_FISH
1C	1	1	0.0000	0.000	0.0000	8.4000	LTV_FISH
IHP	1	1	0.0020	0.002	0.0002	8.4000	LTV_FISH
MWC	1	1	0.0010	0.001	0.0001	8.4000	LTV_FISH
WOL	1	1	0.0000	0.000	0.0000	8.4000	LTV_FISH
1C	1	1	0.0000	0.000	0.0000	3.3600	S_ACU_V
IHP	1	1	0.0020	0.002	0.0006	3.3600	S_ACU_V
MWC	1	1	0.0010	0.001	0.0003	3.3600	S_ACU_V
WOL	1	1	0.0000	0.000	0.0000	3.3600	S_ACU_V
1C	1	1	0.0000	0.000	0.0000	0.1880	S_CHR_V
IHP	1	1	0.0020	0.002	0.0106	0.1880	S_CHR_V
MWC	1	1	0.0010	0.001	0.0053	0.1880	S_CHR_V
WOL	1	1	0.0000	0.000	0.0000	0.1880	S_CHR_V

IRON

1C	9	8	0.3924	0.592	3.7449	0.1580	LCV_ALLO
BWC	2	1	0.1270	0.127	0.8038	0.1580	LCV_ALLO
HRT	8	8	0.2448	0.305	1.9294	0.1580	LCV_ALLO
IHP	12	8	0.2020	0.357	2.2619	0.1580	LCV_ALLO
LMB	29	28	0.1870	0.231	1.4639	0.1580	LCV_ALLO
LWC	2	2	0.1700	0.334	2.1149	0.1580	LCV_ALLO
MWC	18	16	0.1442	0.166	1.0498	0.1580	LCV_ALLO
RAC	2	1	0.2590	0.259	1.6392	0.1580	LCV_ALLO
UMB	9	9	0.3292	0.461	2.9168	0.1580	LCV_ALLO
W4T	16	15	0.5062	0.761	4.8151	0.1580	LCV_ALLO
WOL	13	13	1.0815	1.346	8.5173	0.1580	LCV_ALLO
WS	10	10	0.6578	1.021	6.4590	0.1580	LCV_ALLO
1C	9	8	0.3924	0.592	3.7449	0.1580	LCV_DAPH
BWC	2	1	0.1270	0.127	0.8038	0.1580	LCV_DAPH
HRT	8	8	0.2448	0.305	1.9294	0.1580	LCV_DAPH
IHP	12	8	0.2020	0.357	2.2619	0.1580	LCV_DAPH
LMB	29	28	0.1870	0.231	1.4639	0.1580	LCV_DAPH
LWC	2	2	0.1700	0.334	2.1149	0.1580	LCV_DAPH
MWC	18	16	0.1442	0.166	1.0498	0.1580	LCV_DAPH
RAC	2	1	0.2590	0.259	1.6392	0.1580	LCV_DAPH
UMB	9	9	0.3292	0.461	2.9168	0.1580	LCV_DAPH
W4T	16	15	0.5062	0.761	4.8151	0.1580	LCV_DAPH
WOL	13	13	1.0815	1.346	8.5173	0.1580	LCV_DAPH
WS	10	10	0.6578	1.021	6.4590	0.1580	LCV_DAPH
1C	9	8	0.3924	0.592	0.4552	1.3000	LCV_FISH
BWC	2	1	0.1270	0.127	0.0977	1.3000	LCV_FISH
HRT	8	8	0.2448	0.305	0.2345	1.3000	LCV_FISH
IHP	12	8	0.2020	0.357	0.2749	1.3000	LCV_FISH
LMB	29	28	0.1870	0.231	0.1779	1.3000	LCV_FISH
LWC	2	2	0.1700	0.334	0.2570	1.3000	LCV_FISH
MWC	18	16	0.1442	0.166	0.1276	1.3000	LCV_FISH
RAC	2	1	0.2590	0.259	0.1992	1.3000	LCV_FISH
UMB	9	9	0.3292	0.461	0.3545	1.3000	LCV_FISH
W4T	16	15	0.5062	0.761	0.5852	1.3000	LCV_FISH
WOL	13	13	1.0815	1.346	1.0352	1.3000	LCV_FISH
WS	10	10	0.6578	1.021	0.7850	1.3000	LCV_FISH
1C	9	8	0.3924	0.592	36.9811	0.0160	LTV_DAPH
BWC	2	1	0.1270	0.127	7.9375	0.0160	LTV_DAPH
HRT	8	8	0.2448	0.305	19.0530	0.0160	LTV_DAPH
IHP	12	8	0.2020	0.357	22.3365	0.0160	LTV_DAPH
LMB	29	28	0.1870	0.231	14.4561	0.0160	LTV_DAPH
LWC	2	2	0.1700	0.334	20.8848	0.0160	LTV_DAPH
MWC	18	16	0.1442	0.166	10.3667	0.0160	LTV_DAPH
RAC	2	1	0.2590	0.259	16.1875	0.0160	LTV_DAPH
UMB	9	9	0.3292	0.461	28.8034	0.0160	LTV_DAPH
W4T	16	15	0.5062	0.761	47.5493	0.0160	LTV_DAPH
WOL	13	13	1.0815	1.346	84.1088	0.0160	LTV_DAPH
WS	10	10	0.6578	1.021	63.7822	0.0160	LTV_DAPH
1C	9	8	0.3924	0.592	0.5917	1.0000	NAWQ_CHR

B-14

BWC	2	1	0.1270	0.127	0.1270	1.0000	NAWQ_CHR
HRT	8	8	0.2448	0.305	0.3048	1.0000	NAWQ_CHR
IHP	12	8	0.2020	0.357	0.3574	1.0000	NAWQ_CHR
LMB	29	28	0.1870	0.231	0.2313	1.0000	NAWQ_CHR
LWC	2	2	0.1700	0.334	0.3342	1.0000	NAWQ_CHR
MWC	18	16	0.1442	0.166	0.1659	1.0000	NAWQ_CHR
RAC	2	1	0.2590	0.259	0.2590	1.0000	NAWQ_CHR
UMB	9	9	0.3292	0.461	0.4609	1.0000	NAWQ_CHR
W4T	16	15	0.5062	0.761	0.7608	1.0000	NAWQ_CHR
WOL	13	13	1.0815	1.346	1.3457	1.0000	NAWQ_CHR
WS	10	10	0.6578	1.021	1.0205	1.0000	NAWQ_CHR

LEAD

1C	8	1	0.0020	0.002	0.1631	0.0123	LCV_ALLO
BWC	2	1	0.0040	0.004	0.3263	0.0123	LCV_ALLO
IHP	11	1	0.0040	0.004	0.3263	0.0123	LCV_ALLO
LMB	23	2	0.0021	0.002	0.1840	0.0123	LCV_ALLO
LWC	2	1	0.0020	0.002	0.1631	0.0123	LCV_ALLO
W4T	9	2	0.0013	0.002	0.1740	0.0123	LCV_ALLO
WOL	12	8	0.0028	0.003	0.2721	0.0123	LCV_ALLO
WS	8	1	0.0020	0.002	0.1631	0.0123	LCV_ALLO
1C	8	1	0.0020	0.002	0.0040	0.5000	LCV_AQPL
BWC	2	1	0.0040	0.004	0.0080	0.5000	LCV_AQPL
IHP	11	1	0.0040	0.004	0.0080	0.5000	LCV_AQPL
LMB	23	2	0.0021	0.002	0.0045	0.5000	LCV_AQPL
LWC	2	1	0.0020	0.002	0.0040	0.5000	LCV_AQPL
W4T	9	2	0.0013	0.002	0.0043	0.5000	LCV_AQPL
WOL	12	8	0.0028	0.003	0.0067	0.5000	LCV_AQPL
WS	8	1	0.0020	0.002	0.0040	0.5000	LCV_AQPL
1C	8	1	0.0020	0.002	0.1631	0.0123	LCV_DAPH
BWC	2	1	0.0040	0.004	0.3263	0.0123	LCV_DAPH
IHP	11	1	0.0040	0.004	0.3263	0.0123	LCV_DAPH
LMB	23	2	0.0021	0.002	0.1840	0.0123	LCV_DAPH
LWC	2	1	0.0020	0.002	0.1631	0.0123	LCV_DAPH
W4T	9	2	0.0013	0.002	0.1740	0.0123	LCV_DAPH
WOL	12	8	0.0028	0.003	0.2721	0.0123	LCV_DAPH
WS	8	1	0.0020	0.002	0.1631	0.0123	LCV_DAPH
1C	8	1	0.0020	0.002	0.1059	0.0189	LCV_FISH
BWC	2	1	0.0040	0.004	0.2119	0.0189	LCV_FISH
IHP	11	1	0.0040	0.004	0.2119	0.0189	LCV_FISH
LMB	23	2	0.0021	0.002	0.1195	0.0189	LCV_FISH
LWC	2	1	0.0020	0.002	0.1059	0.0189	LCV_FISH
W4T	9	2	0.0013	0.002	0.1130	0.0189	LCV_FISH
WOL	12	8	0.0028	0.003	0.1767	0.0189	LCV_FISH
WS	8	1	0.0020	0.002	0.1059	0.0189	LCV_FISH
1C	8	1	0.0020	0.002	0.0786	0.0255	LCV_NDI
BWC	2	1	0.0040	0.004	0.1571	0.0255	LCV_NDI
IHP	11	1	0.0040	0.004	0.1571	0.0255	LCV_NDI
LMB	23	2	0.0021	0.002	0.0886	0.0255	LCV_NDI
LWC	2	1	0.0020	0.002	0.0786	0.0255	LCV_NDI
W4T	9	2	0.0013	0.002	0.0838	0.0255	LCV_NDI
WOL	12	8	0.0028	0.003	0.1310	0.0255	LCV_NDI
WS	8	1	0.0020	0.002	0.0786	0.0255	LCV_NDI
1C	8	1	0.0020	0.002	0.0909	0.0220	LTV_FISH
BWC	2	1	0.0040	0.004	0.1818	0.0220	LTV_FISH
IHP	11	1	0.0040	0.004	0.1818	0.0220	LTV_FISH
LMB	23	2	0.0021	0.002	0.1026	0.0220	LTV_FISH
LWC	2	1	0.0020	0.002	0.0909	0.0220	LTV_FISH
W4T	9	2	0.0013	0.002	0.0970	0.0220	LTV_FISH
WOL	12	8	0.0028	0.003	0.1516	0.0220	LTV_FISH
WS	8	1	0.0020	0.002	0.0909	0.0220	LTV_FISH
1C	8	1	0.0020	0.002	0.0244	0.0820	NAWQ_ACU
BWC	2	1	0.0040	0.004	0.0488	0.0820	NAWQ_ACU
IHP	11	1	0.0040	0.004	0.0488	0.0820	NAWQ_ACU
LMB	23	2	0.0021	0.002	0.0275	0.0820	NAWQ_ACU
LWC	2	1	0.0020	0.002	0.0244	0.0820	NAWQ_ACU
W4T	9	2	0.0013	0.002	0.0260	0.0820	NAWQ_ACU
WOL	12	8	0.0028	0.003	0.0407	0.0820	NAWQ_ACU
WS	8	1	0.0020	0.002	0.0244	0.0820	NAWQ_ACU
1C	8	1	0.0020	0.002	0.6250	0.0032	NAWQ_CHR
BWC	2	1	0.0040	0.004	1.2500	0.0032	NAWQ_CHR
IHP	11	1	0.0040	0.004	1.2500	0.0032	NAWQ_CHR

B-15

	LMB	23	2	0.0021	0.002	0.7051	0.0032	NAWQ_CHR
	LWC	2	1	0.0020	0.002	0.6250	0.0032	NAWQ_CHR
	W4T	9	2	0.0013	0.002	0.6667	0.0032	NAWQ_CHR
	WOL	12	8	0.0028	0.003	1.0423	0.0032	NAWQ_CHR
	WS	8	1	0.0020	0.002	0.6250	0.0032	NAWQ_CHR
MAGNESIUM	1C	9	9	9.3944	10.548	0.1286	82.0000	LCV_ALLO
	1WC	2	2	9.5050	11.936	0.1456	82.0000	LCV_ALLO
	BWC	2	2	11.6500	43.534	0.5309	82.0000	LCV_ALLO
	HRT	8	8	9.2375	10.597	0.1292	82.0000	LCV_ALLO
	IHP	12	12	9.6750	10.468	0.1277	82.0000	LCV_ALLO
	LMB	29	29	11.7938	14.053	0.1714	82.0000	LCV_ALLO
	LWC	2	2	9.1950	19.329	0.2357	82.0000	LCV_ALLO
	MWC	18	18	9.1383	9.782	0.1193	82.0000	LCV_ALLO
	NWT	2	2	8.8550	22.398	0.2731	82.0000	LCV_ALLO
	RAC	2	2	4.7600	4.886	0.0596	82.0000	LCV_ALLO
	UMB	9	9	12.4356	18.128	0.2211	82.0000	LCV_ALLO
	W4T	16	16	16.0956	17.779	0.2168	82.0000	LCV_ALLO
	WOL	13	13	8.6508	9.471	0.1155	82.0000	LCV_ALLO
	WS	10	10	8.6190	10.216	0.1246	82.0000	LCV_ALLO
	1C	9	9	9.3944	10.548	0.1286	82.0000	LCV_DAPH
	1WC	2	2	9.5050	11.936	0.1456	82.0000	LCV_DAPH
	BWC	2	2	11.6500	43.534	0.5309	82.0000	LCV_DAPH
	HRT	8	8	9.2375	10.597	0.1292	82.0000	LCV_DAPH
	IHP	12	12	9.6750	10.468	0.1277	82.0000	LCV_DAPH
	LMB	29	29	11.7938	14.053	0.1714	82.0000	LCV_DAPH
	LWC	2	2	9.1950	19.329	0.2357	82.0000	LCV_DAPH
	MWC	18	18	9.1383	9.782	0.1193	82.0000	LCV_DAPH
	NWT	2	2	8.8550	22.398	0.2731	82.0000	LCV_DAPH
	RAC	2	2	4.7600	4.886	0.0596	82.0000	LCV_DAPH
	UMB	9	9	12.4356	18.128	0.2211	82.0000	LCV_DAPH
	W4T	16	16	16.0956	17.779	0.2168	82.0000	LCV_DAPH
	WOL	13	13	8.6508	9.471	0.1155	82.0000	LCV_DAPH
	WS	10	10	8.6190	10.216	0.1246	82.0000	LCV_DAPH
MANGANESE	1C	9	9	0.0258	0.034	0.0308	1.1000	LCV_ALLO
	1WC	2	2	0.0195	0.023	0.0206	1.1000	LCV_ALLO
	BWC	2	1	0.0080	0.008	0.0073	1.1000	LCV_ALLO
	HRT	8	8	0.0444	0.051	0.0462	1.1000	LCV_ALLO
	IHP	12	12	0.0248	0.030	0.0270	1.1000	LCV_ALLO
	LMB	29	29	0.1169	0.138	0.1255	1.1000	LCV_ALLO
	LWC	2	2	0.0345	0.057	0.0515	1.1000	LCV_ALLO
	MWC	18	18	0.0292	0.031	0.0284	1.1000	LCV_ALLO
	NWT	2	2	0.0350	0.149	0.1351	1.1000	LCV_ALLO
	RAC	2	2	0.7330	5.323	4.8392	1.1000	LCV_ALLO
	UMB	9	9	0.0552	0.075	0.0678	1.1000	LCV_ALLO
	W4T	16	16	0.7425	1.378	1.2531	1.1000	LCV_ALLO
	WOL	13	13	0.1461	0.177	0.1608	1.1000	LCV_ALLO
	WS	10	10	1.6078	3.223	2.9304	1.1000	LCV_ALLO
	1C	9	9	0.0258	0.034	0.0308	1.1000	LCV_DAPH
	1WC	2	2	0.0195	0.023	0.0206	1.1000	LCV_DAPH
	BWC	2	1	0.0080	0.008	0.0073	1.1000	LCV_DAPH
	HRT	8	8	0.0444	0.051	0.0462	1.1000	LCV_DAPH
	IHP	12	12	0.0248	0.030	0.0270	1.1000	LCV_DAPH
	LMB	29	29	0.1169	0.138	0.1255	1.1000	LCV_DAPH
	LWC	2	2	0.0345	0.057	0.0515	1.1000	LCV_DAPH
	MWC	18	18	0.0292	0.031	0.0284	1.1000	LCV_DAPH
	NWT	2	2	0.0350	0.149	0.1351	1.1000	LCV_DAPH
	RAC	2	2	0.7330	5.323	4.8392	1.1000	LCV_DAPH
	UMB	9	9	0.0552	0.075	0.0678	1.1000	LCV_DAPH
	W4T	16	16	0.7425	1.378	1.2531	1.1000	LCV_DAPH
	WOL	13	13	0.1461	0.177	0.1608	1.1000	LCV_DAPH
	WS	10	10	1.6078	3.223	2.9304	1.1000	LCV_DAPH
	1C	9	9	0.0258	0.034	0.0191	1.7700	LCV_FISH
	1WC	2	2	0.0195	0.023	0.0128	1.7700	LCV_FISH
	BWC	2	1	0.0080	0.008	0.0045	1.7700	LCV_FISH
	HRT	8	8	0.0444	0.051	0.0287	1.7700	LCV_FISH
	IHP	12	12	0.0248	0.030	0.0168	1.7700	LCV_FISH
	LMB	29	29	0.1169	0.138	0.0780	1.7700	LCV_FISH
	LWC	2	2	0.0345	0.057	0.0320	1.7700	LCV_FISH
	MWC	18	18	0.0292	0.031	0.0177	1.7700	LCV_FISH

B-16

NWT	2	2	0.0350	0.149	0.0840	1.7700	LCV_FISH	
RAC	2	2	0.7330	5.323	3.0074	1.7700	LCV_FISH	
UMB	9	9	0.0552	0.075	0.0421	1.7700	LCV_FISH	
W4T	16	16	0.7425	1.378	0.7787	1.7700	LCV_FISH	
WOL	13	13	0.1461	0.177	0.1000	1.7700	LCV_FISH	
WS	10	10	1.6078	3.223	1.8211	1.7700	LCV_FISH	
1C	9	9	0.0258	0.034	0.0308	1.1000	LTV_DAPH	
1WC	2	2	0.0195	0.023	0.0206	1.1000	LTV_DAPH	
BWC	2	1	0.0080	0.008	0.0073	1.1000	LTV_DAPH	
HRT	8	8	0.0444	0.051	0.0462	1.1000	LTV_DAPH	
IHP	12	12	0.0248	0.030	0.0270	1.1000	LTV_DAPH	
LMB	29	29	0.1169	0.138	0.1255	1.1000	LTV_DAPH	
LWC	2	2	0.0345	0.057	0.0515	1.1000	LTV_DAPH	
MWC	18	18	0.0292	0.031	0.0284	1.1000	LTV_DAPH	
NWT	2	2	0.0350	0.149	0.1351	1.1000	LTV_DAPH	
RAC	2	2	0.7330	5.323	4.8392	1.1000	LTV_DAPH	
UMB	9	9	0.0552	0.075	0.0678	1.1000	LTV_DAPH	
W4T	16	16	0.7425	1.378	1.2531	1.1000	LTV_DAPH	
WOL	13	13	0.1461	0.177	0.1608	1.1000	LTV_DAPH	
WS	10	10	1.6078	3.223	2.9304	1.1000	LTV_DAPH	
1C	9	9	0.0258	0.034	0.0267	1.2700	LTV_FISH	
1WC	2	2	0.0195	0.023	0.0178	1.2700	LTV_FISH	
BWC	2	1	0.0080	0.008	0.0063	1.2700	LTV_FISH	
HRT	8	8	0.0444	0.051	0.0401	1.2700	LTV_FISH	
IHP	12	12	0.0248	0.030	0.0234	1.2700	LTV_FISH	
LMB	29	29	0.1169	0.138	0.1087	1.2700	LTV_FISH	
LWC	2	2	0.0345	0.057	0.0446	1.2700	LTV_FISH	
MWC	18	18	0.0292	0.031	0.0246	1.2700	LTV_FISH	
NWT	2	2	0.0350	0.149	0.1170	1.2700	LTV_FISH	
RAC	2	2	0.7330	5.323	4.1914	1.2700	LTV_FISH	
UMB	9	9	0.0552	0.075	0.0587	1.2700	LTV_FISH	
W4T	16	16	0.7425	1.378	1.0853	1.2700	LTV_FISH	
WOL	13	13	0.1461	0.177	0.1393	1.2700	LTV_FISH	
WS	10	10	1.6078	3.223	2.5381	1.2700	LTV_FISH	
1C	9	9	0.0258	0.034	0.0230	1.4700	S_ACU_V	
1WC	2	2	0.0195	0.023	0.0154	1.4700	S_ACU_V	
BWC	2	1	0.0080	0.008	0.0054	1.4700	S_ACU_V	
HRT	8	8	0.0444	0.051	0.0346	1.4700	S_ACU_V	
IHP	12	12	0.0248	0.030	0.0202	1.4700	S_ACU_V	
LMB	29	29	0.1169	0.138	0.0939	1.4700	S_ACU_V	
LWC	2	2	0.0345	0.057	0.0385	1.4700	S_ACU_V	
MWC	18	18	0.0292	0.031	0.0213	1.4700	S_ACU_V	
NWT	2	2	0.0350	0.149	0.1011	1.4700	S_ACU_V	
RAC	2	2	0.7330	5.323	3.6212	1.4700	S_ACU_V	
UMB	9	9	0.0552	0.075	0.0507	1.4700	S_ACU_V	
W4T	16	16	0.7425	1.378	0.9377	1.4700	S_ACU_V	
WOL	13	13	0.1461	0.177	0.1204	1.4700	S_ACU_V	
WS	10	10	1.6078	3.223	2.1928	1.4700	S_ACU_V	
1C	9	9	0.0258	0.034	0.4219	0.0803	S_CHR_V	
1WC	2	2	0.0195	0.023	0.2822	0.0803	S_CHR_V	
BWC	2	1	0.0080	0.008	0.0996	0.0803	S_CHR_V	
HRT	8	8	0.0444	0.051	0.6335	0.0803	S_CHR_V	
IHP	12	12	0.0248	0.030	0.3697	0.0803	S_CHR_V	
LMB	29	29	0.1169	0.138	1.7190	0.0803	S_CHR_V	
LWC	2	2	0.0345	0.057	0.7048	0.0803	S_CHR_V	
MWC	18	18	0.0292	0.031	0.3896	0.0803	S_CHR_V	
NWT	2	2	0.0350	0.149	1.8512	0.0803	S_CHR_V	
RAC	2	2	0.7330	5.323	66.2901	0.0803	S_CHR_V	
UMB	9	9	0.0552	0.075	0.9281	0.0803	S_CHR_V	
W4T	16	16	0.7425	1.378	17.1654	0.0803	S_CHR_V	
WOL	13	13	0.1461	0.177	2.2034	0.0803	S_CHR_V	
WS	10	10	1.6078	3.223	40.1422	0.0803	S_CHR_V	
MERCURY	LMB	17	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	MWC	13	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	WOL	10	2	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	WS	7	1	0.0000	0.000	0.0000	0.0002	LCV_ALLO
	LMB	17	1	0.0000	0.000	0.0000	0.0050	LCV_AQPL
	MWC	13	1	0.0000	0.000	0.0000	0.0050	LCV_AQPL
	WOL	10	2	0.0000	0.000	0.0000	0.0050	LCV_AQPL
	WS	7	1	0.0000	0.000	0.0000	0.0050	LCV_AQPL

B-17

	LMB	17	1	0.0000	0.000	0.0000	0.0010	LCV_DAPH
	MWC	13	1	0.0000	0.000	0.0000	0.0010	LCV_DAPH
	WOL	10	2	0.0000	0.000	0.0000	0.0010	LCV_DAPH
	WS	7	1	0.0000	0.000	0.0000	0.0010	LCV_DAPH
	LMB	17	1	0.0000	0.000	0.0000	0.0002	LCV_FISH
	MWC	13	1	0.0000	0.000	0.0000	0.0002	LCV_FISH
	WOL	10	2	0.0000	0.000	0.0000	0.0002	LCV_FISH
	WS	7	1	0.0000	0.000	0.0000	0.0002	LCV_FISH
	LMB	17	1	0.0000	0.000	0.0000	0.0009	LTV_DAPH
	MWC	13	1	0.0000	0.000	0.0000	0.0009	LTV_DAPH
	WOL	10	2	0.0000	0.000	0.0000	0.0009	LTV_DAPH
	WS	7	1	0.0000	0.000	0.0000	0.0009	LTV_DAPH
	LMB	17	1	0.0000	0.000	0.0000	0.0009	LTV_FISH
	MWC	13	1	0.0000	0.000	0.0000	0.0009	LTV_FISH
	WOL	10	2	0.0000	0.000	0.0000	0.0009	LTV_FISH
	WS	7	1	0.0000	0.000	0.0000	0.0009	LTV_FISH
	LMB	17	1	0.0000	0.000	0.0000	0.0024	NAWQ_ACU
	MWC	13	1	0.0000	0.000	0.0000	0.0024	NAWQ_ACU
	WOL	10	2	0.0000	0.000	0.0000	0.0024	NAWQ_ACU
	WS	7	1	0.0000	0.000	0.0000	0.0024	NAWQ_ACU
	LMB	17	1	0.0000	0.000	0.0000	0.0013	S_CHR_V
	MWC	13	1	0.0000	0.000	0.0000	0.0013	S_CHR_V
	WOL	10	2	0.0000	0.000	0.0000	0.0013	S_CHR_V
	WS	7	1	0.0000	0.000	0.0000	0.0013	S_CHR_V
HOLYBDENUM	RAC	2	1	0.0120	0.012	0.0136	0.8800	LCV_ALLO
	W4T	16	6	0.0152	0.019	0.0221	0.8800	LCV_ALLO
	RAC	2	1	0.0120	0.012	0.0136	0.8800	LCV_DAPH
	W4T	16	6	0.0152	0.019	0.0221	0.8800	LCV_DAPH
	RAC	2	1	0.0120	0.012	0.0333	0.3600	LTV_DAPH
	W4T	16	6	0.0152	0.019	0.0541	0.3600	LTV_DAPH
	RAC	2	1	0.0120	0.012	0.0012	10.1000	S_ACU_V
	W4T	16	6	0.0152	0.019	0.0019	10.1000	S_ACU_V
	RAC	2	1	0.0120	0.012	0.0502	0.2390	S_CHR_V
	W4T	16	6	0.0152	0.019	0.0816	0.2390	S_CHR_V
Methylene chloride	1C	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	LMB	1	1	0.0010	0.001	0.0000	42.6670	LCV_ALLO
	MWC	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	NWT	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	RAC	1	1	0.0020	0.002	0.0000	42.6670	LCV_ALLO
	UMB	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	W4T	3	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	WOL	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	WS	1	1	0.0000	0.000	0.0000	42.6670	LCV_ALLO
	1C	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	LMB	1	1	0.0010	0.001	0.0000	42.6670	LCV_DAPH
	MWC	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	NWT	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	RAC	1	1	0.0020	0.002	0.0000	42.6670	LCV_DAPH
	UMB	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	W4T	3	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	WOL	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	WS	1	1	0.0000	0.000	0.0000	42.6670	LCV_DAPH
	1C	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	LMB	1	1	0.0010	0.001	0.0000	108.0000	LCV_FISH
	MWC	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	NWT	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	RAC	1	1	0.0020	0.002	0.0000	108.0000	LCV_FISH
	UMB	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	W4T	3	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	WOL	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	WS	1	1	0.0000	0.000	0.0000	108.0000	LCV_FISH
	1C	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
	LMB	1	1	0.0010	0.001	0.0024	0.4100	LTV_FISH
	MWC	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
	NWT	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
	RAC	1	1	0.0020	0.002	0.0049	0.4100	LTV_FISH
	UMB	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
	W4T	3	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
	WOL	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH

B-18

WS	1	1	0.0000	0.000	0.0000	0.4100	LTV_FISH
1C	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
LMB	1	1	0.0010	0.001	0.0000	25.6000	S_ACU_V
MWC	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
NWT	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
RAC	1	1	0.0020	0.002	0.0001	25.6000	S_ACU_V
UMB	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
W4T	3	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
WOL	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
WS	1	1	0.0000	0.000	0.0000	25.6000	S_ACU_V
1C	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
LMB	1	1	0.0010	0.001	0.0004	2.2400	S_CHR_V
MWC	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
NWT	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
RAC	1	1	0.0020	0.002	0.0009	2.2400	S_CHR_V
UMB	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
W4T	3	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
WOL	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V
WS	1	1	0.0000	0.000	0.0000	2.2400	S_CHR_V

NICKEL

LMB	29	1	0.0070	0.007	1.4000	0.0050	LCV_ALLO
W4T	16	5	0.0136	0.015	3.0380	0.0050	LCV_ALLO
WS	10	4	0.0104	0.014	2.8210	0.0050	LCV_ALLO
LMB	29	1	0.0070	0.007	1.4000	0.0050	LCV_AQPL
W4T	16	5	0.0136	0.015	3.0380	0.0050	LCV_AQPL
WS	10	4	0.0104	0.014	2.8210	0.0050	LCV_AQPL
LMB	29	1	0.0070	0.007	1.4000	0.0050	LCV_DAPH
W4T	16	5	0.0136	0.015	3.0380	0.0050	LCV_DAPH
WS	10	4	0.0104	0.014	2.8210	0.0050	LCV_DAPH
LMB	29	1	0.0070	0.007	0.2000	0.0350	LCV_FISH
W4T	16	5	0.0136	0.015	0.4340	0.0350	LCV_FISH
WS	10	4	0.0104	0.014	0.4030	0.0350	LCV_FISH
LMB	29	1	0.0070	0.007	0.0545	0.1284	LCV_NDI
W4T	16	5	0.0136	0.015	0.1183	0.1284	LCV_NDI
WS	10	4	0.0104	0.014	0.1099	0.1284	LCV_NDI
LMB	29	1	0.0070	0.007	0.1556	0.0450	LTV_DAPH
W4T	16	5	0.0136	0.015	0.3376	0.0450	LTV_DAPH
WS	10	4	0.0104	0.014	0.3134	0.0450	LTV_DAPH
LMB	29	1	0.0070	0.007	0.1129	0.0620	LTV_FISH
W4T	16	5	0.0136	0.015	0.2450	0.0620	LTV_FISH
WS	10	4	0.0104	0.014	0.2275	0.0620	LTV_FISH
LMB	29	1	0.0070	0.007	0.0050	1.4000	NAWQ_ACU
W4T	16	5	0.0136	0.015	0.0108	1.4000	NAWQ_ACU
WS	10	4	0.0104	0.014	0.0101	1.4000	NAWQ_ACU
LMB	29	1	0.0070	0.007	0.0438	0.1600	NAWQ_CHR
W4T	16	5	0.0136	0.015	0.0949	0.1600	NAWQ_CHR
WS	10	4	0.0104	0.014	0.0882	0.1600	NAWQ_CHR

POTASSIUM

1C	9	7	0.9363	1.066	0.0201	53.0000	LCV_ALLO
1WC	2	2	1.6850	2.032	0.0383	53.0000	LCV_ALLO
BWC	2	1	0.8870	0.887	0.0167	53.0000	LCV_ALLO
HRT	8	8	1.8813	2.108	0.0398	53.0000	LCV_ALLO
IHP	12	11	1.8167	2.057	0.0388	53.0000	LCV_ALLO
LMB	29	28	2.0054	2.276	0.0429	53.0000	LCV_ALLO
LWC	2	1	1.2000	1.200	0.0226	53.0000	LCV_ALLO
MWC	18	17	1.7764	1.980	0.0374	53.0000	LCV_ALLO
NWT	2	1	1.5900	1.590	0.0300	53.0000	LCV_ALLO
RAC	2	1	0.8990	0.899	0.0170	53.0000	LCV_ALLO
UMB	9	9	2.5233	3.478	0.0656	53.0000	LCV_ALLO
W4T	16	16	7.0494	9.129	0.1722	53.0000	LCV_ALLO
WOL	13	12	1.7679	2.005	0.0378	53.0000	LCV_ALLO
WS	10	8	1.7448	2.089	0.0394	53.0000	LCV_ALLO
1C	9	7	0.9363	1.066	0.0201	53.0000	LCV_DAPH
1WC	2	2	1.6850	2.032	0.0383	53.0000	LCV_DAPH
BWC	2	1	0.8870	0.887	0.0167	53.0000	LCV_DAPH
HRT	8	8	1.8813	2.108	0.0398	53.0000	LCV_DAPH
IHP	12	11	1.8167	2.057	0.0388	53.0000	LCV_DAPH
LMB	29	28	2.0054	2.276	0.0429	53.0000	LCV_DAPH
LWC	2	1	1.2000	1.200	0.0226	53.0000	LCV_DAPH
MWC	18	17	1.7764	1.980	0.0374	53.0000	LCV_DAPH
NWT	2	1	1.5900	1.590	0.0300	53.0000	LCV_DAPH

B-19

	RAC	2	1	0.8990	0.899	0.0170	53.0000	LCV_DAPH
	UMB	9	9	2.5233	3.478	0.0656	53.0000	LCV_DAPH
	W4T	16	16	7.0494	9.129	0.1722	53.0000	LCV_DAPH
	WOL	13	12	1.7679	2.005	0.0378	53.0000	LCV_DAPH
	WS	10	8	1.7448	2.089	0.0394	53.0000	LCV_DAPH
SILVER	BWC	2	1	0.0040	0.004	33.3333	0.0001	LCV_ALLO
	IHP	12	1	0.0040	0.004	33.3333	0.0001	LCV_ALLO
	W4T	16	1	0.0040	0.004	33.3333	0.0001	LCV_ALLO
	BWC	2	1	0.0040	0.004	0.1333	0.0300	LCV_AQPL
	IHP	12	1	0.0040	0.004	0.1333	0.0300	LCV_AQPL
	W4T	16	1	0.0040	0.004	0.1333	0.0300	LCV_AQPL
	BWC	2	1	0.0040	0.004	1.5385	0.0026	LCV_DAPH
	IHP	12	1	0.0040	0.004	1.5385	0.0026	LCV_DAPH
	W4T	16	1	0.0040	0.004	1.5385	0.0026	LCV_DAPH
	BWC	2	1	0.0040	0.004	33.3333	0.0001	LCV_FISH
	IHP	12	1	0.0040	0.004	33.3333	0.0001	LCV_FISH
	W4T	16	1	0.0040	0.004	33.3333	0.0001	LCV_FISH
	BWC	2	1	0.0040	0.004	7.1429	0.0006	LTV_DAPH
	IHP	12	1	0.0040	0.004	7.1429	0.0006	LTV_DAPH
	W4T	16	1	0.0040	0.004	7.1429	0.0006	LTV_DAPH
	BWC	2	1	0.0040	0.004	20.0000	0.0002	LTV_FISH
	IHP	12	1	0.0040	0.004	20.0000	0.0002	LTV_FISH
	W4T	16	1	0.0040	0.004	20.0000	0.0002	LTV_FISH
	BWC	2	1	0.0040	0.004	0.9756	0.0041	NAWQ_ACU
	IHP	12	1	0.0040	0.004	0.9756	0.0041	NAWQ_ACU
	W4T	16	1	0.0040	0.004	0.9756	0.0041	NAWQ_ACU
	BWC	2	1	0.0040	0.004	11.1111	0.0004	S_CHR_V
	IHP	12	1	0.0040	0.004	11.1111	0.0004	S_CHR_V
	W4T	16	1	0.0040	0.004	11.1111	0.0004	S_CHR_V
SODIUM	1C	9	9	4.5044	6.144	0.0090	680.0000	LCV_ALLO
	1WC	2	2	18.8500	40.001	0.0588	680.0000	LCV_ALLO
	BWC	2	1	0.5230	0.523	0.0008	680.0000	LCV_ALLO
	HRT	8	8	5.0038	5.694	0.0084	680.0000	LCV_ALLO
	IHP	12	12	16.4375	19.317	0.0284	680.0000	LCV_ALLO
	LMB	29	29	7.4876	8.950	0.0132	680.0000	LCV_ALLO
	LWC	2	2	17.4800	72.536	0.1067	680.0000	LCV_ALLO
	MWC	18	18	15.6189	18.004	0.0265	680.0000	LCV_ALLO
	NWT	2	2	3.4500	6.354	0.0093	680.0000	LCV_ALLO
	RAC	2	2	2.3700	3.570	0.0052	680.0000	LCV_ALLO
	UMB	9	9	7.6311	11.164	0.0164	680.0000	LCV_ALLO
	W4T	16	16	13.3956	15.115	0.0222	680.0000	LCV_ALLO
	WOL	13	13	13.3846	15.932	0.0234	680.0000	LCV_ALLO
	WS	10	10	26.6400	41.874	0.0616	680.0000	LCV_ALLO
	1C	9	9	4.5044	6.144	0.0090	680.0000	LCV_DAPH
	1WC	2	2	18.8500	40.001	0.0588	680.0000	LCV_DAPH
	BWC	2	1	0.5230	0.523	0.0008	680.0000	LCV_DAPH
	HRT	8	8	5.0038	5.694	0.0084	680.0000	LCV_DAPH
	IHP	12	12	16.4375	19.317	0.0284	680.0000	LCV_DAPH
	LMB	29	29	7.4876	8.950	0.0132	680.0000	LCV_DAPH
	LWC	2	2	17.4800	72.536	0.1067	680.0000	LCV_DAPH
	MWC	18	18	15.6189	18.004	0.0265	680.0000	LCV_DAPH
	NWT	2	2	3.4500	6.354	0.0093	680.0000	LCV_DAPH
	RAC	2	2	2.3700	3.570	0.0052	680.0000	LCV_DAPH
	UMB	9	9	7.6311	11.164	0.0164	680.0000	LCV_DAPH
	W4T	16	16	13.3956	15.115	0.0222	680.0000	LCV_DAPH
	WOL	13	13	13.3846	15.932	0.0234	680.0000	LCV_DAPH
	WS	10	10	26.6400	41.874	0.0616	680.0000	LCV_DAPH
STRONTIUM	1C	9	9	0.0829	0.095	0.0023	42.0000	LCV_ALLO
	1WC	2	2	0.0925	0.134	0.0032	42.0000	LCV_ALLO
	BWC	2	1	0.0360	0.036	0.0009	42.0000	LCV_ALLO
	HRT	8	8	0.0941	0.107	0.0026	42.0000	LCV_ALLO
	IHP	12	12	0.0973	0.106	0.0025	42.0000	LCV_ALLO
	LMB	29	29	0.1344	0.156	0.0037	42.0000	LCV_ALLO
	LWC	2	2	0.1000	0.207	0.0049	42.0000	LCV_ALLO
	MWC	18	18	0.0924	0.100	0.0024	42.0000	LCV_ALLO
	NWT	2	2	0.0965	0.169	0.0040	42.0000	LCV_ALLO
	RAC	2	2	0.1250	0.232	0.0055	42.0000	LCV_ALLO
	UMB	9	9	0.1467	0.200	0.0048	42.0000	LCV_ALLO

B-20

W4T	16	16	0.1990	0.230	0.0055	42.0000	LCV_ALLO
WOL	13	13	0.0960	0.105	0.0025	42.0000	LCV_ALLO
WS	10	10	0.0938	0.116	0.0028	42.0000	LCV_ALLO
1C	9	9	0.0829	0.095	0.0023	42.0000	LCV_DAPH
1WC	2	2	0.0925	0.134	0.0032	42.0000	LCV_DAPH
BWC	2	1	0.0360	0.036	0.0009	42.0000	LCV_DAPH
HRT	8	8	0.0941	0.107	0.0026	42.0000	LCV_DAPH
IHP	12	12	0.0973	0.106	0.0025	42.0000	LCV_DAPH
LMB	29	29	0.1344	0.156	0.0037	42.0000	LCV_DAPH
LWC	2	2	0.1000	0.207	0.0049	42.0000	LCV_DAPH
MWC	18	18	0.0924	0.100	0.0024	42.0000	LCV_DAPH
NWT	2	2	0.0965	0.169	0.0040	42.0000	LCV_DAPH
RAC	2	2	0.1250	0.232	0.0055	42.0000	LCV_DAPH
UMB	9	9	0.1467	0.200	0.0048	42.0000	LCV_DAPH
W4T	16	16	0.1990	0.230	0.0055	42.0000	LCV_DAPH
WOL	13	13	0.0960	0.105	0.0025	42.0000	LCV_DAPH
WS	10	10	0.0938	0.116	0.0028	42.0000	LCV_DAPH
1C	9	9	0.0829	0.095	0.0156	6.1000	S_ACU_V
1WC	2	2	0.0925	0.134	0.0219	6.1000	S_ACU_V
BWC	2	1	0.0360	0.036	0.0059	6.1000	S_ACU_V
HRT	8	8	0.0941	0.107	0.0176	6.1000	S_ACU_V
IHP	12	12	0.0973	0.106	0.0174	6.1000	S_ACU_V
LMB	29	29	0.1344	0.156	0.0256	6.1000	S_ACU_V
LWC	2	2	0.1000	0.207	0.0340	6.1000	S_ACU_V
MWC	18	18	0.0924	0.100	0.0163	6.1000	S_ACU_V
NWT	2	2	0.0965	0.169	0.0277	6.1000	S_ACU_V
RAC	2	2	0.1250	0.232	0.0381	6.1000	S_ACU_V
UMB	9	9	0.1467	0.200	0.0327	6.1000	S_ACU_V
W4T	16	16	0.1990	0.230	0.0377	6.1000	S_ACU_V
WOL	13	13	0.0960	0.105	0.0172	6.1000	S_ACU_V
WS	10	10	0.0938	0.116	0.0191	6.1000	S_ACU_V
1C	9	9	0.0829	0.095	0.1535	0.6200	S_CHR_V
1WC	2	2	0.0925	0.134	0.2154	0.6200	S_CHR_V
BWC	2	1	0.0360	0.036	0.0581	0.6200	S_CHR_V
HRT	8	8	0.0941	0.107	0.1731	0.6200	S_CHR_V
IHP	12	12	0.0973	0.106	0.1707	0.6200	S_CHR_V
LMB	29	29	0.1344	0.156	0.2519	0.6200	S_CHR_V
LWC	2	2	0.1000	0.207	0.3344	0.6200	S_CHR_V
MWC	18	18	0.0924	0.100	0.1607	0.6200	S_CHR_V
NWT	2	2	0.0965	0.169	0.2728	0.6200	S_CHR_V
RAC	2	2	0.1250	0.232	0.3747	0.6200	S_CHR_V
UMB	9	9	0.1467	0.200	0.3221	0.6200	S_CHR_V
W4T	16	16	0.1990	0.230	0.3711	0.6200	S_CHR_V
WOL	13	13	0.0960	0.105	0.1691	0.6200	S_CHR_V
WS	10	10	0.0938	0.116	0.1877	0.6200	S_CHR_V
Trichloroethene	RAC	1	0.0010	0.001	0.0001	7.2570	LCV_ALLO
	W4T	3	0.0010	0.001	0.0001	7.2570	LCV_ALLO
	RAC	1	0.0010	0.001	0.0001	7.2570	LCV_DAPH
	W4T	3	0.0010	0.001	0.0001	7.2570	LCV_DAPH
	RAC	1	0.0010	0.001	0.0001	14.8670	LCV_FISH
	W4T	3	0.0010	0.001	0.0001	14.8670	LCV_FISH
	RAC	1	0.0010	0.001	0.0002	5.7580	LTV_FISH
	W4T	3	0.0010	0.001	0.0002	5.7580	LTV_FISH
	RAC	1	0.0010	0.001	0.0002	4.3500	S_ACU_V
	W4T	3	0.0010	0.001	0.0002	4.3500	S_ACU_V
	RAC	1	0.0010	0.001	0.0022	0.4650	S_CHR_V
	W4T	3	0.0010	0.001	0.0022	0.4650	S_CHR_V
VANADIUM	IHP	12	0.0040	0.004	0.0500	0.0800	LCV_ALLO
	LMB	29	0.0020	0.002	0.0250	0.0800	LCV_ALLO
	UMB	9	0.0020	0.002	0.0250	0.0800	LCV_ALLO
	WOL	13	0.0033	0.004	0.0480	0.0800	LCV_ALLO
	IHP	12	0.0040	0.004	0.0043	0.9400	LCV_DAPH
	LMB	29	0.0020	0.002	0.0021	0.9400	LCV_DAPH
	UMB	9	0.0020	0.002	0.0021	0.9400	LCV_DAPH
	WOL	13	0.0033	0.004	0.0041	0.9400	LCV_DAPH
	IHP	12	0.0040	0.004	0.0500	0.0800	LCV_FISH
	LMB	29	0.0020	0.002	0.0250	0.0800	LCV_FISH
	UMB	9	0.0020	0.002	0.0250	0.0800	LCV_FISH
	WOL	13	0.0033	0.004	0.0480	0.0800	LCV_FISH

B-21

	IHP	12	1	0.0040	0.004	0.0093	0.4300	LTV_DAPH
	LMB	29	1	0.0020	0.002	0.0047	0.4300	LTV_DAPH
	UMB	9	1	0.0020	0.002	0.0047	0.4300	LTV_DAPH
	WOL	13	2	0.0033	0.004	0.0089	0.4300	LTV_DAPH
	IHP	12	1	0.0040	0.004	0.0976	0.0410	LTV_FISH
	LMB	29	1	0.0020	0.002	0.0488	0.0410	LTV_FISH
	UMB	9	1	0.0020	0.002	0.0488	0.0410	LTV_FISH
	WOL	13	2	0.0033	0.004	0.0936	0.0410	LTV_FISH
	IHP	12	1	0.0040	0.004	0.0141	0.2840	S_ACU_V
	LMB	29	1	0.0020	0.002	0.0070	0.2840	S_ACU_V
	UMB	9	1	0.0020	0.002	0.0070	0.2840	S_ACU_V
	WOL	13	2	0.0033	0.004	0.0135	0.2840	S_ACU_V
	IHP	12	1	0.0040	0.004	0.2094	0.0191	S_CHR_V
	LMB	29	1	0.0020	0.002	0.1047	0.0191	S_CHR_V
	UMB	9	1	0.0020	0.002	0.1047	0.0191	S_CHR_V
	WOL	13	2	0.0033	0.004	0.2010	0.0191	S_CHR_V
Vinyl chloride	W4T	3	1	0.0000	0.000	0.0000	28.8790	LCV_ALLO
	W4T	3	1	0.0000	0.000	0.0000	28.8790	LCV_FISH
	W4T	3	1	0.0000	0.000	0.0000	14.5200	LTV_FISH
	W4T	3	1	0.0000	0.000	0.0000	1.5700	S_ACU_V
	W4T	3	1	0.0000	0.000	0.0000	0.0878	S_CHR_V
ZINC	1C	9	4	0.0193	0.028	0.9435	0.0300	LCV_ALLO
	1WC	2	2	0.0660	0.262	8.7242	0.0300	LCV_ALLO
	HRT	8	3	0.0044	0.007	0.2177	0.0300	LCV_ALLO
	IHP	11	9	0.0373	0.045	1.4882	0.0300	LCV_ALLO
	LMB	28	18	0.0124	0.014	0.4730	0.0300	LCV_ALLO
	MWC	17	11	0.0251	0.027	0.9161	0.0300	LCV_ALLO
	UMB	9	6	0.0569	0.114	3.7985	0.0300	LCV_ALLO
	W4T	16	6	0.0083	0.013	0.4240	0.0300	LCV_ALLO
	WOL	12	7	0.0363	0.053	1.7608	0.0300	LCV_ALLO
	WS	8	5	0.0083	0.015	0.5070	0.0300	LCV_ALLO
	1C	9	4	0.0193	0.028	0.9435	0.0300	LCV_AQPL
	1WC	2	2	0.0660	0.262	8.7242	0.0300	LCV_AQPL
	HRT	8	3	0.0044	0.007	0.2177	0.0300	LCV_AQPL
	IHP	11	9	0.0373	0.045	1.4882	0.0300	LCV_AQPL
	LMB	28	18	0.0124	0.014	0.4730	0.0300	LCV_AQPL
	MWC	17	11	0.0251	0.027	0.9161	0.0300	LCV_AQPL
	UMB	9	6	0.0569	0.114	3.7985	0.0300	LCV_AQPL
	W4T	16	6	0.0083	0.013	0.4240	0.0300	LCV_AQPL
	WOL	12	7	0.0363	0.053	1.7608	0.0300	LCV_AQPL
	WS	8	5	0.0083	0.015	0.5070	0.0300	LCV_AQPL
	1C	9	4	0.0193	0.028	0.6057	0.0467	LCV_DAPH
	1WC	2	2	0.0660	0.262	5.6008	0.0467	LCV_DAPH
	HRT	8	3	0.0044	0.007	0.1398	0.0467	LCV_DAPH
	IHP	11	9	0.0373	0.045	0.9554	0.0467	LCV_DAPH
	LMB	28	18	0.0124	0.014	0.3037	0.0467	LCV_DAPH
	MWC	17	11	0.0251	0.027	0.5881	0.0467	LCV_DAPH
	UMB	9	6	0.0569	0.114	2.4386	0.0467	LCV_DAPH
	W4T	16	6	0.0083	0.013	0.2722	0.0467	LCV_DAPH
	WOL	12	7	0.0363	0.053	1.1304	0.0467	LCV_DAPH
	WS	8	5	0.0083	0.015	0.3255	0.0467	LCV_DAPH
	1C	9	4	0.0193	0.028	0.7774	0.0364	LCV_FISH
	1WC	2	2	0.0660	0.262	7.1883	0.0364	LCV_FISH
	HRT	8	3	0.0044	0.007	0.1794	0.0364	LCV_FISH
	IHP	11	9	0.0373	0.045	1.2262	0.0364	LCV_FISH
	LMB	28	18	0.0124	0.014	0.3898	0.0364	LCV_FISH
	MWC	17	11	0.0251	0.027	0.7548	0.0364	LCV_FISH
	UMB	9	6	0.0569	0.114	3.1298	0.0364	LCV_FISH
	W4T	16	6	0.0083	0.013	0.3494	0.0364	LCV_FISH
	WOL	12	7	0.0363	0.053	1.4508	0.0364	LCV_FISH
	WS	8	5	0.0083	0.015	0.4177	0.0364	LCV_FISH
	1C	9	4	0.0193	0.028	0.0054	5.2430	LCV_NDI
	1WC	2	2	0.0660	0.262	0.0499	5.2430	LCV_NDI
	HRT	8	3	0.0044	0.007	0.0012	5.2430	LCV_NDI
	IHP	11	9	0.0373	0.045	0.0085	5.2430	LCV_NDI
	LMB	28	18	0.0124	0.014	0.0027	5.2430	LCV_NDI
	MWC	17	11	0.0251	0.027	0.0052	5.2430	LCV_NDI
	UMB	9	6	0.0569	0.114	0.0217	5.2430	LCV_NDI
	W4T	16	6	0.0083	0.013	0.0024	5.2430	LCV_NDI

B-22

WOL	12	7	0.0363	0.053	0.0101	5.2430	LCV_NDI
WS	8	5	0.0083	0.015	0.0029	5.2430	LCV_NDI
1C	9	4	0.0193	0.028	0.6022	0.0470	LTV_FISH
1WC	2	2	0.0660	0.262	5.5686	0.0470	LTV_FISH
HRT	8	3	0.0044	0.007	0.1390	0.0470	LTV_FISH
IHP	11	9	0.0373	0.045	0.9499	0.0470	LTV_FISH
LMB	28	18	0.0124	0.014	0.3019	0.0470	LTV_FISH
MWC	17	11	0.0251	0.027	0.5847	0.0470	LTV_FISH
UMB	9	6	0.0569	0.114	2.4246	0.0470	LTV_FISH
W4T	16	6	0.0083	0.013	0.2706	0.0470	LTV_FISH
WOL	12	7	0.0363	0.053	1.1239	0.0470	LTV_FISH
WS	8	5	0.0083	0.015	0.3236	0.0470	LTV_FISH
1C	9	4	0.0193	0.028	0.2359	0.1200	NAWQ_ACU
1WC	2	2	0.0660	0.262	2.1811	0.1200	NAWQ_ACU
HRT	8	3	0.0044	0.007	0.0544	0.1200	NAWQ_ACU
IHP	11	9	0.0373	0.045	0.3721	0.1200	NAWQ_ACU
LMB	28	18	0.0124	0.014	0.1183	0.1200	NAWQ_ACU
MWC	17	11	0.0251	0.027	0.2290	0.1200	NAWQ_ACU
UMB	9	6	0.0569	0.114	0.9496	0.1200	NAWQ_ACU
W4T	16	6	0.0083	0.013	0.1060	0.1200	NAWQ_ACU
WOL	12	7	0.0363	0.053	0.4402	0.1200	NAWQ_ACU
WS	8	5	0.0083	0.015	0.1267	0.1200	NAWQ_ACU
1C	9	4	0.0193	0.028	0.2573	0.1100	NAWQ_CHR
1WC	2	2	0.0660	0.262	2.3793	0.1100	NAWQ_CHR
HRT	8	3	0.0044	0.007	0.0594	0.1100	NAWQ_CHR
IHP	11	9	0.0373	0.045	0.4059	0.1100	NAWQ_CHR
LMB	28	18	0.0124	0.014	0.1290	0.1100	NAWQ_CHR
MWC	17	11	0.0251	0.027	0.2498	0.1100	NAWQ_CHR
UMB	9	6	0.0569	0.114	1.0360	0.1100	NAWQ_CHR
W4T	16	6	0.0083	0.013	0.1156	0.1100	NAWQ_CHR
WOL	12	7	0.0363	0.053	0.4802	0.1100	NAWQ_CHR
WS	8	5	0.0083	0.015	0.1383	0.1100	NAWQ_CHR

Table B.2. Screening of aqueous concentrations of chemicals detected in seeps and ephemeral tributaries against chronic NAWQC or Secondary Chronic Values (Suter and Mabrey 1996). ALL concentrations are mg/L.

Chemical	Reach	Seep	Samples	Detects	P.L. Mean	95% UCB	Quotient	Benchmark Value	Benchmark
1,1,1-Trichloroethane	MWC	SH2-3	2	1	0.0000	0.000	0.000	0.0621	S_CHR_V
	W4T LMB	BTT MID. DRAIN.	1 1	1 1	0.0010 0.0010	0.001 0.001	0.021 0.021	0.0466 0.0466	S_CHR_V S_CHR_V
ALUMINIUM	W4T	BTT	7	4	0.0336	0.040	0.461	0.0870	NAWQ_CHR
	LVC	EAST SEEP	9	9	0.8376	1.262	14.508	0.0870	NAWQ_CHR
	LMB	MBTRIB-3	2	2	0.1730	0.735	8.447	0.0870	NAWQ_CHR
	LMB	MID. DRAIN.	10	6	0.0713	0.108	1.242	0.0870	NAWQ_CHR
	LVC	MV-1	2	2	0.1990	0.275	3.158	0.0870	NAWQ_CHR
	UMB	MV-3	1	1	0.1620	0.162	1.862	0.0870	NAWQ_CHR
	WOL	RS-1	1	1	0.2980	0.298	3.425	0.0870	NAWQ_CHR
	WOL	RS-3A	10	10	10.0120	12.730	146.319	0.0870	NAWQ_CHR
	WOL	RS-3B	2	2	1.3450	1.819	20.903	0.0870	NAWQ_CHR
	MHC	SW2-1	1	1	0.1860	0.186	2.138	0.0870	NAWQ_CHR
	LMB	SW2-5	10	6	0.0686	0.098	1.124	0.0870	NAWQ_CHR
	Unknown	SW2-6	3	2	0.0753	0.201	2.313	0.0870	NAWQ_CHR
	Unknown	SW2-7	4	2	0.0380	0.038	0.437	0.0870	NAWQ_CHR
	W4T	SW4-2	6	2	0.0318	0.041	0.477	0.0870	NAWQ_CHR
	LMB	SW5-2	1	1	0.0780	0.078	0.897	0.0870	NAWQ_CHR
	LMB	SW5-4	6	2	0.0473	0.051	0.584	0.0870	NAWQ_CHR
	WOL	SW7-2	2	2	0.5740	1.066	12.258	0.0870	NAWQ_CHR
	LVC	SW7-3	8	8	0.4520	0.548	6.299	0.0870	NAWQ_CHR
	LVC	SW7-5	10	10	0.8017	1.122	12.898	0.0870	NAWQ_CHR
	LVC	SW7-6	1	1	0.7610	0.761	8.747	0.0870	NAWQ_CHR
	LVC	SW7-8	1	1	0.4120	0.412	4.736	0.0870	NAWQ_CHR
	HRT	SW9-2	3	2	0.9780	1.791	20.581	0.0870	NAWQ_CHR
	W4T	WAG4 MS1	11	10	0.1551	0.313	3.602	0.0870	NAWQ_CHR
	W4T	WAG4 T2A	7	7	0.4713	0.710	8.163	0.0870	NAWQ_CHR
	LVC	WCTRIB-1	2	1	0.9570	0.957	11.000	0.0870	NAWQ_CHR
	MHC	WCTRIB-2	1	1	0.1940	0.194	2.230	0.0870	NAWQ_CHR
	EVC	WOCET	2	2	1.0710	3.780	43.444	0.0870	NAWQ_CHR
ANTIMONY	W4T	SW4-1	2	1	0.0450	0.045	0.433	0.1040	S_CHR_V
ARSENIC	LVC	EAST SEEP	8	2	0.0016	0.003	0.016	0.1900	NAWQ_CHR
	LMB	MID. DRAIN.	7	2	0.0010	0.001	0.005	0.1900	NAWQ_CHR
	LVC	MV-1	2	1	0.0020	0.002	0.011	0.1900	NAWQ_CHR
	WOL	RS-3A	8	8	0.0169	0.020	0.107	0.1900	NAWQ_CHR
	Unknown	SW2-6	2	1	0.0010	0.001	0.005	0.1900	NAWQ_CHR
	Unknown	SW2-7	3	1	0.0010	0.001	0.005	0.1900	NAWQ_CHR
	W4T	SW4-1	2	1	0.0060	0.006	0.032	0.1900	NAWQ_CHR
	LVC	SW7-3	7	2	0.0030	0.003	0.016	0.1900	NAWQ_CHR

Acetone

IHP	SNNT	1	1	0.0080	0.008	0.001	11.2000	S_CHR_V
LWC	MV-1	1	1	0.0010	0.001	0.000	11.2000	S_CHR_V
LWC	MV-2	1	1	0.0030	0.003	0.000	11.2000	S_CHR_V
MHC	SH2-1	1	1	0.0110	0.011	0.000	11.2000	S_CHR_V
UMC	SH2-2	1	1	0.0010	0.001	0.000	11.2000	S_CHR_V
MHC	SH2-4	1	1	0.0140	0.014	0.000	11.2000	S_CHR_V
WOL	SH6-1	1	1	0.0030	0.003	0.000	11.2000	S_CHR_V
LWC	SH7-5	1	1	0.0140	0.014	0.001	11.2000	S_CHR_V
MHC	WCTRIB-2	1	1	0.0020	0.002	0.000	11.2000	S_CHR_V
ENC	WOCET	1	1	0.0090	0.009	0.001	11.2000	S_CHR_V

BARIUM

W4T	BTT	7	7	0.2804	0.309	81.285	0.0038	S_CHR_V
LWC	EAST SEEP	9	7	0.0312	0.036	9.479	0.0038	S_CHR_V
UMB	MBTRIB-2A	1	1	0.0500	0.050	13.158	0.0038	S_CHR_V
LMB	MBTRIB-3	2	2	0.0570	0.133	34.938	0.0038	S_CHR_V
LWC	MID. DRAIN.	10	10	0.0914	0.113	29.641	0.0038	S_CHR_V
UMB	MV-1	2	2	0.1505	0.160	42.098	0.0038	S_CHR_V
UMB	MV-3	1	1	0.0490	0.049	12.895	0.0038	S_CHR_V
WOL	RS-1	1	1	0.0360	0.036	9.474	0.0038	S_CHR_V
WOL	RS-3A	10	8	0.0515	0.061	16.107	0.0038	S_CHR_V
WOL	RS-3B	2	1	0.0290	0.029	7.632	0.0038	S_CHR_V
MHC	SH2-1	1	1	0.0840	0.084	22.105	0.0038	S_CHR_V
UMC	SH2-2	1	1	0.1040	0.104	27.368	0.0038	S_CHR_V
MHC	SH2-3	1	1	0.0380	0.038	10.000	0.0038	S_CHR_V
MHC	SH2-4	2	2	0.0935	0.324	85.251	0.0038	S_CHR_V
LMB	SH2-5	10	10	0.2801	0.301	79.263	0.0038	S_CHR_V
Unknown	SH2-6	3	3	0.1530	0.168	44.256	0.0038	S_CHR_V
Unknown	SH2-7	4	4	0.1308	0.140	36.864	0.0038	S_CHR_V
W4T	SH4-1	2	2	0.2455	0.362	95.343	0.0038	S_CHR_V
W4T	SH4-2	6	6	0.3733	0.428	112.569	0.0038	S_CHR_V
LMB	SH5-2	1	1	0.0640	0.064	16.842	0.0038	S_CHR_V
LMB	SH5-4	6	6	0.1057	0.127	33.423	0.0038	S_CHR_V
WOL	SH7-1	2	2	0.0720	0.078	20.609	0.0038	S_CHR_V
WOL	SH7-2	2	1	0.0320	0.032	8.421	0.0038	S_CHR_V
LWC	SH7-3	8	3	0.0207	0.022	5.771	0.0038	S_CHR_V
LWC	SH7-5	10	10	0.0546	0.070	18.538	0.0038	S_CHR_V
LWC	SH7-6	1	1	0.0490	0.049	12.895	0.0038	S_CHR_V
HRT	SH7-8	1	1	0.0300	0.030	7.895	0.0038	S_CHR_V
HRT	SH9-1	3	3	0.1360	0.147	38.560	0.0038	S_CHR_V
HRT	SH9-2	3	3	0.2117	0.287	75.593	0.0038	S_CHR_V
W4T	WAG4 MS1	11	10	0.1425	0.154	40.471	0.0038	S_CHR_V
W4T	WAG4 T2A	7	7	0.0941	0.106	27.767	0.0038	S_CHR_V
LWC	WCTRIB-1	2	2	0.0545	0.140	36.773	0.0038	S_CHR_V
MHC	WCTRIB-2	1	1	0.0380	0.038	10.000	0.0038	S_CHR_V
IHP	WCTRIB-3	1	1	0.0620	0.062	16.316	0.0038	S_CHR_V
ENC	WOCET	2	2	0.0490	0.112	29.510	0.0038	S_CHR_V
W4T	BTT	7	1	0.0000	0.000	0.000	0.0051	S_CHR_V
LWC	EAST SEEP	9	1	0.0000	0.000	0.000	0.0051	S_CHR_V
LMB	MID. DRAIN.	10	2	0.0000	0.000	0.000	0.0051	S_CHR_V
WOL	RS-3A	10	5	0.0004	0.001	0.141	0.0051	S_CHR_V

BERYLLIUM

Element	Sample ID	Count	Efficiency	Concentration	Unit	Method	Notes
BORON	LMB	10	0.0000	0.000	0.000	SW2-5	
	Unknown	3	0.0000	0.000	0.000	SW2-6	
	Unknown	4	0.0000	0.000	0.000	SW2-7	
	W4T	6	0.0000	0.000	0.000	SW4-2	
	LWC	8	0.0000	0.000	0.000	SW7-3	
	LWC	10	0.0010	0.001	0.196	SW7-5	
	W4T	11	0.0002	0.001	0.111	WAG4 MS1	
	W4T	7	0.0000	0.000	0.000	WAG4 T2A	
	W4T	7	0.0826	0.101	0.185	BTT	
	LWC	9	0.0528	0.079	0.145	EAST SEEP	
	LMB	10	0.1654	0.201	0.367	MID. DRAIN.	
	WOL	10	0.0924	0.107	0.195	RS-3A	
	WOL	2	0.0340	0.034	0.062	RS-3B	
	MWC	1	0.0190	0.019	0.035	SH2-1	
	LMB	5	0.1890	0.237	0.434	SH2-5	
	Unknown	3	0.1170	0.120	0.219	SH2-6	
	Unknown	4	0.1397	0.141	0.259	SH2-7	
	W4T	6	0.1368	0.181	0.331	SH4-2	
	LMB	1	0.1750	0.175	0.320	SH5-2	
	LMB	6	0.1307	0.178	0.325	SH5-4	
LWC	8	0.0743	0.087	0.160	SW7-3		
LWC	10	0.0557	0.062	0.113	SW7-5		
W4T	11	0.1559	0.174	0.317	WAG4 MS1		
W4T	7	0.1293	0.169	0.309	WAG4 T2A		
IHP	1	0.1990	0.199	0.364	VCTRIB-3		
CADMIUM	W4T	6	0.0000	0.000	0.000	BTT	
	LMB	9	0.0000	0.000	0.000	MID. DRAIN.	
	WOL	8	0.0005	0.002	1.509	RS-3A	
	LMB	7	0.0000	0.000	0.000	SW2-5	
	W4T	5	0.0000	0.000	0.000	SW4-2	
	LMB	5	0.0020	0.002	1.818	SW5-4	
	LWC	6	0.0000	0.000	0.000	SW7-3	
	LWC	7	0.0000	0.000	0.000	SW7-5	
	W4T	9	0.0000	0.000	0.000	WAG4 MS1	
	W4T	6	0.0000	0.000	0.000	WAG4 T2A	
	W4T	7	0.0030	0.003	0.273	BTT	
	LWC	9	0.0176	0.023	2.075	EAST SEEP	
	LMB	10	0.0060	0.006	0.545	MID. DRAIN.	
	WOL	10	0.0784	0.084	7.648	RS-3A	
LMB	10	0.0033	0.004	0.367	SW2-5		
Unknown	3	0.0080	0.008	0.727	SH2-6		
Unknown	4	0.0060	0.006	0.545	SH2-7		
LWC	8	0.0561	0.070	6.382	SH7-3		
LWC	10	0.0053	0.006	0.549	SH7-5		
W4T	11	0.0060	0.006	0.545	WAG4 MS1		
W4T	7	0.0350	0.035	3.182	WAG4 T2A		
W4T	7	0.0030	0.003	0.980	BTT		
CHROMIUM	W4T	6	0.0011	0.011	0.000	NAHQ CHR	
	LMB	9	0.0011	0.011	0.000	NAHQ CHR	
	WOL	8	0.0011	0.011	1.509	NAHQ CHR	
	LMB	7	0.0011	0.011	0.000	NAHQ CHR	
	W4T	5	0.0011	0.011	0.000	NAHQ CHR	
	LMB	5	0.0011	0.011	1.818	NAHQ CHR	
	LWC	6	0.0011	0.011	0.000	NAHQ CHR	
	LWC	7	0.0011	0.011	0.000	NAHQ CHR	
	W4T	9	0.0011	0.011	0.000	NAHQ CHR	
	W4T	6	0.0011	0.011	0.000	NAHQ CHR	
	W4T	7	0.0110	0.110	0.273	NAHQ CHR	
	LWC	9	0.0110	0.110	2.075	NAHQ CHR	
	LMB	10	0.0110	0.110	0.545	NAHQ CHR	
	WOL	10	0.0110	0.110	7.648	NAHQ CHR	
LMB	10	0.0110	0.110	0.367	NAHQ CHR		
Unknown	3	0.0110	0.110	0.727	NAHQ CHR		
Unknown	4	0.0110	0.110	0.545	NAHQ CHR		
LWC	8	0.0110	0.110	6.382	NAHQ CHR		
LWC	10	0.0110	0.110	0.549	NAHQ CHR		
W4T	11	0.0110	0.110	0.545	NAHQ CHR		
W4T	7	0.0110	0.110	3.182	NAHQ CHR		
W4T	7	0.0031	0.031	0.980	S_CHR_V		
COBALT	W4T	6	0.0011	0.011	0.000	NAHQ CHR	
	LMB	9	0.0011	0.011	0.000	NAHQ CHR	
	WOL	8	0.0011	0.011	1.509	NAHQ CHR	
	LMB	7	0.0011	0.011	0.000	NAHQ CHR	
	W4T	5	0.0011	0.011	0.000	NAHQ CHR	
	LMB	5	0.0011	0.011	1.818	NAHQ CHR	
	LWC	6	0.0011	0.011	0.000	NAHQ CHR	
	LWC	7	0.0011	0.011	0.000	NAHQ CHR	
	W4T	9	0.0011	0.011	0.000	NAHQ CHR	
	W4T	6	0.0011	0.011	0.000	NAHQ CHR	
	W4T	7	0.0110	0.110	0.273	NAHQ CHR	
	LWC	9	0.0110	0.110	2.075	NAHQ CHR	
	LMB	10	0.0110	0.110	0.545	NAHQ CHR	
	WOL	10	0.0110	0.110	7.648	NAHQ CHR	
LMB	10	0.0110	0.110	0.367	NAHQ CHR		
Unknown	3	0.0110	0.110	0.727	NAHQ CHR		
Unknown	4	0.0110	0.110	0.545	NAHQ CHR		
LWC	8	0.0110	0.110	6.382	NAHQ CHR		
LWC	10	0.0110	0.110	0.549	NAHQ CHR		
W4T	11	0.0110	0.110	0.545	NAHQ CHR		
W4T	7	0.0110	0.110	3.182	NAHQ CHR		
W4T	7	0.0031	0.031	0.980	S_CHR_V		

LWC	EAST SEEP	9	6	0.0087	0.011	3.668	0.0031	S_CHR_V
LWC	MV-1	2	1	0.0060	0.006	1.961	0.0031	S_CHR_V
WOL	RS-3A	10	9	0.0286	0.031	10.206	0.0031	S_CHR_V
LMB	SW2-5	10	6	0.0049	0.006	2.086	0.0031	S_CHR_V
Unknown	SW2-6	3	1	0.0050	0.005	1.634	0.0031	S_CHR_V
W4T	SW4-1	2	1	0.0080	0.008	2.614	0.0031	S_CHR_V
W4T	SW4-2	6	4	0.0198	0.027	8.961	0.0031	S_CHR_V
WOL	SW7-2	2	2	0.0940	0.347	113.252	0.0031	S_CHR_V
LWC	SW7-3	8	5	0.0133	0.017	5.546	0.0031	S_CHR_V
LWC	SW7-5	10	5	0.0095	0.014	4.471	0.0031	S_CHR_V
W4T	WAG4 MS1	11	3	0.0031	0.004	1.444	0.0031	S_CHR_V
W4T	BTT	7	2	0.0026	0.004	0.335	0.0120	NAWQ_CHR
LWC	EAST SEEP	9	6	0.0057	0.008	0.672	0.0120	NAWQ_CHR
UMB	MBTRIB-2A	1	1	0.0160	0.016	1.333	0.0120	NAWQ_CHR
LMB	MID. DRAIN.	10	2	0.0015	0.002	0.201	0.0120	NAWQ_CHR
WOL	RS-3A	10	8	0.0180	0.022	1.802	0.0120	NAWQ_CHR
LMB	SW2-5	10	4	0.0029	0.004	0.301	0.0120	NAWQ_CHR
Unknown	SW2-6	3	1	0.0020	0.002	0.167	0.0120	NAWQ_CHR
Unknown	SW2-7	4	1	0.0020	0.002	0.167	0.0120	NAWQ_CHR
W4T	SW4-2	6	2	0.0025	0.004	0.317	0.0120	NAWQ_CHR
WOL	SW7-2	2	1	0.0250	0.025	2.083	0.0120	NAWQ_CHR
LWC	SW7-3	8	6	0.0053	0.006	0.523	0.0120	NAWQ_CHR
LWC	SW7-5	10	4	0.0038	0.006	0.511	0.0120	NAWQ_CHR
HRT	SW9-1	3	1	0.0070	0.007	0.583	0.0120	NAWQ_CHR
HRT	SW9-2	3	2	0.0090	0.009	0.750	0.0120	NAWQ_CHR
W4T	WAG4 MS1	11	4	0.0032	0.004	0.365	0.0120	NAWQ_CHR
W4T	WAG4 T2A	7	2	0.0027	0.004	0.362	0.0120	NAWQ_CHR
IHP	5NNT	1	1	0.1400	0.140	15.748	0.0089	S_CHR_V
MWC	SNST	1	1	0.0040	0.004	0.450	0.0089	S_CHR_V
W4T	BTT	1	1	0.0000	0.000	0.000	0.0089	S_CHR_V
LWC	EAST SEEP	1	1	0.0110	0.011	1.237	0.0089	S_CHR_V
LMB	MBTRIB-3	1	1	0.0630	0.063	7.087	0.0089	S_CHR_V
LMB	MID. DRAIN.	1	1	0.0160	0.016	1.800	0.0089	S_CHR_V
LHC	HV-2	1	1	0.0000	0.000	0.000	0.0089	S_CHR_V
UMB	MV-3	1	1	0.0010	0.001	0.112	0.0089	S_CHR_V
WOL	RS-1	1	1	0.0250	0.025	2.812	0.0089	S_CHR_V
WOL	RS-3B	1	1	0.2600	0.260	29.246	0.0089	S_CHR_V
MWC	SW2-1	1	1	0.1200	0.120	13.498	0.0089	S_CHR_V
MWC	SW2-3	2	2	0.0375	0.074	30.851	0.0089	S_CHR_V
MWC	SW2-4	1	1	0.0410	0.041	4.612	0.0089	S_CHR_V
LMB	SW2-5	1	1	0.2400	0.240	26.997	0.0089	S_CHR_V
WOL	SW6-1	1	1	0.0020	0.002	0.225	0.0089	S_CHR_V
WOL	SW6-2	1	1	0.0100	0.010	1.125	0.0089	S_CHR_V
WOL	SW7-1	1	1	0.4100	0.410	46.119	0.0089	S_CHR_V
WOL	SW7-2	1	1	0.0670	0.067	7.537	0.0089	S_CHR_V
LWC	SW7-5	1	1	0.3000	0.300	33.746	0.0089	S_CHR_V
LWC	SW7-6	1	1	0.1100	0.110	12.373	0.0089	S_CHR_V
LWC	SW7-7	1	1	0.0430	0.043	4.837	0.0089	S_CHR_V
LWC	SW7-8	1	1	0.0000	0.000	0.000	0.0089	S_CHR_V

COPPER

Carbon disulfide

LWC	EAST SEEP	7	2	0.0025	0.004	1.153	0.0032	NAWQ_CHR
LMB	MID. DRAIN.	5	1	0.0010	0.001	0.313	0.0032	NAWQ_CHR
WOL	RS-1	1	1	0.0030	0.003	0.938	0.0032	NAWQ_CHR
WOL	RS-3A	6	4	0.0043	0.006	1.800	0.0032	NAWQ_CHR
WOL	RS-3B	1	1	0.0040	0.004	1.250	0.0032	NAWQ_CHR
MWC	SH2-1	1	1	0.0030	0.003	0.938	0.0032	NAWQ_CHR
MWC	SH2-4	2	1	0.0080	0.008	2.500	0.0032	NAWQ_CHR
LMB	SH2-5	6	3	0.0012	0.002	0.513	0.0032	NAWQ_CHR
Unknown	SH2-6	1	1	0.0020	0.002	0.625	0.0032	NAWQ_CHR
Unknown	SH2-7	2	1	0.0020	0.002	0.625	0.0032	NAWQ_CHR
LMB	SH5-4	3	1	0.0110	0.011	3.438	0.0032	NAWQ_CHR
WOL	SH7-2	2	1	0.0020	0.002	0.625	0.0032	NAWQ_CHR
LWC	SH7-3	6	1	0.0030	0.003	0.938	0.0032	NAWQ_CHR
LWC	SH7-5	8	2	0.0040	0.004	1.250	0.0032	NAWQ_CHR
LWC	SH7-8	1	1	0.0030	0.003	0.938	0.0032	NAWQ_CHR
W4T	WAG4 T2A	5	1	0.0010	0.001	0.313	0.0032	NAWQ_CHR
W4T	SH4-2	1	1	0.1130	0.113	7.842	0.0144	SCV_NEW
W4T	BTT	7	4	0.0083	0.012	0.146	0.0803	S_CHR_V
LWC	EAST SEEP	9	9	0.2593	0.358	4.452	0.0803	S_CHR_V
LMB	MBTRIB-2A	1	1	0.1420	0.142	1.768	0.0803	S_CHR_V
LMB	MBTRIB-3	2	2	0.3685	0.561	6.987	0.0803	S_CHR_V
LMB	MID. DRAIN.	10	10	1.8748	3.000	37.362	0.0803	S_CHR_V
LWC	MV-1	2	2	3.9650	4.123	51.343	0.0803	S_CHR_V
WOL	MV-3	1	1	0.0190	0.019	0.237	0.0803	S_CHR_V
WOL	RS-1	1	1	1.3500	1.350	16.812	0.0803	S_CHR_V
WOL	RS-3A	10	10	0.2344	0.311	3.871	0.0803	S_CHR_V
WOL	RS-3B	2	2	0.0560	0.138	1.720	0.0803	S_CHR_V
MWC	SH2-1	1	1	0.9920	0.992	12.354	0.0803	S_CHR_V
WMC	SH2-2	1	1	0.7960	0.796	9.913	0.0803	S_CHR_V
MWC	SH2-3	1	1	0.0930	0.093	1.158	0.0803	S_CHR_V
MWC	SH2-4	2	2	1.5775	5.887	73.308	0.0803	S_CHR_V
LMB	SH2-5	10	10	0.9450	1.423	17.718	0.0803	S_CHR_V
Unknown	SH2-6	3	3	1.0000	1.563	19.459	0.0803	S_CHR_V
Unknown	SH2-7	4	4	1.4475	1.968	24.504	0.0803	S_CHR_V
W4T	SH4-1	2	2	2.0935	12.931	161.034	0.0803	S_CHR_V
W4T	SH4-2	6	6	0.7535	1.061	13.209	0.0803	S_CHR_V
LMB	SH5-2	1	1	0.0170	0.017	0.212	0.0803	S_CHR_V
LMB	SH5-4	6	6	0.7902	1.546	19.256	0.0803	S_CHR_V
WOL	SH7-1	2	2	6.3900	6.453	80.363	0.0803	S_CHR_V
LWC	SH7-2	8	3	0.1270	0.253	3.154	0.0803	S_CHR_V
LWC	SH7-3	10	8	0.0054	0.006	0.077	0.0803	S_CHR_V
LWC	SH7-5	1	1	0.2716	0.703	8.755	0.0803	S_CHR_V
LWC	SH7-6	1	1	0.0710	0.071	0.884	0.0803	S_CHR_V
LWC	SH7-8	1	1	0.1260	0.126	1.569	0.0803	S_CHR_V
HRT	SH9-1	3	3	0.2477	0.360	4.485	0.0803	S_CHR_V
HRT	SH9-2	3	3	1.1750	2.706	33.694	0.0803	S_CHR_V
W4T	WAG4 MS1	11	10	0.2138	0.338	4.214	0.0803	S_CHR_V
W4T	WAG4 T2A	7	6	0.1250	0.226	2.818	0.0803	S_CHR_V
LWC	WCTRIB-1	2	2	0.2130	1.198	14.918	0.0803	S_CHR_V

LITHIUM

MANGANESE

Element	Sample	Count	Area	CPM	Efficiency	Concentration	Unit
MERCURY	WCTRIB-2	1	0.0240	0.024	0.299	0.0803	S_CHR_V
	WCTRIB-3	1	0.5850	0.585	7.285	0.0803	S_CHR_V
	WOCET	2	0.3545	2.214	27.570	0.0803	S_CHR_V
	RS-3A	10	0.0009	0.001	0.833	0.0013	S_CHR_V
	SW2-1	1	0.0000	0.000	0.000	0.0013	S_CHR_V
	SW2-4	2	0.0000	0.000	0.000	0.0013	S_CHR_V
	SW4-2	4	0.0000	0.000	0.000	0.0013	S_CHR_V
	SW7-3	8	0.0000	0.000	0.000	0.0013	S_CHR_V
	W4T	7	0.0612	0.114	0.479	0.2390	S_CHR_V
	EAST SEEP	9	0.0150	0.015	0.063	0.2390	S_CHR_V
MOLYBDENUM	RS-3A	10	0.0483	0.052	0.216	0.2390	S_CHR_V
	SW4-2	6	0.0080	0.008	0.033	0.2390	S_CHR_V
	SW7-3	8	0.0123	0.014	0.059	0.2390	S_CHR_V
	WAG4 MS1	11	0.0065	0.007	0.031	0.2390	S_CHR_V
	SNHT	1	0.0130	0.013	0.006	2.2400	S_CHR_V
	SNST	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	BTT	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	EAST SEEP	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	MID. DRAIN.	1	0.0010	0.001	0.000	2.2400	S_CHR_V
	MV-1	1	0.0010	0.001	0.000	2.2400	S_CHR_V
Methylene chloride	MV-2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	MV-3	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	RS-1	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	RS-3A	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW2-1	1	0.0010	0.001	0.000	2.2400	S_CHR_V
	SW2-2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW2-3	2	0.0020	0.008	0.004	2.2400	S_CHR_V
	SW2-4	1	0.0010	0.001	0.000	2.2400	S_CHR_V
	SW6-1	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW6-2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
NICKEL	SW7-2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW7-3	2	0.0005	0.004	0.002	2.2400	S_CHR_V
	SW7-4	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW7-5	1	0.0300	0.030	0.013	2.2400	S_CHR_V
	SW7-8	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	SW9-2	2	0.0010	0.001	0.000	2.2400	S_CHR_V
	WAG4 T2A	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	WAG6 MS2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	WCTRIB-1	1	0.0000	0.000	0.000	2.2400	S_CHR_V
	WCTRIB-2	1	0.0000	0.000	0.000	2.2400	S_CHR_V
WOCET	1	0.0010	0.001	0.000	2.2400	S_CHR_V	
WSTRIB-1	1	0.0000	0.000	0.000	2.2400	S_CHR_V	
W4T	7	0.0395	0.048	0.300	0.1600	MAVQ_CHR	
LWC	9	0.0269	0.032	0.199	0.1600	MAVQ_CHR	
LMB	10	0.0060	0.006	0.038	0.1600	MAVQ_CHR	
WOL	10	0.0970	0.103	0.645	0.1600	MAVQ_CHR	
LMB	10	0.0104	0.013	0.082	0.1600	MAVQ_CHR	

Tetrachloroethene	WOL	FRENCH DR S	1	1	0.0550	0.055	0.440	0.1250	S_CHR_V
	LMB	MID. DRAIN.	1	1	0.0000	0.000	0.000	0.1250	S_CHR_V
	W4T	WAG4 T2A	1	1	0.0000	0.000	0.000	0.1250	S_CHR_V
	WOL	WAG6 MS3B	1	1	0.0010	0.001	0.008	0.1250	S_CHR_V
Trichloroethene	WOL	FRENCH DR S	1	1	1.0000	1.000	2.151	0.4650	S_CHR_V
	LMB	MID. DRAIN.	1	1	0.0010	0.001	0.002	0.4650	S_CHR_V
	HRT	SH9-2	2	1	0.0000	0.000	0.000	0.4650	S_CHR_V
	W4T	WAG4 T2A	1	1	0.0020	0.002	0.004	0.4650	S_CHR_V
VANADIUM	WOL	WAG6 MS2	1	1	0.0030	0.003	0.006	0.4650	S_CHR_V
	WOL	RS-3A	10	9	0.0217	0.027	1.388	0.0191	S_CHR_V
	LMB	SH2-5	10	1	0.0030	0.003	0.157	0.0191	S_CHR_V
	LWC	SH7-3	8	2	0.0040	0.004	0.209	0.0191	S_CHR_V
Vinyl chloride	LWC	SH7-5	9	1	0.0020	0.002	0.105	0.0191	S_CHR_V
	LMB	MID. DRAIN.	1	1	0.0010	0.001	0.011	0.0878	S_CHR_V
	W4T	WAG4 MS1	1	1	0.0460	0.046	0.524	0.0878	S_CHR_V
	W4T	WAG4 T2A	1	1	0.0000	0.000	0.000	0.0878	S_CHR_V
ZINC	W4T	BTT	7	2	0.0118	0.016	0.149	0.1100	NAVQ_CHR
	LVC	EAST SEEP	8	5	0.0110	0.015	0.138	0.1100	NAVQ_CHR
	LMB	MID. DRAIN.	10	3	0.0053	0.006	0.052	0.1100	NAVQ_CHR
	WOL	RS-3A	9	7	0.0386	0.049	0.449	0.1100	NAVQ_CHR
	WOL	RS-3B	2	1	0.0090	0.009	0.082	0.1100	NAVQ_CHR
	MVC	SH2-4	2	1	0.0040	0.004	0.036	0.1100	NAVQ_CHR
	LMB	SH2-5	10	6	0.0168	0.024	0.220	0.1100	NAVQ_CHR
	Unknown	SH2-6	3	2	0.0110	0.017	0.153	0.1100	NAVQ_CHR
	Unknown	SH2-7	4	2	0.0305	0.067	0.605	0.1100	NAVQ_CHR
	W4T	SH4-1	2	1	0.0300	0.030	0.273	0.1100	NAVQ_CHR
	W4T	SH4-2	6	4	0.0344	0.056	0.512	0.1100	NAVQ_CHR
	LMB	SH5-4	6	3	0.0066	0.009	0.081	0.1100	NAVQ_CHR
	LWC	SH7-3	7	5	0.0281	0.031	0.283	0.1100	NAVQ_CHR
	LWC	SH7-5	8	5	0.0116	0.018	0.164	0.1100	NAVQ_CHR
	HRT	SH9-1	3	2	0.0107	0.033	0.301	0.1100	NAVQ_CHR
	HRT	SH9-2	3	3	0.1143	0.368	3.346	0.1100	NAVQ_CHR
	W4T	WAG4 MS1	11	3	0.0059	0.008	0.069	0.1100	NAVQ_CHR
W4T	WAG4 T2A	7	3	0.0120	0.019	0.177	0.1100	NAVQ_CHR	

Table B.3. Screening of aqueous concentrations of radionuclides detected in streams against screening benchmarks for aquatic life. Concentrations are pCi/L.

Radionuclide	Reach	Samples	Detects	P.L. Mean	95% UCB	Quotient	Benchmark Value	Benchmark
Cesium-137	1WC	5	5	112.62	179.30	0.004733	37882.0000	WAT_RAD
	IHP	9	9	109.87	134.53	0.003551	37882.0000	WAT_RAD
	LMB	12	1	15.90	15.90	0.000420	37882.0000	WAT_RAD
	LWC	3	2	40.10	85.62	0.002260	37882.0000	WAT_RAD
	MWC	12	12	66.95	85.18	0.002249	37882.0000	WAT_RAD
	W4T	16	3	40.31	50.19	0.001325	37882.0000	WAT_RAD
	WOL	12	12	67.33	79.17	0.002090	37882.0000	WAT_RAD
Cobalt-60	WS	7	1	31.30	31.30	0.000061	510314.000	WAT_RAD
Curium-244	1WC	2	2	0.00	0.00	0.000000	5537.0000	WAT_RAD
	IHP	1	1	0.00	0.00	0.000000	5537.0000	WAT_RAD
Plutonium-238	1WC	2	2	0.00	0.00	0.000000	5845.0000	WAT_RAD
	IHP	1	1	0.00	0.00	0.000000	5845.0000	WAT_RAD
	LMB	1	1	0.16	0.16	0.000027	5845.0000	WAT_RAD
Strontium-90	1C	2	2	477.00	1973.36	0.005694	346570.000	WAT_RAD
	1WC	4	2	87.50	212.88	0.000614	346570.000	WAT_RAD
	HRT	19	19	1057.61	1444.71	0.004169	346570.000	WAT_RAD
	IHP	3	1	196.00	196.00	0.000566	346570.000	WAT_RAD
	LMB	21	21	641.60	804.60	0.002322	346570.000	WAT_RAD
	LWC	1	1	326.00	326.00	0.000941	346570.000	WAT_RAD
	MWC	3	2	130.80	287.08	0.000828	346570.000	WAT_RAD
	NWT	2	1	96.00	96.00	0.000277	346570.000	WAT_RAD
	UMB	2	1	20.00	20.00	0.000058	346570.000	WAT_RAD
	W4T	20	20	9778.88	10681.82	0.030822	346570.000	WAT_RAD
	WOL	3	3	224.00	330.21	0.000953	346570.000	WAT_RAD
	WS	3	2	179.67	225.75	0.000651	346570.000	WAT_RAD
	1C	7	7	269.80	411.81	0.001188	346570.000	WAT_RAD
	1WC	10	8	77.56	118.79	0.000343	346570.000	WAT_RAD
	HRT	7	7	947.00	1242.22	0.003584	346570.000	WAT_RAD
	IHP	13	13	80.95	94.42	0.000272	346570.000	WAT_RAD
	LMB	32	32	532.49	643.31	0.001856	346570.000	WAT_RAD
	LWC	10	9	178.04	212.35	0.000613	346570.000	WAT_RAD
	MWC	24	24	105.43	119.69	0.000345	346570.000	WAT_RAD
	NWT	6	6	36.55	58.92	0.000170	346570.000	WAT_RAD
UMB	4	2	0.00	0.00	0.000000	346570.000	WAT_RAD	
W4T	1	1	7168.00	7168.00	0.020683	346570.000	WAT_RAD	
WOL	12	12	194.00	213.04	0.000615	346570.000	WAT_RAD	
WS	7	7	135.79	183.74	0.000530	346570.000	WAT_RAD	
Thorium-228	LMB	1	1	1.25	1.25	0.000916	1365.0000	WAT_RAD
Thorium-230	LMB	1	1	0.17	0.17	0.000080	2064.0000	WAT_RAD
Thorium-232	LMB	1	1	0.01	0.01	0.000004	2389.0000	WAT_RAD
Tritium	1C	9	9	1.15	2.03	0.000000	3445329521	WAT_RAD
	1WC	3	3	11.54	27.71	0.000000	3445329521	WAT_RAD
	BWC	2	2	1.57	6.75	0.000000	3445329521	WAT_RAD
	HRT	52	51	1784.99	2190.70	0.000001	3445329521	WAT_RAD
	IHP	16	16	8.50	12.23	0.000000	3445329521	WAT_RAD
	LMB	147	147	751.34	816.59	0.000000	3445329521	WAT_RAD
	LWC	24	24	132.94	156.69	0.000000	3445329521	WAT_RAD
	MWC	34	34	28.33	33.13	0.000000	3445329521	WAT_RAD
	NWT	3	3	0.49	1.21	0.000000	3445329521	WAT_RAD
	RAC	2	2	1.34	5.25	0.000000	3445329521	WAT_RAD
	UMB	15	15	17.62	26.87	0.000000	3445329521	WAT_RAD
	W4T	110	110	11631.42	13062.04	0.000004	3445329521	WAT_RAD
	WOL	15	15	156.62	191.31	0.000000	3445329521	WAT_RAD
	WS	10	10	49.55	115.28	0.000000	3445329521	WAT_RAD
	1C	1	1	1.68	1.68	0.000000	3445329521	WAT_RAD
	1WC	1	1	4.94	4.94	0.000000	3445329521	WAT_RAD

B-33

	HRT	1	1	1.88	1.88	0.000000	3445329521	WAT_RAD
	IHP	6	6	10.01	17.44	0.000000	3445329521	WAT_RAD
	LMB	8	8	1111.21	1514.15	0.000000	3445329521	WAT_RAD
	MWC	10	10	28.34	38.52	0.000000	3445329521	WAT_RAD
	RAC	1	1	0.81	0.81	0.000000	3445329521	WAT_RAD
	WOL	9	9	151.34	199.66	0.000000	3445329521	WAT_RAD
Uranium-235	LMB	1	1	0.03	0.03	0.000001	21833.0000	WAT_RAD
Uranium-238	LMB	1	1	0.53	0.53	0.000023	22727.0000	WAT_RAD

Table B.4. Screening of aqueous concentrations of radionuclides detected in seeps and ephemeral tributaries against screening benchmarks for aquatic life. Concentrations are pCi/L.

Radionuclide	Reach	Seep ID	Samples	Detects	P.L. Mean	95% UCB	Quotient Value	Benchmark	Benchmark	
Cesium-137	LWC	SW7-4	1	1	28.50	28.50	0.00075	37882.0000	WAT_RAD	
	LWC	SW7-6	2	2	304.30	529.07	0.01397	37882.0000	WAT_RAD	
	LWC	WTRIB-1	3	1	29.20	29.20	0.00077	37882.0000	WAT_RAD	
	W4T	BTT	5	5	300.29	690.41	0.01823	37882.0000	WAT_RAD	
	W4T	SW4-2	5	5	247.79	450.83	0.01190	37882.0000	WAT_RAD	
	W4T	WAG4 T2A	4	4	254.11	401.08	0.01059	37882.0000	WAT_RAD	
	WS	RS-1	1	1	35.90	35.90	0.00095	37882.0000	WAT_RAD	
	WS	RS-3B	2	1	13.30	13.30	0.00035	37882.0000	WAT_RAD	
	WS	SW7-2	2	1	43.20	43.20	0.00114	37882.0000	WAT_RAD	
	Cobalt-60	LWC	EAST SEEP	7	6	166.51	222.17	0.00044	510314.0000	WAT_RAD
		LWC	SW7-3	8	8	653.45	1057.22	0.00207	510314.0000	WAT_RAD
		LWC	SW7-4	1	1	179.30	179.30	0.00035	510314.0000	WAT_RAD
LWC		SW7-5	6	6	2550.38	5002.84	0.00980	510314.0000	WAT_RAD	
LWC		WTRIB-1	3	2	65.27	89.77	0.00018	510314.0000	WAT_RAD	
WS		RS-3A	8	8	835.72	972.64	0.00191	510314.0000	WAT_RAD	
WS		RS-3B	2	2	97.05	364.44	0.00071	510314.0000	WAT_RAD	
WS		SW7-2	2	2	1172.20	2869.34	0.00562	510314.0000	WAT_RAD	
WS		WS-2	2	1	111.80	111.80	0.00022	510314.0000	WAT_RAD	
Plutonium-238		LMB	SW5-4	1	1	0.45	0.45	0.00008	5845.0000	WAT_RAD
		LWC	SW7-3	1	1	0.02	0.02	0.00000	5845.0000	WAT_RAD
		W4T	BTT	1	1	0.03	0.03	0.00000	5845.0000	WAT_RAD
	W4T	SW4-1	1	1	0.16	0.16	0.00003	5845.0000	WAT_RAD	
	W4T	SW4-2	1	1	0.00	-0.07	-0.00001	5845.0000	WAT_RAD	
	W4T	WAG4 MS1	1	1	0.04	0.04	0.00001	5845.0000	WAT_RAD	
	WS	RS-1	1	1	0.11	0.11	0.00002	5845.0000	WAT_RAD	
	WS	RS-3A	1	1	0.16	0.16	0.00003	5845.0000	WAT_RAD	
	WS	RS-3B	1	1	0.23	0.23	0.00004	5845.0000	WAT_RAD	
	Strontium-90	LMB	MID. DRAIN.	2	2	2333.25	2483.20	0.00717	346570.0000	WAT_RAD
		LWC	SW2-5	3	3	163422.33	176320.66	0.50876	346570.0000	WAT_RAD
		Unknown	EAST SEEP	3	1	68.00	68.00	0.00020	346570.0000	WAT_RAD
Unknown		SW2-6	1	1	564388.00	564388.00	1.62850	346570.0000	WAT_RAD	
Unknown		SW2-7	2	2	186311.50	672088.38	1.93926	346570.0000	WAT_RAD	
W4T		BTT	1	1	19733.00	19733.00	0.05694	346570.0000	WAT_RAD	
W4T		SW4-2	1	1	13100.00	13100.00	0.03780	346570.0000	WAT_RAD	
W4T		WAG4 MS1	2	2	11123.00	21250.26	0.06132	346570.0000	WAT_RAD	
W4T		WAG4 T2A	1	1	9138.00	9138.00	0.02637	346570.0000	WAT_RAD	
WOL		WAG6 MS3	1	1	248.00	248.00	0.00072	346570.0000	WAT_RAD	
WS		WS-1	1	1	196.00	196.00	0.00057	346570.0000	WAT_RAD	
LMB		MID. DRAIN.	3	3	1516.67	1835.79	0.00530	346570.0000	WAT_RAD	
LMB	SW5-4	2	2	355176.00	854612.69	2.46592	346570.0000	WAT_RAD		

UMC	SW2-2	1	562.00	562.00	0.00162	346570.000	WAT_RAD
W4T	WAG4 MS1	3	10214.67	12498.47	0.03606	346570.000	WAT_RAD
W4T	WAG4 T2A	3	7481.83	8923.30	0.02575	346570.000	WAT_RAD
WS	RS-3A	4	5.54	8.05	0.00002	346570.000	WAT_RAD
WS	RS-3B	2	28.70	115.83	0.00033	346570.000	WAT_RAD
WOL	WAG6 MS3	1	280.00	280.00	0.00081	346570.000	WAT_RAD
WS	WS-1	3	138.97	210.84	0.00061	346570.000	WAT_RAD
WS	WS-2	2	159.50	206.85	0.00060	346570.000	WAT_RAD
WS	WS-3	2	196.50	395.38	0.00114	346570.000	WAT_RAD
WS	WSTRIB-1	3	156.20	257.66	0.00074	346570.000	WAT_RAD
Thorium-228							
LMB	SW5-4	1	0.68	0.68	0.00050	1365.0000	WAT_RAD
LWC	SW7-3	1	1.04	1.04	0.00076	1365.0000	WAT_RAD
W4T	BTT	1	2.35	2.35	0.00172	1365.0000	WAT_RAD
W4T	SW4-1	1	0.95	0.95	0.00070	1365.0000	WAT_RAD
W4T	SW4-2	1	1.30	1.30	0.00095	1365.0000	WAT_RAD
W4T	WAG4 MS1	1	3.50	3.50	0.00256	1365.0000	WAT_RAD
WS	RS-1	1	0.00	0.00	0.00000	1365.0000	WAT_RAD
WS	RS-3A	1	0.00	0.00	0.00000	1365.0000	WAT_RAD
WS	RS-3B	1	0.00	0.00	0.00000	1365.0000	WAT_RAD
Thorium-230							
LMB	SW5-4	1	0.08	0.08	0.00004	2064.0000	WAT_RAD
LWC	SW7-3	1	0.12	0.12	0.00006	2064.0000	WAT_RAD
W4T	BTT	1	0.04	0.04	0.00002	2064.0000	WAT_RAD
W4T	SW4-1	1	0.12	0.12	0.00006	2064.0000	WAT_RAD
W4T	SW4-2	1	0.14	0.14	0.00007	2064.0000	WAT_RAD
W4T	WAG4 MS1	1	0.08	0.08	0.00004	2064.0000	WAT_RAD
WS	RS-1	1	0.12	0.12	0.00006	2064.0000	WAT_RAD
WS	RS-3A	1	0.21	0.21	0.00010	2064.0000	WAT_RAD
WS	RS-3B	1	0.09	0.09	0.00004	2064.0000	WAT_RAD
Thorium-232							
LMB	SW5-4	1	0.04	0.04	0.00002	2389.0000	WAT_RAD
LWC	SW7-3	1	0.02	0.02	0.00001	2389.0000	WAT_RAD
W4T	BTT	1	0.00	-0.01	-0.00000	2389.0000	WAT_RAD
W4T	SW4-1	1	0.01	0.01	0.00000	2389.0000	WAT_RAD
W4T	SW4-2	1	0.03	0.03	0.00001	2389.0000	WAT_RAD
W4T	WAG4 MS1	1	0.04	0.04	0.00002	2389.0000	WAT_RAD
WS	RS-1	1	0.00	0.00	0.00000	2389.0000	WAT_RAD
WS	RS-3A	1	0.20	0.20	0.00008	2389.0000	WAT_RAD
WS	RS-3B	1	0.04	0.04	0.00002	2389.0000	WAT_RAD
Tritium							
EWC	WOCET	2	0.81	2.96	0.00000	3445329521	WAT_RAD
IHP	WCTRIB-3	2	103.19	273.12	0.00000	3445329521	WAT_RAD
LMB	MBTRIB-3	3	2.51	3.35	0.00000	3445329521	WAT_RAD
LMB	MID. DRAIN.	10	7387.80	8140.39	0.00000	3445329521	WAT_RAD
LMB	SW2-5	10	9420.15	10890.74	0.00000	3445329521	WAT_RAD
LMB	SW5-2	2	20.20	42.42	0.00000	3445329521	WAT_RAD
LMB	SW5-4	7	4792.71	5825.63	0.00000	3445329521	WAT_RAD
LWC	EAST SEEP	10	24.81	30.86	0.00000	3445329521	WAT_RAD
LHC	MV-1	1	1.80	1.80	0.00000	3445329521	WAT_RAD
LWC	SW7-3	4	21.46	29.73	0.00000	3445329521	WAT_RAD

LWC	SW7-5	3	3	55.73	159.07	0.00000	3445329521	WAT_RAD
LWC	SW7-6	2	2	3.58	4.05	0.00000	3445329521	WAT_RAD
LWC	SW7-8	1	1	8.26	8.26	0.00000	3445329521	WAT_RAD
LWC	WCTRIB-1	3	3	9.53	20.67	0.00000	3445329521	WAT_RAD
MHC	SW2-1	1	1	11.38	11.38	0.00000	3445329521	WAT_RAD
MHC	SW2-3	1	1	6.08	6.08	0.00000	3445329521	WAT_RAD
MHC	SW2-4	2	2	1028.85	1667.49	0.00000	3445329521	WAT_RAD
MHC	WCTRIB-2	1	1	2.60	2.60	0.00000	3445329521	WAT_RAD
NRT	SW9-1	1	1	2.72	2.72	0.00000	3445329521	WAT_RAD
NRT	SW9-2	1	1	0.90	0.90	0.00000	3445329521	WAT_RAD
UMB	MBTRIB-1	2	2	1.25	4.41	0.00000	3445329521	WAT_RAD
UMB	MBTRIB-2A	3	3	56.20	171.54	0.00000	3445329521	WAT_RAD
UMB	MBTRIB-2B	3	3	0.59	1.63	0.00000	3445329521	WAT_RAD
UMB	MV-3	1	1	0.00	-0.13	-0.00000	3445329521	WAT_RAD
UMC	SW2-2	1	1	3.91	3.91	0.00000	3445329521	WAT_RAD
Unknown	SPD	2	2	10454.50	15975.88	0.00000	3445329521	WAT_RAD
Unknown	SW2-6	4	4	2441.75	2999.32	0.00000	3445329521	WAT_RAD
Unknown	SW2-7	7	7	2491.00	2734.15	0.00000	3445329521	WAT_RAD
W4T	BTT	7	7	29.98	38.14	0.00000	3445329521	WAT_RAD
W4T	SW4-1	2	2	204.68	1370.98	0.00000	3445329521	WAT_RAD
W4T	SW4-2	7	7	17.48	21.73	0.00000	3445329521	WAT_RAD
W4T	WAG4 MS1	10	10	7446.35	11140.71	0.00000	3445329521	WAT_RAD
W4T	WAG4 T2A	7	7	6054.14	8853.05	0.00000	3445329521	WAT_RAD
WS	RS-1	1	1	15.27	15.27	0.00000	3445329521	WAT_RAD
WS	RS-3A	4	4	37.98	49.66	0.00000	3445329521	WAT_RAD
WS	RS-3B	2	2	12.57	41.64	0.00000	3445329521	WAT_RAD
LWC	SW7-1	1	1	26.30	26.30	0.00000	3445329521	WAT_RAD
LWC	SW7-2	2	2	8.38	9.96	0.00000	3445329521	WAT_RAD
WOL	WAG6 MS2	3	3	2538.50	3616.08	0.00000	3445329521	WAT_RAD
WOL	WAG6 MS3	1	1	2314.00	2314.00	0.00000	3445329521	WAT_RAD
WOL	WAG6 MS3B	3	3	2618.00	3287.14	0.00000	3445329521	WAT_RAD
WS	WS-1	3	3	15.22	29.34	0.00000	3445329521	WAT_RAD
WS	WS-2	2	2	22.98	97.04	0.00000	3445329521	WAT_RAD
WS	WS-3	2	2	10.65	12.63	0.00000	3445329521	WAT_RAD
WS	WSTRIB-1	3	3	20.57	50.85	0.00000	3445329521	WAT_RAD
W4T	SW4-2	1	1	17.77	17.77	0.00000	3445329521	WAT_RAD
W4T	WAG4 MS1	3	3	9344.67	25014.93	0.00001	3445329521	WAT_RAD
W4T	WAG4 T2A	1	1	12000.00	12000.00	0.00000	3445329521	WAT_RAD
WS	WS-3	1	1	6.57	6.57	0.00000	3445329521	WAT_RAD
LMB	SW5-4	1	1	0.01	0.01	0.00000	21833.0000	WAT_RAD
LWC	SW7-3	1	1	9.97	9.97	0.00046	21833.0000	WAT_RAD
W4T	BTT	1	1	5.32	5.32	0.00024	21833.0000	WAT_RAD
W4T	SW4-1	1	1	0.01	0.01	0.00000	21833.0000	WAT_RAD
W4T	SW4-2	1	1	0.06	0.06	0.00000	21833.0000	WAT_RAD
W4T	WAG4 MS1	1	1	0.68	0.68	0.00003	21833.0000	WAT_RAD
WS	RS-1	1	1	0.01	0.01	0.00000	21833.0000	WAT_RAD
WS	RS-3A	1	1	6.22	6.22	0.00028	21833.0000	WAT_RAD
WS	RS-3B	1	1	0.52	0.52	0.00002	21833.0000	WAT_RAD
LMB	SW5-4	1	1	0.02	0.02	0.00000	22727.0000	WAT_RAD

Uranium-235

Uranium-238

Table B.5. Screening of aqueous concentrations of chemicals (including radionuclides) detected in sediments against screening benchmarks for benthic invertebrates (Hull and Suter 1996). All concentrations are mg/L. See Table 4.1 for definitions of benchmarks.

Chemical	Benchmark	Reach	Samples	Detects	P.L. Mean	Maximum	Quotient	Benchmark Value
2-Methylnaphthalene	ER_L	IHP	5	2	0.03	0.030830	0.440423	0.070000
		LWC	7	1	0.08	0.083000	1.185714	0.070000
2-Methylnaphthalene	ER_M	IHP	5	2	0.03	0.030830	0.046014	0.670000
		LWC	7	1	0.08	0.083000	0.123881	0.670000
2-Methylnaphthalene	REG_IV	IHP	5	2	0.03	0.030830	0.093423	0.330000
		LWC	7	1	0.08	0.083000	0.251515	0.330000
2-Methylnaphthalene	SED_PEL	IHP	5	2	0.03	0.030830	0.153381	0.201000
		LWC	7	1	0.08	0.083000	0.412935	0.201000
2-Methylnaphthalene	SED_TEL	IHP	5	2	0.03	0.030830	1.526219	0.020200
		LWC	7	1	0.08	0.083000	4.108911	0.020200
4-Methylphenol	AET	IHP	5	1	0.22	0.220000	0.328358	0.670000
AM-241	SED_RAD	IHP	5	5	1.99	3.115066	0.00764	4076.000000
		LMB	5	1	0.13	0.130000	0.000032	4076.000000
		LWC	4	4	2.65	7.968475	0.001955	4076.000000
		MWC	2	2	2.95	4.528438	0.001111	4076.000000
		IHP	5	2	0.08	0.187724	0.144403	1.300000
Acenaphthene	EPASQC_A	LWC	7	2	0.06	0.125011	0.096163	1.300000
		MWC	4	1	0.54	0.540000	0.415385	1.300000
		IHP	5	2	0.08	0.187724	11.732762	0.016000
Acenaphthene	ER_L	LWC	7	2	0.06	0.125011	7.813207	0.016000
		MWC	4	1	0.54	0.540000	33.750000	0.016000
		IHP	5	2	0.08	0.187724	0.375448	0.500000
Acenaphthene	ER_M	LWC	7	2	0.06	0.125011	0.250023	0.500000
		MWC	4	1	0.54	0.540000	1.080000	0.500000
		IHP	5	2	0.08	0.187724	0.568861	0.330000
Acenaphthene	REG_IV	LWC	7	2	0.06	0.125011	0.378822	0.330000
		MWC	4	1	0.54	0.540000	1.636364	0.330000
		IHP	5	2	0.08	0.187724	2.111633	0.088900
Acenaphthene	SED_PEL	LWC	7	2	0.06	0.125011	1.406201	0.088900
		MWC	4	1	0.54	0.540000	6.074241	0.088900
		IHP	5	2	0.08	0.187724	27.976779	0.006710
Acenaphthene	SED_TEL	LWC	7	2	0.06	0.125011	18.630598	0.006710
		MWC	4	1	0.54	0.540000	80.476902	0.006710
		IHP	5	2	0.08	0.187724	27.976779	0.006710

Anthracene	EQSQB_A	IHP	5	2	0.13	0.325262	1084.205837	0.000300
		LWC	7	2	0.06	0.095920	319.734751	0.000300
		MWC	4	2	0.28	0.706312	2354.373817	0.000300
Anthracene	ER_L	IHP	5	2	0.13	0.325262	3.826609	0.085000
		LWC	7	2	0.06	0.095920	1.128476	0.085000
		MWC	4	2	0.28	0.706312	8.309555	0.085000
Anthracene	ER_M	IHP	5	2	0.13	0.325262	0.295693	1.100000
		LWC	7	2	0.06	0.095920	0.087200	1.100000
		MWC	4	2	0.28	0.706312	0.642102	1.100000
Anthracene	REG_IV	IHP	5	2	0.13	0.325262	0.985642	0.330000
		LWC	7	2	0.06	0.095920	0.290668	0.330000
		MWC	4	2	0.28	0.706312	2.140340	0.330000
Anthracene	SED_PEL	IHP	5	2	0.13	0.325262	1.327599	0.245000
		LWC	7	2	0.06	0.095920	0.391512	0.245000
		MWC	4	2	0.28	0.706312	2.882907	0.245000
Anthracene	SED_TEL	IHP	5	2	0.13	0.325262	6.935218	0.046900
		LWC	7	2	0.06	0.095920	2.045212	0.046900
		MWC	4	2	0.28	0.706312	15.059960	0.046900
Antimony	ER_L	IHP	3	2	0.51	0.918798	0.459399	2.000000
		LMB	5	5	0.92	1.052792	0.526396	2.000000
		LWC	3	3	0.99	1.442676	0.721338	2.000000
Antimony	ER_M	MWC	2	2	0.58	0.732844	0.366422	2.000000
		IHP	3	2	0.51	0.918798	0.036752	25.000000
		LMB	5	5	0.92	1.052792	0.042112	25.000000
Antimony	REG_IV	LWC	3	3	0.99	1.442676	0.057707	25.000000
		MWC	2	2	0.58	0.732844	0.029314	25.000000
		IHP	3	2	0.51	0.918798	0.076566	12.000000
Aroclor-1254	EQSQB_A	LMB	5	5	0.92	1.052792	0.087733	12.000000
		LWC	3	3	0.99	1.442676	0.120223	12.000000
		MWC	2	2	0.58	0.732844	0.061070	12.000000
Aroclor-1254	ER_L	IHP	5	3	0.25	0.449889	2.646404	0.170000
		LWC	7	1	0.05	0.047000	0.276471	0.170000
		IHP	5	3	0.25	0.449889	19.560381	0.023000
Aroclor-1254	ER_M	LWC	7	1	0.05	0.047000	2.043478	0.023000
		IHP	5	3	0.25	0.449889	2.499382	0.180000
		LWC	7	1	0.05	0.047000	0.261111	0.180000
Aroclor-1260	EQSQB_A	IHP	5	5	0.22	0.381206	0.000347	1099.000000
		LMB	4	1	0.04	0.038000	0.000035	1099.000000
		LWC	7	3	0.07	0.152737	0.000139	1099.000000

Aroclor-1260	ER_L	MWC	4	4	0.23	0.525667	0.000478	1099.000000
		IHP	5	5	0.22	0.381206	16.574153	0.023000
		LMB	4	1	0.04	0.038000	1.652174	0.023000
		LWC	7	3	0.07	0.152737	6.640733	0.023000
		MWC	4	4	0.23	0.525667	22.855074	0.023000
Aroclor-1260	ER_M	IHP	5	5	0.22	0.381206	2.117808	0.180000
		LMB	4	1	0.04	0.038000	0.211111	0.180000
		LWC	7	3	0.07	0.152737	0.848538	0.180000
		MWC	4	4	0.23	0.525667	2.920371	0.180000
Arsenic	ER_L	IHP	3	3	4.40	8.860352	1.080531	8.200000
		LMB	5	5	5.04	7.292707	0.889355	8.200000
		LWC	3	3	4.40	8.821955	1.075848	8.200000
		MWC	2	2	5.80	15.270627	1.862272	8.200000
Arsenic	ER_M	IHP	3	3	4.40	8.860352	0.126576	70.000000
		LMB	5	5	5.04	7.292707	0.104182	70.000000
		LWC	3	3	4.40	8.821955	0.126028	70.000000
		MWC	2	2	5.80	15.270627	0.218152	70.000000
Arsenic	LEL_MOE	IHP	3	3	4.40	8.860352	1.476725	6.000000
		LMB	5	5	5.04	7.292707	1.215451	6.000000
		LWC	3	3	4.40	8.821955	1.470326	6.000000
		MWC	2	2	5.80	15.270627	2.545105	6.000000
Arsenic	REG_IV	IHP	3	3	4.40	8.860352	1.107544	8.000000
		LMB	5	5	5.04	7.292707	0.911588	8.000000
		LWC	3	3	4.40	8.821955	1.102744	8.000000
		MWC	2	2	5.80	15.270627	1.908828	8.000000
Arsenic	SED_PEL	IHP	3	3	4.40	8.860352	0.212989	41.600001
		LMB	5	5	5.04	7.292707	0.175305	41.600001
		LWC	3	3	4.40	8.821955	0.212066	41.600001
		MWC	2	2	5.80	15.270627	0.367082	41.600001
Arsenic	SED_TEL	IHP	3	3	4.40	8.860352	1.223806	7.240000
		LMB	5	5	5.04	7.292707	1.007280	7.240000
		LWC	3	3	4.40	8.821955	1.218502	7.240000
		MWC	2	2	5.80	15.270627	2.109203	7.240000
Barium	REG_V_L	IHP	3	3	102.77	196.451396	9.822570	20.000000
		LMB	5	5	358.20	681.723747	34.086187	20.000000
		LWC	3	3	197.33	297.160025	14.858001	20.000000
		MWC	2	2	118.00	130.627503	6.531375	20.000000
Barium	REG_V_M	IHP	3	3	102.77	196.451396	3.274190	60.000000
		LMB	5	5	358.20	681.723747	11.362062	60.000000
		LWC	3	3	197.33	297.160025	4.952667	60.000000
		MWC	2	2	118.00	130.627503	2.177125	60.000000

Benzo(a)anthracene	EQPSQB_A	IHP	5	4	0.23	0.546377	5.059042	0.108000
		LWC	7	1	0.03	0.032000	0.296296	0.108000
		MWC	4	2	0.60	0.995223	9.215028	0.108000
Benzo(a)anthracene	ER_L	IHP	5	4	0.23	0.546377	2.101448	0.260000
		LWC	7	1	0.03	0.032000	0.123077	0.260000
		MWC	4	2	0.60	0.995223	3.827781	0.260000
Benzo(a)anthracene	ER_M	IHP	5	4	0.23	0.546377	0.341485	1.600000
		LWC	7	1	0.03	0.032000	0.020000	1.600000
		MWC	4	2	0.60	0.995223	0.622014	1.600000
Benzo(a)anthracene	REG_IV	IHP	5	4	0.23	0.546377	1.655686	0.330000
		LWC	7	1	0.03	0.032000	0.096970	0.330000
		MWC	4	2	0.60	0.995223	3.015827	0.330000
Benzo(a)anthracene	SED_PEL	IHP	5	4	0.23	0.546377	0.788422	0.693000
		LWC	7	1	0.03	0.032000	0.046176	0.693000
		MWC	4	2	0.60	0.995223	1.436108	0.693000
Benzo(a)anthracene	SED_TEL	IHP	5	4	0.23	0.546377	7.304499	0.074800
		LWC	7	1	0.03	0.032000	0.427807	0.074800
		MWC	4	2	0.60	0.995223	13.305121	0.074800
Benzo(a)pyrene	EQPSQB_A	IHP	5	4	0.20	0.395901	2.827864	0.140000
		LWC	7	3	0.08	0.126524	0.903744	0.140000
		MWC	4	3	0.47	0.954074	6.814816	0.140000
Benzo(a)pyrene	ER_L	IHP	5	4	0.20	0.395901	0.920700	0.430000
		LWC	7	3	0.08	0.126524	0.294242	0.430000
		MWC	4	3	0.47	0.954074	2.218777	0.430000
Benzo(a)pyrene	ER_M	IHP	5	4	0.20	0.395901	0.247438	1.600000
		LWC	7	3	0.08	0.126524	0.079078	1.600000
		MWC	4	3	0.47	0.954074	0.596296	1.600000
Benzo(a)pyrene	REG_IV	IHP	5	4	0.20	0.395901	1.199700	0.330000
		LWC	7	3	0.08	0.126524	0.383406	0.330000
		MWC	4	3	0.47	0.954074	2.891134	0.330000
Benzo(a)pyrene	SED_PEL	IHP	5	4	0.20	0.395901	0.518874	0.763000
		LWC	7	3	0.08	0.126524	0.165825	0.763000
		MWC	4	3	0.47	0.954074	1.250425	0.763000
Benzo(a)pyrene	SED_TEL	IHP	5	4	0.20	0.395901	4.458344	0.088800
		LWC	7	3	0.08	0.126524	1.424821	0.088800
		MWC	4	3	0.47	0.954074	10.744080	0.088800
CM-244	SED_RAD	IHP	2	2	4.70	13.539252	0.001223	11069.000000
		LMB	5	3	0.94	1.691524	0.000153	11069.000000
		LWC	2	2	0.04	0.143677	0.000013	11069.000000

Chrysene	ER_L	IHP	5	5	0.25	0.526580	1.385738	0.380000
		LWC	7	1	0.04	0.035000	0.092105	0.380000
		MWC	4	2	0.34	0.771989	2.031550	0.380000
Chrysene	ER_M	IHP	5	5	0.25	0.526580	0.188064	2.800000
		LWC	7	1	0.04	0.035000	0.012500	2.800000
		MWC	4	2	0.34	0.771989	0.275710	2.800000
Chrysene	REG_IV	IHP	5	5	0.25	0.526580	1.595698	0.330000
		LWC	7	1	0.04	0.035000	0.106061	0.330000
		MWC	4	2	0.34	0.771989	2.339360	0.330000
Chrysene	SED_PEL	IHP	5	5	0.25	0.526580	0.622436	0.846000
		LWC	7	1	0.04	0.035000	0.041371	0.846000
		MWC	4	2	0.34	0.771989	0.912516	0.846000
Chrysene	SED_TEL	IHP	5	5	0.25	0.526580	4.875745	0.108000
		LWC	7	1	0.04	0.035000	0.324074	0.108000
		MWC	4	2	0.34	0.771989	7.148045	0.108000
Copper	ER_L	IHP	3	3	35.43	79.478143	2.337592	34.000000
		LMB	5	5	23.38	30.480170	0.896476	34.000000
		LWC	3	3	29.73	66.341745	1.951228	34.000000
Copper	ER_M	IHP	3	3	35.43	79.478143	0.294363	270.000000
		LMB	5	5	23.38	30.480170	0.112890	270.000000
		LWC	3	3	29.73	66.341745	0.245710	270.000000
Copper	REG_IV	IHP	3	3	35.43	79.478143	1.076037	270.000000
		LMB	5	5	23.38	30.480170	0.088578	28.000000
		LWC	3	3	29.73	66.341745	2.369348	28.000000
Copper	SED_PEL	IHP	3	3	35.43	79.478143	10.376073	28.000000
		LMB	5	5	23.38	30.480170	0.735909	108.000000
		LWC	3	3	29.73	66.341745	0.282224	108.000000
Copper	SED_TEL	IHP	3	3	35.43	79.478143	0.614275	108.000000
		LMB	5	5	23.38	30.480170	2.690093	108.000000
		LWC	3	3	29.73	66.341745	4.250168	18.700000
Di-n-butylphthalate	EQPSQB_A	IHP	5	4	0.28	0.449471	0.010676	42.100000
		LWC	7	2	0.36	0.578466	0.013740	42.100000
		MWC	4	2	0.22	0.564621	0.013411	42.100000
Di-n-octylphthalate	EQPSQB_A	LMB	5	1	0.06	0.055000	0.000000	7800000.000000

Dibenz(a,h)anthracene	ER_L	IHP MWC	5 4	1 1	0.24 0.10	0.240000 0.100500	3.809524 1.595238	0.063000 0.063000
Dibenz(a,h)anthracene	ER_M	IHP MWC	5 4	1 1	0.24 0.10	0.240000 0.100500	0.923077 0.386538	0.260000 0.260000
Dibenz(a,h)anthracene	REG_IV	IHP MWC	5 4	1 1	0.24 0.10	0.240000 0.100500	0.727273 0.304545	0.330000 0.330000
Dibenz(a,h)anthracene	SED_PEL	IHP MWC	5 4	1 1	0.24 0.10	0.240000 0.100500	1.777778 0.744444	0.135000 0.135000
Dibenz(a,h)anthracene	SED_TEL	IHP MWC	5 4	1 1	0.24 0.10	0.240000 0.100500	38.585210 16.157557	0.006220 0.006220
Dibenzofuran	EQSQB_A	IHP LWC MWC	5 7 4	1 2 1	0.07 0.16 0.43	0.067000 0.362266 0.430000	0.029258 0.158195 0.187773	2.290000 2.290000 2.290000
Fluoranthene	EPASQC_A	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	0.159715 0.017742 0.034323 0.196552	6.200000 6.200000 6.200000 6.200000
Fluoranthene	ER_L	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	1.650391 0.183333 0.354670 2.031041	0.600000 0.600000 0.600000 0.600000
Fluoranthene	ER_M	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	0.194164 0.021569 0.041726 0.238946	5.100000 5.100000 5.100000 5.100000
Fluoranthene	REG_IV	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	2.605880 0.289474 0.560005 3.206907	0.380000 0.380000 0.380000 0.380000
Fluoranthene	SED_PEL	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	0.662808 0.073628 0.142438 0.815679	1.494000 1.494000 1.494000 1.494000
Fluoranthene	SED_TEL	IHP LMB LWC MWC	5 5 7 4	5 1 3 3	0.49 0.11 0.12 0.48	0.990234 0.110000 0.212802 1.218625	8.763137 0.973451 1.883203 10.784288	0.113000 0.113000 0.113000 0.113000
Fluorene	ER_L	IHP LWC	5 7	2 2	0.09 0.21	0.197724 0.478767	10.406536 25.198283	0.019000 0.019000

Fluorene	ER_M	IHP LWC	5 7	2 2	0.09 0.21	0.197724 0.478767	0.366156 0.886606	0.540000 0.540000
Fluorene	REG_IV	IHP LWC	5 7	2 2	0.09 0.21	0.197724 0.478767	0.599164 1.450810	0.330000 0.330000
Fluorene	SED_PEL	IHP LWC	5 7	2 2	0.09 0.21	0.197724 0.478767	1.373085 3.324773	0.144000 0.144000
Fluorene	SED_TEL	IHP LWC	5 7	2 2	0.09 0.21	0.197724 0.478767	9.326612 22.583366	0.021200 0.021200
Iron	LEL_MOE	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	21133.33 49080.00 40600.00 21850.00	29098.586823 67472.093481 68827.869767 32899.065151	0.969953 2.249070 2.294262 1.096636	30000.000000 30000.000000 30000.000000 30000.000000
Iron	SEL_MOE	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	21133.33 49080.00 40600.00 21850.00	29098.586823 67472.093481 68827.869767 32899.065151	0.727465 1.686802 1.720697 0.822477	40000.000000 40000.000000 40000.000000 40000.000000
Lead	ER_L	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	35.03 13.70 15.77 71.80	64.883115 19.143937 22.827828 121.678637	1.380492 0.407318 0.485698 2.588907	47.000000 47.000000 47.000000 47.000000
Lead	ER_M	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	35.03 13.70 15.77 71.80	64.883115 19.143937 22.827828 121.678637	0.297629 0.087816 0.104715 0.558159	218.000000 218.000000 218.000000 218.000000
Lead	REG_IV	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	35.03 13.70 15.77 71.80	64.883115 19.143937 22.827828 121.678637	3.089672 0.911616 1.087039 5.794221	21.000000 21.000000 21.000000 21.000000
Lead	SED_PEL	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	35.03 13.70 15.77 71.80	64.883115 19.143937 22.827828 121.678637	0.579314 0.170928 0.203820 1.086416	112.000000 112.000000 112.000000 112.000000
Lead	SED_TEL	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	35.03 13.70 15.77 71.80	64.883115 19.143937 22.827828 121.678637	2.148447 0.633905 0.755888 4.029094	30.200000 30.200000 30.200000 30.200000
Manganese	LEL_MOE	IHP LMB LWC MVC	3 5 3 2	3 5 3 2	803.33 3211.20 1760.00 747.00	1650.715184 6021.788706 2289.905652 1018.491315	3.588511 13.090845 4.978056 2.214112	460.000000 460.000000 460.000000 460.000000

Manganese	SEL_MOE	IHP	3	803.33	1650.715184	1.487131	1110.000000
		LMB	5	3211.20	6021.788706	5.425035	1110.000000
		LWC	3	1760.00	2289.905652	2.062978	1110.000000
		MWC	2	747.00	1018.491315	0.917560	1110.000000
Mercury	ER_L	IHP	3	2.03	4.072683	27.151220	0.150000
		LMB	5	0.12	0.142359	0.949060	0.150000
		LWC	3	0.17	0.199085	1.327235	0.150000
		MWC	2	4.60	5.862750	39.085002	0.150000
Mercury	ER_M	IHP	3	2.03	4.072683	5.736173	0.710000
		LMB	5	0.12	0.142359	0.200506	0.710000
		LWC	3	0.17	0.199085	0.280402	0.710000
		MWC	2	4.60	5.862750	8.257395	0.710000
Mercury	REG_IV	IHP	3	2.03	4.072683	40.726831	0.100000
		LMB	5	0.12	0.142359	1.423590	0.100000
		LWC	3	0.17	0.199085	1.990852	0.100000
		MWC	2	4.60	5.862750	58.627503	0.100000
Mercury	SED_PEL	IHP	3	2.03	4.072683	5.851556	0.696000
		LMB	5	0.12	0.142359	0.204539	0.696000
		LWC	3	0.17	0.199085	0.286042	0.696000
		MWC	2	4.60	5.862750	8.423491	0.696000
Mercury	SED_TEL	IHP	3	2.03	4.072683	31.328332	0.130000
		LMB	5	0.12	0.142359	1.095069	0.130000
		LWC	3	0.17	0.199085	1.531425	0.130000
		MWC	2	4.60	5.862750	45.098080	0.130000
Naphthalene	EQPSQB_A	IHP	5	0.04	0.047791	0.117423	0.407000
		LWC	7	0.03	0.031000	0.076167	0.407000
Naphthalene	ER_L	IHP	5	0.04	0.047791	0.298694	0.160000
		LWC	7	0.03	0.031000	0.193750	0.160000
Naphthalene	ER_M	IHP	5	0.04	0.047791	0.022758	2.100000
		LWC	7	0.03	0.031000	0.014762	2.100000
Naphthalene	REG_IV	IHP	5	0.04	0.047791	0.144821	0.330000
		LWC	7	0.03	0.031000	0.093939	0.330000
Naphthalene	SED_PEL	IHP	5	0.04	0.047791	0.122228	0.391000
		LWC	7	0.03	0.031000	0.079284	0.391000
Naphthalene	SED_TEL	IHP	5	0.04	0.047791	1.381245	0.034600
		LWC	7	0.03	0.031000	0.895954	0.034600
Nickel	ER_L	IHP	3	45.90	145.505469	6.928832	21.000000
		LMB	5	38.30	42.993937	2.047330	21.000000
		LWC	3	30.23	43.938960	2.092331	21.000000

PU-238	SED_RAD	IHP	5	0.31	0.440587	0.000017	26272.000000
		LMB	5	1.10	1.534837	0.000058	26272.000000
		LWC	4	0.37	1.178685	0.000045	26272.000000
		MWC	2	0.86	1.139119	0.000043	26272.000000
PU-239/40	SED_RAD	IHP	5	5.59	8.759958	0.000313	27975.000000
		LWC	4	18.64	59.758823	0.002136	27975.000000
		MWC	2	5.30	7.194125	0.000257	27975.000000
		AET	5	0.03	0.029000	0.080556	0.360000
Phenanthrene	EPASQC_A	IHP	5	0.31	0.654559	0.363644	1.800000
		LMB	5	0.07	0.065000	0.036111	1.800000
		LWC	7	0.37	0.818758	0.454866	1.800000
		MWC	4	0.93	2.517209	1.398450	1.800000
Phenanthrene	ER_L	IHP	5	0.31	0.654559	2.727328	0.240000
		LMB	5	0.07	0.065000	0.270833	0.240000
		LWC	7	0.37	0.818758	3.411493	0.240000
		MWC	4	0.93	2.517209	10.488372	0.240000
Phenanthrene	ER_M	IHP	5	0.31	0.654559	0.436372	1.500000
		LMB	5	0.07	0.065000	0.043333	1.500000
		LWC	7	0.37	0.818758	0.545839	1.500000
		MWC	4	0.93	2.517209	1.678140	1.500000
Phenanthrene	REG_IV	IHP	5	0.31	0.654559	1.983511	0.330000
		LMB	5	0.07	0.065000	0.196970	0.330000
		LWC	7	0.37	0.818758	2.481086	0.330000
		MWC	4	0.93	2.517209	7.627907	0.330000
Phenanthrene	SED_PEL	IHP	5	0.31	0.654559	1.203233	0.544000
		LMB	5	0.07	0.065000	0.119485	0.544000
		LWC	7	0.37	0.818758	1.505070	0.544000
		MWC	4	0.93	2.517209	4.627223	0.544000
Phenanthrene	SED_TEL	IHP	5	0.31	0.654559	7.549696	0.086700
		LMB	5	0.07	0.065000	0.749712	0.086700
		LWC	7	0.37	0.818758	9.443578	0.086700
		MWC	4	0.93	2.517209	29.033556	0.086700
Pyrene	ER_L	IHP	5	0.39	0.831804	1.260309	0.660000
		LMB	5	0.09	0.086000	0.130303	0.660000
		LWC	7	0.08	0.132275	0.200416	0.660000
		MWC	4	0.67	1.499339	2.271726	0.660000
Pyrene	ER_M	IHP	5	0.39	0.831804	0.319925	2.600000
		LMB	5	0.09	0.086000	0.033077	2.600000
		LWC	7	0.08	0.132275	0.050875	2.600000
		MWC	4	0.67	1.499339	0.576669	2.600000

Pyrene	REG_IV	IHP	5	5	0.39	0.831804	2.520618	0.330000
		LMB	5	1	0.09	0.086000	0.260606	0.330000
		LWC	7	3	0.08	0.132275	0.400832	0.330000
		MWC	4	4	0.67	1.499339	4.543453	0.330000
Pyrene	SED_PEL	IHP	5	5	0.39	0.831804	0.594996	1.398000
		LMB	5	1	0.09	0.086000	0.061516	1.398000
		LWC	7	3	0.08	0.132275	0.094617	1.398000
		MWC	4	4	0.67	1.499339	1.072489	1.398000
Pyrene	SED_TEL	IHP	5	5	0.39	0.831804	5.436627	0.153000
		LMB	5	1	0.09	0.086000	0.562092	0.153000
		LWC	7	3	0.08	0.132275	0.864540	0.153000
		MWC	4	4	0.67	1.499339	9.799604	0.153000
Silver	ER_L	IHP	3	3	7.37	21.870242	21.870242	1.000000
		LMB	5	5	0.29	0.324491	0.324491	1.000000
		LWC	3	3	0.29	0.332266	0.332266	1.000000
		MWC	2	2	24.05	31.310814	31.310814	1.000000
Silver	ER_H	IHP	3	3	7.37	21.870242	5.910876	3.700000
		LMB	5	5	0.29	0.324491	0.087700	3.700000
		LWC	3	3	0.29	0.332266	0.089802	3.700000
		MWC	2	2	24.05	31.310814	8.462382	3.700000
Silver	REG_IV	IHP	3	3	7.37	21.870242	10.935121	2.000000
		LMB	5	5	0.29	0.324491	0.162246	2.000000
		LWC	3	3	0.29	0.332266	0.166133	2.000000
		MWC	2	2	24.05	31.310814	15.655407	2.000000
Silver	SED_PEL	IHP	3	3	7.37	21.870242	12.356069	1.770000
		LMB	5	5	0.29	0.324491	0.183329	1.770000
		LWC	3	3	0.29	0.332266	0.187721	1.770000
		MWC	2	2	24.05	31.310814	17.689726	1.770000
Silver	SED_TEL	IHP	3	3	7.37	21.870242	29.836617	0.733000
		LMB	5	5	0.29	0.324491	0.442690	0.733000
		LWC	3	3	0.29	0.332266	0.453297	0.733000
		MWC	2	2	24.05	31.310814	42.715979	0.733000
TH-228	SED_RAD	IHP	5	5	1.44	1.759990	0.000101	17450.000000
		LMB	5	5	1.14	1.602714	0.000092	17450.000000
		LWC	4	4	6.03	6.720305	0.000385	17450.000000
		MWC	1	1	1.50	1.500000	0.000086	17450.000000
TH-230	SED_RAD	IHP	5	5	1.01	1.144154	0.000004	306208.000000
		LMB	5	5	0.84	1.173528	0.000004	306208.000000
		LWC	4	4	1.06	1.192956	0.000004	306208.000000
		MWC	1	1	0.88	0.880000	0.000003	306208.000000
TH-232	SED_RAD	IHP	5	5	1.17	1.253080	0.000049	25440.000000

	LMB	5	5	1.14	1.575231	0.000062	25440.000000
	LWC	4	4	1.28	1.517579	0.000060	25440.000000
	MWC	1	1	1.10	1.100000	0.000043	25440.000000
TOTAL RADIO-STRONTIUM	SED_RAD	5	5	47.58	89.007899	0.005099	17455.000000
	IHP	5	4	28.63	55.535909	0.003182	17455.000000
	LMB	4	3	6.25	13.833095	0.000793	17455.000000
	LWC	2	2	25.50	72.853136	0.004174	17455.000000
	MWC	2	2				
U-233/4	SED_RAD	5	5	2.85	4.177064	0.000464	9001.000000
	IHP	5	5	0.81	1.135598	0.000126	9001.000000
	LMB	2	2	6.95	16.736315	0.001859	9001.000000
	LWC	2	2	4.75	9.485314	0.001054	9001.000000
	MWC	2	2				
U-235	SED_RAD	5	5	0.10	0.161824	0.000018	9207.000000
	IHP	5	5	0.08	0.130476	0.000014	9207.000000
	LMB	2	2	0.07	0.254756	0.000028	9207.000000
	LWC	2	2	0.08	0.282697	0.000031	9207.000000
	MWC	2	2				
U-238	SED_RAD	5	5	1.83	2.767581	0.000331	8351.000000
	IHP	5	5	0.80	1.077611	0.000129	8351.000000
	LMB	2	2	0.24	0.708531	0.000085	8351.000000
	LWC	2	2	1.70	2.331375	0.000279	8351.000000
	MWC	2	2				
Zinc	ER_L	3	3	197.50	393.402164	2.622681	150.000000
	IHP	5	5	399.00	554.862641	3.699084	150.000000
	LMB	3	3	70.27	88.289810	0.588599	150.000000
	LWC	2	2	811.00	1423.433897	9.489559	150.000000
	MWC	2	2				
Zinc	ER_M	3	3	197.50	393.402164	0.959517	410.000000
	IHP	5	5	399.00	554.862641	1.353324	410.000000
	LMB	3	3	70.27	88.289810	0.215341	410.000000
	LWC	2	2	811.00	1423.433897	3.471790	410.000000
	MWC	2	2				
Zinc	REG_IV	3	3	197.50	393.402164	5.785326	68.000000
	IHP	5	5	399.00	554.862641	8.159745	68.000000
	LMB	3	3	70.27	88.289810	1.298380	68.000000
	LWC	2	2	811.00	1423.433897	20.932851	68.000000
	MWC	2	2				
Zinc	SED_PEL	3	3	197.50	393.402164	1.451669	271.000000
	IHP	5	5	399.00	554.862641	2.047464	271.000000
	LMB	3	3	70.27	88.289810	0.325793	271.000000
	LWC	2	2	811.00	1423.433897	5.252524	271.000000
	MWC	2	2				
Zinc	SED_TEL	3	3	197.50	393.402164	3.172598	124.000000
	IHP	5	5	399.00	554.862641	4.474699	124.000000
	LMB	3	3	70.27	88.289810	0.712015	124.000000
	LWC	2	2	811.00	1423.433897	11.479306	124.000000
	MWC	2	2				
bis(2-Ethylhexyl)phthalate	EQPSOB_A	5	5	0.30	0.481422	0.000001	890000.000000
	IHP	5	5	0.11	0.163751	0.000000	890000.000000
	LMB	5	5				

bis(2-Ethylhexyl)phthalate	SED_PEL	LWC	7	5	0.16	0.314450	0.000000	890000.000000
		MWC	4	4	0.96	2.226516	0.000003	890000.000000
		IHP	5	5	0.30	0.481422	0.181875	2.647000
		LMB	5	3	0.11	0.163751	0.061863	2.647000
bis(2-Ethylhexyl)phthalate	SED_TEL	LWC	7	5	0.16	0.314450	0.118795	2.647000
		MWC	4	4	0.96	2.226516	0.841147	2.647000
		IHP	5	5	0.30	0.481422	2.645178	0.182000
		LMB	5	3	0.11	0.163751	0.899729	0.182000
		LWC	7	5	0.16	0.314450	1.727748	0.182000
		MWC	4	4	0.96	2.226516	12.233605	0.182000

Table B.6. Comparison of above-background, maximum-detected soil contaminant concentrations to screening benchmarks for toxicity to soil invertebrates.

Reach	Chemical	Mean Conc. (mg/kg)	Maximum detect. conc. (mg/kg)	Earthworm benchmark (mg/kg)	Hazard Quotient
IHP	Cadmium	1.211	1.5	20	0.08
	Chromium VI²	48.93	71.60	0.4	200
	Copper	28.19	45.1	50	0.9
	Fluorene	0.38	0.14	30	0.005
	Lead	59.39	111	500	0.2
	Mercury	17.33	76.4	0.1	800
	Nickel	53.63	310	200	2
	Zinc	95.92	159	200	0.8
LWOC	Cadmium	0.8325	1	20	0.05
	Chromium VI²	55.45	95.30	0.4	200
	Lead	35.3	50.4	500	0.1
	Mercury	2.958	5.1	0.1	50
	N-Nitrosodiphenylamine ¹	0.4275	0.42	20	0.021
	Selenium	1.075	1.3	70	0.02
	Zinc	104.4	190	200	1
MWOC	Cadmium	0.825	0.86	20	0.04
	Mercury	1.555	2.5	0.1	30
	Selenium	1.23	1.6	70	0.02
	Zinc	83.75	123	200	0.6
LMBC/	Cadmium	0.93	1.1	20	0.06
	Mercury	0.506	2	0.1	20
	Selenium	0.902	1.3	70	0.02

Boldface type indicates exceedence of benchmark.

¹ Background concentration assumed to be zero.

² Concentrations of Chromium VI are compared to benchmarks for total chromium.

Table B.7. Comparison of above-background, maximum-detected soil contaminant concentrations to screening benchmarks for toxicity to plants.

Reach	Chemical	Mean Conc. (mg/kg)	Maximum detect. conc. (mg/kg)	Benchmark (mg/kg)	Hazard Quotient	
IHP	Barium	180.27778	268	500	0.5	
	Boron ¹	6.4055556	10.2	0.5	20	
	Cadmium	1.2111111	1.5	3	0.5	
	Chromium VI ³	48.93	71.60	1	70	
	Copper	28.188889	45.1	100	0.5	
	Di-n-butyl phthalate ²	0.1494	0.11	200	0.0006	
	Lead	59.394444	111	50	2	
	Lithium ¹	26.844444	41.4	2	20	
	Manganese	1260.1111	4150	500	8	
	Mercury	17.33	76.4	0.3	300	
	Molybdenum ¹	4.2722222	5.2	2	3	
	Nickel	53.627778	310	30	10	
	PCB, total ²	1.0866	2.194	40	0.05	
	Silver	6.6	15.2	2	8	
	Thallium	0.8483333	0.91	1	0.9	
	Tin	10.522222	12.8	50	0.3	
	Zinc	95.916667	159	50	3	
	LWOC	Boron ¹	9.75	10.2	0.5	20
		Cadmium	0.8325	1	3	0.3
Chromium VI ³		55.45	95.30	1	100	
Di-n-butyl phthalate ²		0.18975	0.17	200	0.0009	
Lead		35.3	50.4	50	1	
Lithium ¹		13.95	16.2	2	8	
Manganese		969.25	1280	500	3	
Mercury		2.9575	5.1	0.3	20	
Molybdenum ¹		3.6	3.8	2	2	
PCB, total ²		0.5115	0.736	40	0.02	
Selenium		1.075	1.3	1	1	
Silver		3.7	6.8	2	3	
Tin		6.95	7.3	50	0.1	
Zinc		104.35	190	50	4	
MWOC	Boron ¹	10.45	10.9	0.5	20	
	Cadmium	0.825	0.86	3	0.3	
	Di-n-butyl phthalate ²	0.042	0.049	200	0.0002	
	Lithium ¹	11.2	13	2	7	
	Mercury	1.555	2.5	0.3	8	
	Molybdenum ¹	3.85	4	2	2	
	PCB, total ²	0.633	0.94	40	0.02	
	Selenium	1.23	1.6	1	2	

	Silver	2.5	3.9	2	2
	Tin	7.4	7.7	50	0.2
	Zinc	83.75	123	50	2
LMBC/	Antimony	10.66	12.1	5	2
	Barium	220.6	409	500	0.8
	Boron ¹	6.36	9.4	0.5	20
	Cadmium	0.93	1.1	3	0.4
	Di-n-butyl phthalate ²	0.1635714	0.42	200	0.002
	Lead	16.36	18.8	50	0.4
	Lithium ¹	20.02	25.9	2	10
	Manganese	4502	9400	500	20
	Mercury	0.506	2	0.3	7
	Molybdenum ¹	5.56	7.3	2	4
	PCB, total ²	0.3752857	0.315	40	0.008
	Selenium	0.902	1.3	1	1
	Silver	1.208	1.4	2	0.7
	Thallium	1.11	1.4	1	1
	Tin	8.78	11.3	50	0.2

Boldface type indicates exceedence of benchmark.

¹ Background concentration not available.

² Background concentration assumed to be zero.

³ Concentrations of Chromium VI are compared to benchmarks for total chromium.

Table B.8. Comparison of above-background, maximum-detected contaminant concentrations in seeps or springs to screening benchmarks for toxicity to plants in solution.

Reach	Seep	Chemical	Mean conc. (mg/kg)	Maximum detect conc. (mg/kg)	Benchmark (mg/kg)	Hazard Quotient
EWC	WOCET	ALUMINUM	1.071	1.5	0.2	8
	WOCET	IRON	1.084	1.61	10	0.2
	WOCET	MANGANESE	0.3545	0.649	4	0.2
HRT	SW9-1	IRON	0.3663	0.621	10	0.06
	SW9-1	MANGANESE	0.2477	0.311	4	0.08
	SW9-2	ALUMINUM	0.858	1.46	0.2	7
	SW9-2	IRON	1.867	3.37	10	0.3
	SW9-2	MANGANESE	1.175	2.18	4	0.5
	SW9-2	ZINC	0.1143	0.288	0.4	0.7
IHP	WCTRIB-3	BORON ¹	0.199	0.199	1	0.2
	WCTRIB-3	IRON	0.378	0.378	10	0.04
	WCTRIB-3	MANGANESE	0.585	0.585	4	0.1
	SW2-2	IRON	0.394	0.394	10	0.04
	SW2-2	MANGANESE	0.796	0.796	4	0.2
LMB	MBTRIB-3	IRON	0.2945	0.414	10	0.04
	MBTRIB-3	MANGANESE	0.3685	0.399	4	0.1
	MID. DRAIN.	BORON ¹	0.1939	0.274	1	0.3
	MID. DRAIN.	IRON	2.0316	8.61	10	0.9
	MID. DRAIN.	MANGANESE	1.8748	4.74	4	1
	SW2-5	BORON ¹	0.215	0.336	1	0.3
SW2-5	IRON	0.3486	1.54	10	0.2	

SW2-5	MANGANESE	0.945	3.06	4	0.7
SW5-2	BORON 1	0.175	0.175	1	0.2
SW5-2	IRON	0.089	0.089	10	0.009
SW5-4	BORON 1	0.1457	0.234	1	0.2
SW5-4	IRON	0.4035	1.11	10	0.1
SW5-4	MANGANESE	0.7902	2.62	4	0.7
LWC					
EAST SEEP	ALUMINUM	0.8376	2.41	0.2	10
EAST SEEP	ARSENIC	0.0275	0.004	0.001	4
EAST SEEP	BORON 1	0.08756	0.14	1	0.1
EAST SEEP	CHROMIUM	0.015	0.033	0.05	0.7
EAST SEEP	IRON	0.6446	1.7	10	0.2
EAST SEEP	MANGANESE	0.2593	0.552	4	0.1
EAST SEEP	MOLYBDENUM 1	0.01467	0.015	0.5	0.03
EAST SEEP	NICKEL	0.02767	0.04	0.2	0.2
MV-1	IRON	2.1	2.11	10	0.2
MV-1	MANGANESE	3.965	3.99	4	1
SW7-3	ALUMINUM	0.452	0.775	0.2	4
SW7-3	BORON 1	0.08713	0.101	1	0.1
SW7-3	CHROMIUM	0.056	0.091	0.05	2
SW7-3	IRON	0.3488	0.693	10	0.07
SW7-3	MOLYBDENUM 1	0.014	0.017	0.5	0.03
SW7-5	ALUMINUM	0.8017	2.19	0.2	10
SW7-5	BORON 1	0.0951	0.077	1	0.08
SW7-5	IRON	0.8155	2.73	10	0.3
SW7-5	MANGANESE	0.2711	2.35	4	0.6
SW7-6	ALUMINUM	0.761	0.761	0.2	4
SW7-6	IRON	0.431	0.431	10	0.04
SW7-8	ALUMINUM	0.412	0.412	0.2	2
SW7-8	IRON	0.368	0.368	10	0.04
SW7-8	MANGANESE	0.126	0.126	4	0.03
WCTRIB-1	ALUMINUM	0.572	0.957	0.2	5
WCTRIB-1	IRON	0.437	0.659	10	0.07

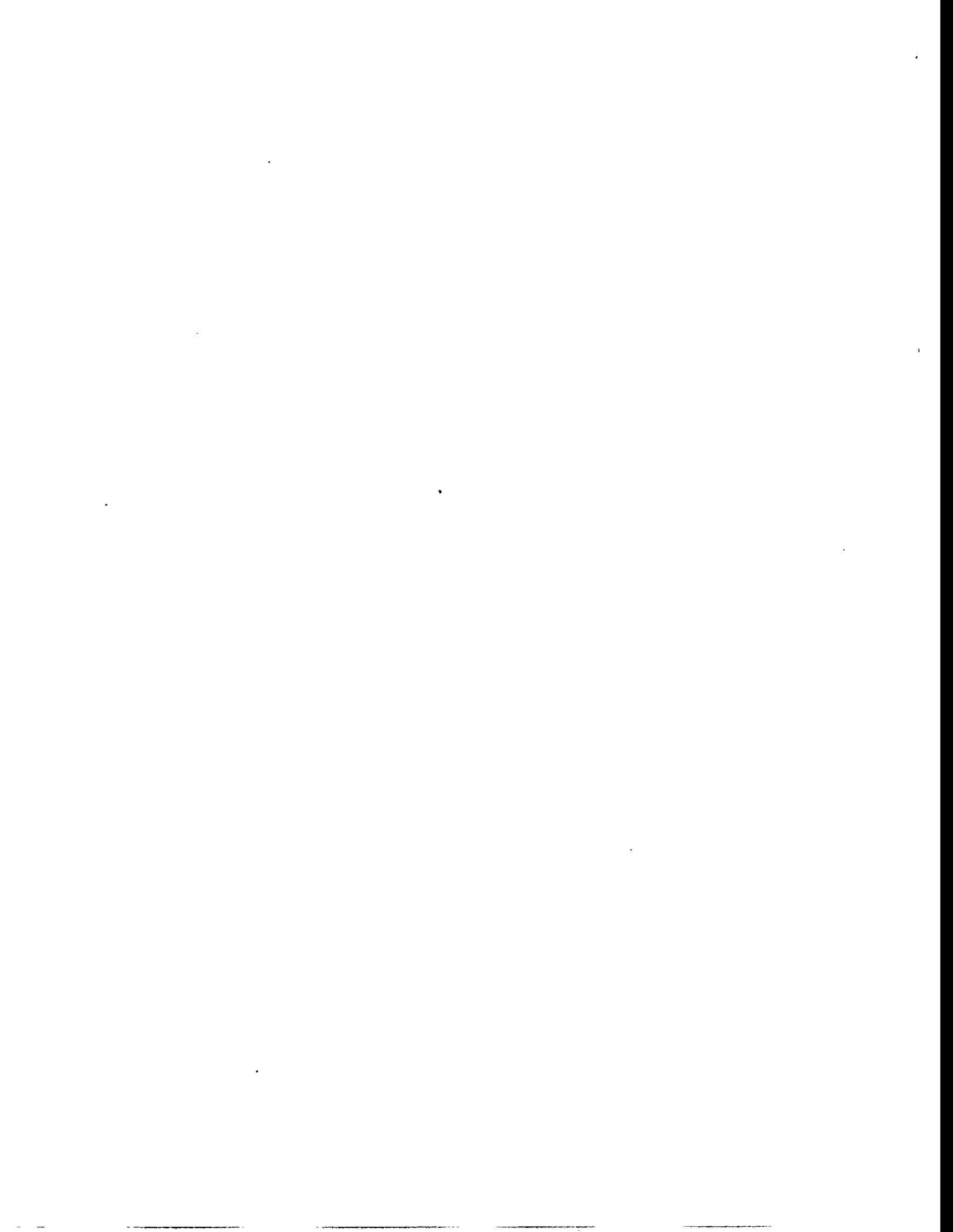
	WCTRIB-1	MANGANESE	0.213	0.369	4	0.09
MWC	SW2-1	BORON ¹	0.019	0.019	1	0.02
	SW2-1	IRON	2.3	2.3	10	0.2
	SW2-1	MANGANESE	0.992	0.992	4	0.2
	SW2-3	IRON	0.225	0.225	10	0.02
	SW2-4	IRON	0.5975	0.809	10	0.08
	SW2-4	MANGANESE	1.5775	2.26	4	0.6
	SW2-4	SILVER ¹	0.003	0.003	0.1	0.03
	WCTRIB-2	IRON	0.188	0.188	10	0.02
UMB	MBTRIB-2A	IRON	0.218	0.218	10	0.02
	MV-3	IRON	0.172	0.172	10	0.02
Unknown	SW2-6	BORON ¹	0.133	0.118	1	0.1
	SW2-6	IRON	2.857	8.52	10	0.9
	SW2-6	MANGANESE	1	1.36	4	0.3
	SW2-7	BORON ¹	0.1465	0.141	1	0.1
	SW2-7	IRON	6.8925	17	10	2
	SW2-7	MANGANESE	1.4475	1.9	4	0.5
W4T	BTT	BORON ¹	0.1126	0.113	1	0.1
	BTT	IRON	0.1414	0.202	10	0.02
	BTT	MOLYBDENUM ¹	0.06743	0.171	0.5	0.3
	BTT	NICKEL	0.04114	0.051	0.2	0.3
	SW4-1	ARSENIC	0.004	0.006	0.001	6
	SW4-1	IRON	14.881	28.8	10	3
	SW4-1	MANGANESE	2.0935	3.81	4	1
	SW4-2	BORON ¹	0.1547	0.231	1	0.2
	SW4-2	IRON	2.4103	7.06	10	0.7
	SW4-2	LITHIUM ¹	0.113	0.113	3	0.04
	SW4-2	MANGANESE	0.7535	1.23	4	0.3
	SW4-2	MOLYBDENUM	0.01467	0.008	0.5	0.02

SW4-2	NICKEL	6.17	9.46	0.2	50
WAG4 MS1	ALUMINUM	0.1535	1.02	0.2	5
WAG4 MS1	BERYLLIUM	0.000909	0.002	0.5	0.004
WAG4 MS1	BORON ¹	0.1735	0.196	1	0.2
WAG4 MS1	IRON	0.1947	1.12	10	0.1
WAG4 MS1	MANGANESE	0.2095	0.644	4	0.2
WAG4 MS1	MOLYBDENUM ¹	0.01418	0.007	0.5	0.01
WAG4 MS1	NICKEL	0.03364	0.067	0.2	0.3
WAG4 T2A	ALUMINUM	0.4713	1	0.2	5
WAG4 T2A	BORON ¹	0.1456	0.206	1	0.2
WAG4 T2A	CHROMIUM	0.008	0.035	0.05	1
WAG4 T2A	IRON	0.4763	1.43	10	0.1
WAG4 T2A	MANGANESE	0.1254	0.41	4	0.1
WOL					
RS-1	IRON	0.601	0.601	10	0.06
RS-1	MANGANESE	1.35	1.35	4	0.3
RS-3A	ALUMINUM	10.012	17.7	0.2	90
RS-3A	ARSENIC	0.01688	0.024	0.001	20
RS-3A	BORON ¹	0.1521	0.118	1	0.1
RS-3A	CHROMIUM	0.078	0.096	0.05	2
RS-3A	COPPER	0.0194	0.028	0.03	0.9
RS-3A	IRON	7.728	13.9	10	1
RS-3A	MANGANESE	0.2344	0.531	4	0.1
RS-3A	MOLYBDENUM ¹	0.0483	0.056	0.5	0.1
RS-3A	NICKEL	0.097	0.108	0.2	0.5
RS-3A	SILVER ¹	0.0046	0.006	0.1	0.06
RS-3A	VANADIUM	0.022	0.034	0.2	0.2
RS-3B	ALUMINUM	1.345	1.42	0.2	7
RS-3B	BORON ¹	0.077	0.034	1	0.03
RS-3B	IRON	1.065	1.08	10	0.1
SW7-1	IRON	6.375	6.57	10	0.7
SW7-1	MANGANESE	6.39	6.4	4	2
SW7-2	ALUMINUM	0.574	0.652	0.2	3

SW7-2	COBALT	0.094	0.134	0.06	2
SW7-2	COPPER	0.015	0.025	0.03	0.8
SW7-2	IRON	0.586	0.642	10	0.06
FRENCH DR S	TETRACHLOROETHENE	0.055	0.055	10	0.006
WAG6 MS3B	TETRACHLOROETHENE	0.001	0.001	10	0.0001

Boldface type indicates exceedence of benchmark.

1 Background concentration not available.



Appendix C

DATA TABLES FOR RISKS TO WILDLIFE

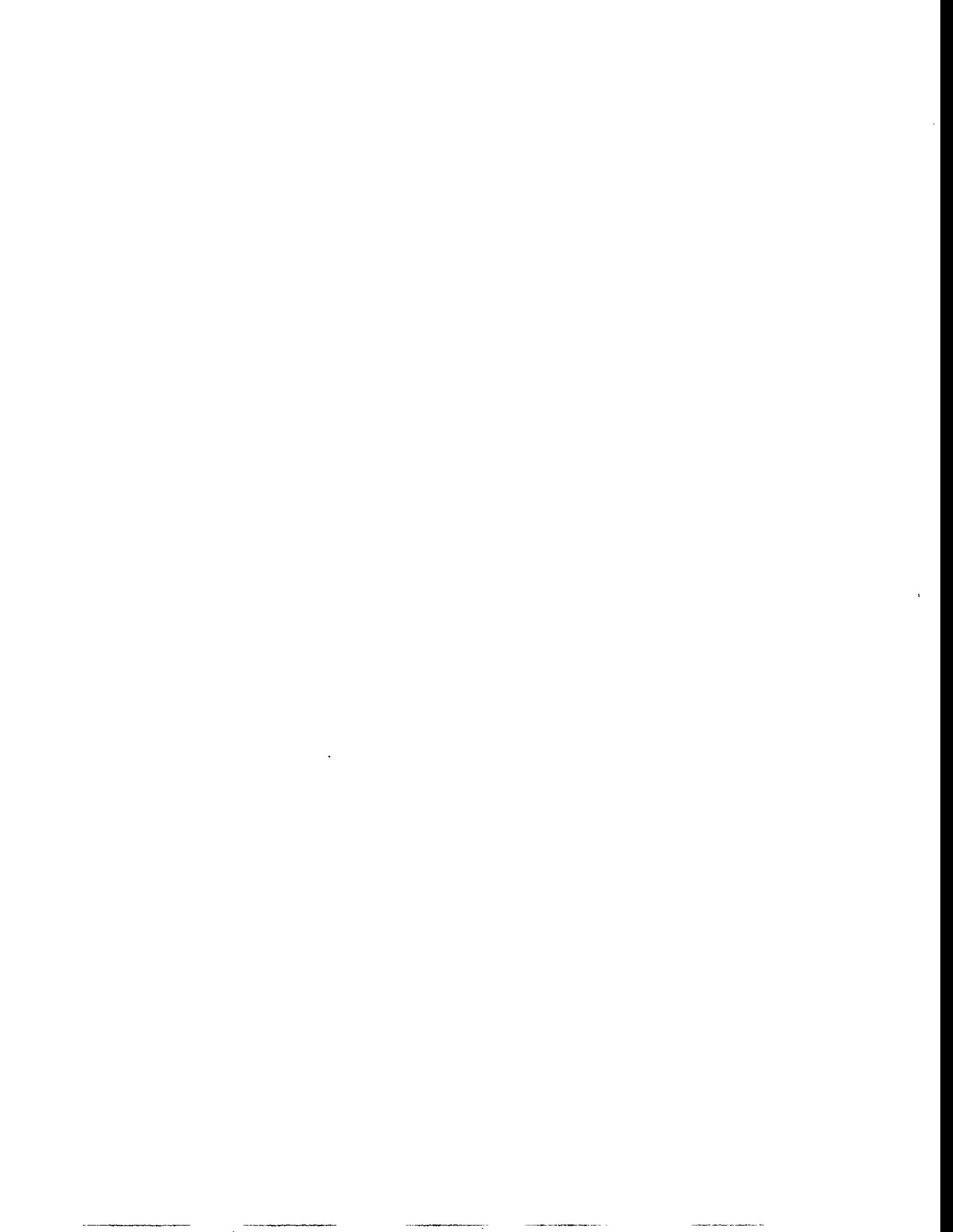


Table C.1. Point Estimates of Contaminant Exposure of Mink to Fish from White Oak Creek.

Location	Contaminant	Dietary Exposure (mg/kg/d)	NOAEL HQ	LOAEL HQ
HINDSCR ¹	Mercury	0.005	0.33	0.20
NTK0.2	Mercury	0.010	0.64	0.38
WCK3.5	Mercury	0.014	0.91	0.55
WCK2.9	Mercury	0.023	1.51	0.91
MEK0.2	Mercury	0.007	0.45	0.27
WCK2.3	Mercury	0.015	0.98	0.59
WOL	Mercury	0.009	0.58	0.35
WCK0.9	Mercury	0.006	0.37	0.22
WCK0.3	Mercury	0.010	0.64	0.38
HINDSCR ¹	PCB-1260	0.003	0.02	0.00
NTK0.2	PCB-1260	0.049	0.35	0.07
WCK3.5	PCB-1260	0.080	0.57	0.12
WCK2.9	PCB-1260	0.093	0.66	0.13
MEK0.2	PCB-1260	0.039	0.28	0.06
WCK2.3	PCB-1260	0.099	0.70	0.14
WOL	PCB-1260	0.129	0.92	0.19
WCK0.9	PCB-1260	0.052	0.37	0.08
WCK0.3	PCB-1260	0.058	0.41	0.08

¹Reference location.

Table C.2. Point Estimates of Contaminant Exposure of Belted Kingfisher to Fish from White Oak Creek.

Location	Contaminant	Dietary Exposure	NOAEL HO	LOAEL HO
HINDSCR ¹	Mercury	0.033	5.54	0.52
NTK0.2	Mercury	0.065	10.80	1.01
WCK3.5	Mercury	0.092	15.41	1.44
WCK2.9	Mercury	0.154	25.60	2.40
MEK0.2	Mercury	0.046	7.69	0.72
WCK2.3	Mercury	0.100	16.66	1.56
WOL	Mercury	0.059	9.83	0.92
WCK0.9	Mercury	0.038	6.26	0.59
WCK0.3	Mercury	0.065	10.77	1.01
HINDSCR ¹	PCB-1260	0.022	0.12	0.01
NTK0.2	PCB-1260	0.335	1.86	0.19
WCK3.5	PCB-1260	0.543	3.02	0.30
WCK2.9	PCB-1260	0.630	3.50	0.35
MEK0.2	PCB-1260	0.263	1.46	0.15
WCK2.3	PCB-1260	0.668	3.71	0.37
WOL	PCB-1260	0.875	4.86	0.49
WCK0.9	PCB-1260	0.354	1.97	0.20
WCK0.3	PCB-1260	0.391	2.17	0.22

¹Reference location.

Table C.3. Summary Statistics for Soil-Biota Uptake Factors

Contaminant	Media	Detects	Mean	Standard Error	UCL95	Maximum	Minimum
Aluminum	Browse	12	0.004	0.001	0.007	0.016	0.001
Americium-241	Browse	4	1.875	0.346	2.553	2.489	0.894
Arsenic	Browse	6	0.008	0.002	0.012	0.015	0.001
Barium	Browse	12	0.090	0.021	0.131	0.241	0.010
Beryllium	Browse	1	0.002	0.000	0.000	0.002	0.002
Cadmium	Browse	8	1.158	0.761	2.650	5.951	0.002
Calcium	Browse	12	1.717	0.376	2.455	4.257	0.117
Chromium	Browse	12	0.024	0.005	0.033	0.065	0.001
Cobalt	Browse	12	0.003	0.001	0.005	0.010	0.001
Copper	Browse	12	0.305	0.071	0.443	0.864	0.001
Iron	Browse	12	0.001	0.000	0.002	0.002	0.000
Lead	Browse	12	0.007	0.002	0.011	0.022	0.000
Lithium	Browse	10	0.005	0.001	0.008	0.010	0.000
Magnesium	Browse	10	0.980	0.198	1.368	2.350	0.489
Manganese	Browse	10	0.118	0.049	0.215	0.414	0.015
Mercury	Browse	9	0.040	0.010	0.059	0.086	0.007
Nickel	Browse	12	0.051	0.018	0.085	0.200	0.001
Plutonium-239/240	Browse	1	1.800	0.000	0.000	1.800	1.800
Potassium	Browse	10	6.596	1.062	8.677	12.285	2.744
Selenium	Browse	3	0.409	0.395	1.184	1.200	0.011
Silver	Browse	1	0.001	0.000	0.000	0.001	0.001
Sodium	Browse	11	0.352	0.060	0.470	0.693	0.011
Uranium	Browse	1	0.000	0.000	0.000	0.000	0.000
Vanadium	Browse	1	0.026	0.000	0.000	0.026	0.026
Zinc	Browse	12	0.365	0.094	0.550	1.203	0.004
Aluminum	Canopy	12	0.006	0.001	0.009	0.016	0.000
Americium-241	Canopy	6	2.774	1.493	5.700	10.194	0.748
Arsenic	Canopy	6	0.017	0.006	0.029	0.039	0.001
Barium	Canopy	12	0.153	0.045	0.241	0.471	0.010
Beryllium	Canopy	5	0.017	0.010	0.036	0.055	0.002
Cadmium	Canopy	7	0.136	0.083	0.299	0.621	0.003
Calcium	Canopy	12	2.511	0.505	3.501	5.808	0.081
Chromium	Canopy	12	0.033	0.011	0.055	0.147	0.001
Cobalt	Canopy	12	0.003	0.001	0.004	0.009	0.001
Copper	Canopy	12	0.118	0.021	0.160	0.258	0.001
Curium-244	Canopy	1	1.067	0.000	0.000	1.067	1.067
Iron	Canopy	12	0.003	0.001	0.006	0.019	0.000
Lead	Canopy	12	0.032	0.015	0.062	0.177	0.000
Lithium	Canopy	10	0.016	0.011	0.037	0.113	0.000
Magnesium	Canopy	10	1.147	0.206	1.550	2.615	0.443
Manganese	Canopy	10	0.177	0.068	0.310	0.733	0.015

Table 6.3 (Continued)

Contaminant	Media	Detects	Mean	Standard Error	UCL95	Maximum	Minimum
Mercury	Canopy	9	0.044	0.012	0.068	0.128	0.009
Nickel	Canopy	12	0.076	0.026	0.126	0.248	0.003
Plutonium-239/240	Canopy	2	4.025	1.175	6.328	5.200	2.850
Potassium	Canopy	10	3.205	0.581	4.345	5.691	0.288
Selenium	Canopy	4	0.024	0.006	0.035	0.040	0.017
Silver	Canopy	3	0.020	0.010	0.040	0.036	0.001
Sodium	Canopy	11	0.336	0.059	0.452	0.700	0.012
Uranium	Canopy	1	0.001	0.000	0.000	0.001	0.001
Zinc	Canopy	12	0.216	0.043	0.301	0.521	0.019
Aluminum	Herbacious	12	0.019	0.004	0.027	0.050	0.002
Americium-241	Herbacious	3	1.497	0.394	2.269	2.178	0.813
Arsenic	Herbacious	11	0.015	0.002	0.018	0.023	0.007
Barium	Herbacious	12	0.096	0.016	0.126	0.188	0.019
Beryllium	Herbacious	6	0.014	0.003	0.019	0.020	0.004
Cadmium	Herbacious	8	0.202	0.096	0.390	0.709	0.013
Calcium	Herbacious	12	0.725	0.179	1.076	1.983	0.024
Chromium	Herbacious	12	0.043	0.007	0.057	0.081	0.002
Cobalt	Herbacious	12	0.007	0.001	0.010	0.014	0.002
Copper	Herbacious	12	0.137	0.028	0.191	0.316	0.001
Curium-244	Herbacious	1	1.093	0.000	0.000	1.093	1.093
Iron	Herbacious	12	0.009	0.002	0.012	0.022	0.001
Lead	Herbacious	12	0.017	0.004	0.025	0.052	0.000
Lithium	Herbacious	10	0.015	0.003	0.020	0.029	0.002
Magnesium	Herbacious	10	0.681	0.124	0.923	1.481	0.244
Manganese	Herbacious	10	0.027	0.007	0.040	0.080	0.011
Mercury	Herbacious	8	0.031	0.008	0.045	0.067	0.005
Nickel	Herbacious	12	0.026	0.008	0.043	0.103	0.001
Plutonium-239/240	Herbacious	2	4.200	0.800	5.768	5.000	3.400
Potassium	Herbacious	10	3.971	1.054	6.036	13.008	1.517
Selenium	Herbacious	2	0.012	0.002	0.015	0.014	0.011
Silver	Herbacious	3	0.006	0.005	0.016	0.016	0.001
Sodium	Herbacious	11	0.364	0.064	0.490	0.707	0.013
Thallium	Herbacious	1	0.006	0.000	0.000	0.006	0.006
Uranium	Herbacious	1	0.002	0.000	0.000	0.002	0.002
Vanadium	Herbacious	4	0.041	0.005	0.050	0.052	0.030
Zinc	Herbacious	12	0.367	0.071	0.507	0.769	0.016
Aluminum	Small	9	0.003	0.000	0.004	0.005	0.001
	Mammal						
Americium-241	Small	5	1.484	0.400	2.268	2.889	0.735
	Mammal						
Arsenic	Small	9	0.004	0.001	0.006	0.008	0.001
	Mammal						

Table 6.3 (Continued)

Contaminant	Media	Detects	Mean	Standard Error	UCL95	Maximum	Minimum
Barium	Small Mammal	9	0.030	0.008	0.045	0.075	0.004
Cadmium	Small Mammal	6	0.069	0.051	0.168	0.320	0.004
Calcium	Small Mammal	9	3.610	1.141	5.846	9.378	0.243
Chromium	Small Mammal	9	0.044	0.023	0.088	0.221	0.001
Cobalt	Small Mammal	9	0.006	0.002	0.010	0.020	0.003
Copper	Small Mammal	9	0.313	0.076	0.461	0.749	0.001
Curium-244	Small Mammal	1	2.693	0.000	0.000	2.693	2.693
Iron	Small Mammal	9	0.004	0.001	0.005	0.006	0.001
Lead	Small Mammal	9	0.015	0.006	0.027	0.047	0.000
Lithium	Small Mammal	8	0.007	0.004	0.015	0.033	0.001
Magnesium	Small Mammal	7	0.484	0.085	0.651	0.916	0.308
Manganese	Small Mammal	7	0.003	0.001	0.005	0.006	0.002
Mercury	Small Mammal	7	0.282	0.146	0.569	1.085	0.009
Nickel	Small Mammal	9	0.070	0.021	0.112	0.196	0.003
PCB-1260	Small Mammal	8	1.831	0.676	3.157	5.220	0.072
Plutonium-239/240	Small Mammal	2	2.267	0.467	3.181	2.733	1.800
Potassium	Small Mammal	7	2.967	0.494	3.935	5.112	1.812
Selenium	Small Mammal	7	0.162	0.021	0.204	0.263	0.098
Silver	Small Mammal	1	0.000	0.000	0.000	0.000	0.000
Sodium	Small Mammal	8	5.275	1.201	7.630	10.223	0.269
Uranium	Small Mammal	2	0.000	0.000	0.000	0.000	0.000
Zinc	Small Mammal	9	0.868	0.197	1.254	1.863	0.016
Aluminum	Earthworm	20	0.053	0.010	0.073	0.197	0.008

Table 6.3 (Continued)

Contaminant	Media	Detects	Mean	Standard Error	UCL95	Maximum	Minimum
Americium-241	Earthworm	4	1.970	0.601	3.148	3.644	0.800
Aroclor-1254	Earthworm	3	0.470	0.154	0.772	0.625	0.162
Aroclor-1260	Earthworm	13	4.256	1.778	7.740	22.500	0.000
Arsenic	Earthworm	17	0.243	0.070	0.380	0.909	0.006
Barium	Earthworm	20	0.088	0.016	0.119	0.310	0.005
Beryllium	Earthworm	10	0.296	0.170	0.628	1.429	0.000
BetaParticle	Earthworm	6	0.078	0.036	0.148	0.240	0.004
CO-60	Earthworm	1	0.000			0.000	0.000
CS-137	Earthworm	5	0.005	0.003	0.011	0.015	0.000
Cadmium	Earthworm	14	6.004	3.018	11.919	44.553	0.253
Calcium	Earthworm	20	0.689	0.157	0.997	2.513	0.023
Chromium	Earthworm	19	2.087	0.710	3.479	11.416	0.065
Cobalt	Earthworm	17	0.139	0.021	0.181	0.321	0.031
Copper	Earthworm	20	0.538	0.266	1.060	5.498	0.002
Curium-244	Earthworm	4	0.587	0.162	0.905	0.886	0.152
Iron	Earthworm	20	0.038	0.006	0.049	0.100	0.006
K-40	Earthworm	6	0.139	0.040	0.217	0.269	0.056
Lead	Earthworm	19	0.171	0.107	0.381	2.087	0.000
Lithium	Earthworm	12	0.083	0.024	0.131	0.253	0.008
Magnesium	Earthworm	18	0.220	0.031	0.281	0.539	0.069
Manganese	Earthworm	18	0.060	0.009	0.077	0.127	0.012
Mercury	Earthworm	15	1.909	1.068	4.002	16.250	0.030
Molybdenum	Earthworm	2	1.939	0.153	2.238	2.091	1.786
Nickel	Earthworm	14	2.551	0.635	3.796	7.802	0.038
Plutonium-239	Earthworm	1	2.500			2.500	2.500
Potassium	Earthworm	18	3.016	0.535	4.064	8.372	0.567
Selenium	Earthworm	14	1.798	0.922	3.606	13.733	0.301
Silver	Earthworm	10	4.527	2.027	8.500	19.500	0.001
Sodium	Earthworm	19	20.976	7.285	35.254	122.388	0.146
Strontium	Earthworm	2	0.212	0.066	0.342	0.278	0.146
Thallium	Earthworm	2	0.000	0.000	0.000	0.000	0.000
Total Radio Strontium	Earthworm	1	0.169			0.169	0.169
Uranium	Earthworm	2	0.033	0.030	0.092	0.063	0.003
Vanadium	Earthworm	6	0.039	0.014	0.067	0.088	0.000
Zinc	Earthworm	20	2.419	0.518	3.435	7.305	0.032

Table C.4. Life history parameters for the short-tailed shrew (*Blarina brevicauda*)

Parameter	Value ^a	Comments	Reference
Body Weight	0.015 ± 0.00078 kg	New Hampshire (field)	Schlessinger and Potter 1974
Food Consumption Rate	0.01 kg/d	larch sawfly diet (lab)	Buckner 1964
	0.00795 ± 0.00017 kg/d mean = 0.009 kg/d	mealworm diet (lab)	Barrett and Stueck 1976
Water Consumption Rate	0.223 ml/g bw/d		Chew 1951
	0.033 L/d	assuming a 0.015 kg bw	
Soil Consumption Rate	13% of diet		Talmage and Walton 1993
	0.00117 kg/d	assuming diet of 0.009 kg/d	
Diet Composition	earthworms 31.4% slugs/snails 27.1% soil/litter invert 13.2% fungi 8.4% misc. animals 8.1% coleoptera 5.9% vegetation 5.4%	percent volume in diet in summer in New York	Whitaker and Ferraro 1963
Home Range	0.39 ± 0.036 ha	Manitoba bog	Buckner 1966
Habitat Requirements	broad and variable but requires >50% herbaceous cover		Miller and Getz 1977
	forest, wetlands, and grasslands. most abundant in hardwood forests with deep litter and humus.		van Zyll de Jong 1983
Population Density	2.3 /ha - winter 5.2 /ha - spring 9.3 /ha -summer 8.1 /ha - fall	Illinois - alfalfa, tallgrass, and bluegrass; means derived from graph.	Getz 1989
	2.5-45/ha	Depending on habitat	
Behavior	nocturnal, semifossorial, spends little time above surface		George et al. 1986
	active year-round - does not hibernate		EPA 1993a

Parameter	Value^a	Comments	Reference
Other	appear to be unpalatable to most predators due to lateral gland		van Zyll de Jong 1983

^a Suggested values for use in exposure assessment are in bold.

Table C.5. Life history parameters for the white-footed mouse (*Peromyscus leucopus*)

Parameter	Value ^a	Comments	Reference
Body Weight	0.022 kg		Green and Millar 1987
Food Consumption Rate	0.0034 kg/d	lab study	Green and Millar 1987
Water Consumption Rate	0.0066 L/d	nonreproductive ♀ (lab)	Oswald et al. 1993
Soil Consumption Rate	<2%		Beyer et al. 1994
	0.000068 kg/d	assuming diet of 0.0034 kg/d and a 2% soil consumption rate	
Diet Composition	omnivorous and opportunistic		
	arthropods - 57% seeds, fruit, vegetation - 34%	Virginia	Wolff et al. 1985
	arthropods - 30% seeds, fruit, vegetation - 67%	Indiana	Whitaker 1966
	arthropods - 50% seeds, fruit, vegetation - 48%	Illinois	Batzli 1977
Home Range	0.059 ha	mean: ♂+♀; Virginia, mixed deciduous forest	Wolff 1985
Habitat Requirements	wooded, brushy areas; sometimes open areas		Burt and Grossenheider 1976
Population Density	6 - 57 /ha	Virginia, mixed deciduous forest	Wolff 1985
Behavior	while semi-arboreal, spends most of time on ground.		Lackey et al. 1985
	primarily nocturnal		
	enters torpor to reduce metabolic demands in winter and during food stress		EPA 1993a

^a Suggested values for use in exposure assessment are in bold.

Table C.6. Life history parameters for white-tailed deer (*Odocoileus virginianus*)

Parameter	Value ^a	Comments	Reference
Body Weight	68 kg (♂) 45 kg (♀) 56.5 kg (mean♂ + ♀)		Smith 1991
Food Consumption Rate	1.74 kg/d		Mautz et al. 1976
Water Consumption Rate	3.7 L/d	Estimated using allometric equation ^b assuming 56.5 kg bw	Calder and Braun 1983
Soil Consumption Rate	<2% 0.0348 kg/d	assuming 2% soil and 1.74 kg/d food consumption rates	Beyer et al. 1994
Diet Composition	exclusively herbivorous diet diverse and variable, depends on availability. major foods: - buds and twigs of trees and shrubs - grasses and forbs (summer) - mast and fruits (fall)		Martin et al. 1951 Smith 1991
Home Range	59 - 520 ha		Marchinton and Hirth 1984
Habitat Requirements	uses a wide variety of habitats; favors forest-field-farmland mosaic; population density directly related to number and distribution of forest openings		Smith 1991
Population Density	0.06 /ha 0.39 - 0.78 /ha 0.1704/ha	eastern mixed deciduous forest - Tennessee oak-hickory forest - midwest (calculated based upon 2000 deer on the ORR and available habitat)	Barber 1984 Torgerson and Porath 1984 J. Evans (pers. comm., 1995)
Behavior	generally crepuscular active year-round; does not hibernate		Smith 1991

^a Suggested values for use in exposure assessment are in bold.

^b Allometric equation for estimation of water consumption by mammals is:

$$W = 0.099(\text{bw})^{0.90}$$

where: **W** = water consumption (L/d)
bw = body weight (kg)

Table C.7. Life history parameters for red fox (*Vulpes fulva*)

Parameter	Value ^a	Comments	Reference
Body Weight	5.25 ± 0.18 kg (♂)	Illinois	Storm et al. 1976
	4.13 ± 0.11 kg (♀)		
	4.82 ± 0.081 kg (♂)	Iowa	
	3.94 ± 0.079 kg (♀)		
	4.5 kg	mean ♂+♀ for both Illinois and Iowa	
Food Consumption Rate	0.596 kg/d	see calculation below ^b	Vogtsberger and Barret 1973
	0.31 kg/d	0.069 g/g/d for nonbreeding adult times 4.5 kg bw	Sargent 1978
	0.45 kg/d	mean of both estimates	
Water Consumption Rate	0.38 L/d	Estimated using allometric equation ^c ; assuming 4.5 kg bw	Calder and Braun 1983
Soil Consumption Rate	2.8%		Beyer et al. 1994
	0.0126 kg/d	assuming diet of 0.45 kg/d	
Diet Composition	mammals - 68.8% birds - 12.0% plants - 10.4% insects - 0.9% misc. - 5.5%	Maryland, Appalachian region	Hockman and Chapman 1983
Home Range	699 ± 137 ha (♀ spring)	Minnesota - forest, field, swamp	Sargent 1972
	717 ha (♂ all year)	Wisconsin - multiple habitats	Ables 1969
	96 ha (♀ all year)		
Habitat Requirements	wide and diverse - occur in many habitats		EPA 1993a
	prefer mixture of forest and open habitat		Burt and Grossenheider 1976
Population Density	0.046 - 0.077 /ha	"good fox range" in North America	EPA 1993a
Behavior	active year round - does not hibernate		EPA 1993a

^a Suggested values for use in exposure assessment are in bold.

^b The following parameters were presented by Vogtsberger and Barret (1973):

food ingestion	=	223 kcal/kg bw/d
energy content of vertebrate food	=	5.606 kcal/g dry wt.
wet-dry weight conversion	=	1 g wet wt = 0.3 g dry wt

therefore:

$$223 \text{ kcal/kg bw/d} \times 4.5 \text{ kg bw} = 1003.5 \text{ kcal/d}$$

$$1003.5 \text{ kcal/d} \times 1 \text{ g dry wt./5.606 kcal} = 179 \text{ g dry/d}$$

$$179 \text{ g dry/d} \times 1 \text{ g wet/0.3 g dry (wet-dry conversion)} = 596 \text{ g/d}$$

^c Allometric equation for estimation of water consumption by mammals is:

$$W = 0.099(\text{bw})^{0.90}$$

where: W = water consumption (L/d)
 bw = body weight (kg)

Table C.8. Life History parameters for red-tailed hawks (*Buteo jamaicensis*)

Parameter	Value ^a	Comments	Reference
Body Weight	1.028 kg (♂) 1.224 kg (♀) 1.126 kg (mean♂+♀)		Dunning 1984
Food Consumption Rate	0.109 kg/d		Craighead and Craighead 1969
Water Consumption Rate	0.064 L/d	Estimated using allometric equation ^b ; assuming 1.126 kg bw	Calder and Braun 1983
Soil Consumption Rate	while some soil attached to prey may be ingested, amount is assumed to negligible		
Diet Composition	predominantly small mammals small mammal - 78.5 % bird - 8.5 % snake - 13.0 %	Oregon - pasture and wheat fields	EPA 1993a Janes 1984
Home Range	233 ha 1936 ha (957 - 2465 ha range)	Oregon - pasture and wheat fields Colorado - prairie-pinyon/juniper woodland; mean of 4 birds; 95 % ellipse and systematic relocation	Janes 1984 Anderson and Rongstad 1989
Habitat Requirements	use wide range of habitats. prefer landscapes containing mixture of oldfields, wetlands and pasture for foraging with trees interspersed for perching and nesting		EPA 1993a DeGraaf et al. 1981
Population Density	0.03 - >0.005 pairs/ha		EPA 1993a
Behavior	territorial throughout year northerly populations migrate; those in the south do not		Brown and Amadon 1968 ^b National Geographic Society 1987

^a Suggested values for use in exposure assessment are in bold.

* Allometric equation for estimation of water consumption by birds is:

$$W=0.059(bw)^{0.67}$$

where: W = water consumption (L/d)
bw = body weight (kg)

Table C.9. Life history parameters for mink

Parameter	Value	Comments	Reference
Body Weight	1.0 kg (mean ♂+♀)		EPA 1993b
Food Consumption Rate	0.137 kg/d (mean ♂+♀)		Bleavins and Aulerich 1981
Water Consumption Rate	0.099 L/d	estimated using allometric equation ^a assuming 1.0 kg bw	Calder and Braun 1983
Diet Composition	Diverse diet includes: mammals, fish, aquatic invertebrates, amphibians, and birds Proportion of aquatic prey (fish, amphibians, inverts, etc.) = 0.546±0.21 fish sizes: 0-10 cm=72% 11-20 cm=28%	Proportion represents means of values from five studies	Hamilton 1940, Sealander 1943, Korschgen 1958, Burgess and Bider 1980 Alexander 1977
Home Range	2.63 km (♂) 1.85 km (♀) 770 ha (♂)	stream - Sweden prairie potholes, Manitoba range size and shape depends on habitat - linear along streams, circular in marshes	Gerell 1970 Arnold and Fritzell 1987 EPA 1993a.
Habitat Requirements	aquatic habitats - streams, lakes, marshes;		Burt and Grossenheider 1976
Population Density	0.03 - 0.085 /ha 0.6/km	river - Montana river - Michigan	Mitchell 1961 EPA 1993a
Behavior	nocturnal active year-round, does not hibernate		EPA 1993a

^a Allometric equation for estimation of water consumption by mammals is:

$$W = 0.099(bw)^{0.90}$$

where: W = water consumption (L/d)
bw = body weight (kg)

Table C.10. Life history parameters for belted kingfisher

Parameter	Value	Comments	Reference
Body Weight	0.148 kg		Dunning 1984
Food Consumption Rate	50% bw		Alexander 1977
	0.075 kg/d	assuming 0.148 kg bw	
Water Consumption Rate	0.016 L/d	estimated using allometric equation* assuming 0.148 kg bw	Calder and Braun 1983
Soil Consumption Rate	as a piscivore, assumed to be negligible		
Diet Composition	Cyprinids - 76.4% other fish - 10.2% crayfish - 13.3%	Ohio - creek	Davis 1982
	lizards, small snakes, frogs, salamanders, and insects may be consumed if fish are unavailable		Landrum et al. 1993
Home Range	1.03 km (breeding) 0.39 km (non-breeding)	Ohio - creek	Davis 1982
	2.19 km (breeding)	Pennsylvania - stream summer	Brooks and Davis 1987
Habitat Requirements	uses a diverse aquatic habitats (stream, river, lake, marsh, coastline)		Brooks and Davis 1987
	require high vertical banks composed of >75% sand and <7% clay for nest construction		
	prefer relatively clear waters free of thick vegetation		Bent 1940.
Population Density	0.11 - 0.19 pairs/km shore	Pennsylvania - stream summer	Brooks and Davis 1987
Behavior	while most migrate from northern parts of range, some may stay in areas where water remains ice-free		Bent 1940.

* Allometric equation for estimation of water consumption by birds is:

$$W = 0.059(bw)^{0.67}$$

where: W = water consumption (L/d)
bw = body weight (kg)

Table C.11. Life History Parameters for the Wild Turkey (*Meleagris gallopavo*)

Parameter	Value ^a	Comments	Reference
Body Weight	7.400 kg (♂)		Dunning 1984
	4.222 kg (♀)		
	5.8 kg (mean♂+♀)		
Food Consumption Rate	13.6 g/lb bw/d		Korschgen 1967
	0.174 kg/d	assuming 5.8 kg bw	
Water Consumption Rate	0.19 L/d	estimated using allometric equation ^c assuming 5.8 kg bw	Calder and Braun 1983
Soil Consumption Rate	9.3 %		Beyer et al. 1994
	0.0162 kg/d	assuming 0.174 kg/d food consumption rates	
Diet Composition	plant material (mast, fruit, seeds, some foliage) - 90.3%		Korschgen 1967
	animal material (insects, crayfish, snails, salamanders) - 9.7 %		
Home Range	150 - 190 ha		Pough 1951 ^b
Habitat Requirements	mast-producing woodlands with associated fields and abundant water		Schorger 1966 ^b
Population Density	0.03 /ha	West Virginia	Uhling 1950 ^b
	0.06 - 0.076 /ha	in 'ideal' habitat	Pough 1951 ^b
	0.0426 /ha (calculated based on @ 500 turkey observed on ORR and suitable habitat)	Oak Ridge Reservation	Personal Communication, Jim Evans 1995
Behavior	forage primarily on the ground		National Geographic Society 1987
	roost in trees at night		
	year-round resident; does not migrate		

^a Suggested values for use in exposure assessment are in bold.

^b Cited in DeGraaf et al. 1981.

° Allometric equation for estimation of water consumption for birds is:

$$WIR = 0.059(BW)^{0.67}$$

where:

WIR = water ingestion rate (L water/individual/day).

Table C.12. Point Estimates of Contaminant Exposure for Short-tailed Shrews in WAG2.

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
IHP	Acetone	0.007	0.00	0.00
MWC	Acetone	0.011	0.00	0.00
IHP	Aluminum	2816.097	1226.96	122.70
LMB	Aluminum	3187.367	1388.72	138.87
LWC	Aluminum	1649.656	718.75	71.87
MWC	Aluminum	2302.725	1003.29	100.33
backg	Aluminum	6078.748	2648.48	264.85
LMB	Antimony	0.917	0.40	0.04
MWC	Antimony	0.086	0.58	0.06
backg	Antimony	0.166	1.12	0.11
LMB	Aroclor-1248	0.002	0.05	0.01
IHP	Aroclor-1254	0.897	13.42	1.34
LMB	Aroclor-1254	0.028	0.42	0.04
IHP	Aroclor-1260	2.190	32.76	3.28
LWC	Aroclor-1260	2.098	31.39	3.14
MWC	Aroclor-1260	10.206	152.70	15.27
IHP	Arsenic	1.918	12.80	1.28
LMB	Arsenic	3.194	21.31	2.13
MWC	Arsenic	0.002	0.01	0.00
backg	Arsenic	3.981	26.57	2.66
IHP	Barium	32.507	2.75	0.75
LMB	Barium	53.811	4.55	1.24
LWC	Barium	21.347	1.80	0.49
MWC	Barium	27.967	2.36	0.64
backg	Barium	36.051	3.05	0.83
IHP	Benzo(a)pyrene	0.027	0.02	0.00
LWC	Benzo(a)pyrene	0.012	0.01	0.00
MWC	Benzo(a)pyrene	0.009	0.01	0.00
backg	Beryllium	0.168	0.12	
IHP	Beryllium	0.645	0.44	
LMB	Beryllium	0.558	0.38	
LWC	Beryllium	0.456	0.31	
MWC	Beryllium	0.594	0.41	
IHP	Boron	0.685	0.01	0.00
LMB	Boron	0.869	0.01	0.00
LWC	Boron	0.792	0.01	0.00
MWC	Boron	1.126	0.02	0.01
IHP	Cadmium	9.444	31.59	3.16
LMB	Cadmium	8.043	26.90	2.69
LWC	Cadmium	7.000	23.41	2.34
MWC	Cadmium	7.564	25.30	2.53

Table 12 (Continued)

Location	Contaminant	Total Exposure*	NOAEL HQ	LOAEL HQ
backg	Cadmium	3.200	10.70	1.07
IHP	Chloroform	0.004	0.00	0.00
MWC	Chloroform	0.002	0.00	0.00
backg	Chromium	110.474	15.32	3.83
IHP	Chromium	127.246	17.65	4.41
LMB	Chromium	88.251	12.24	3.06
LWC	Chromium	198.547	27.54	6.88
MWC	Chromium	215.309	29.87	7.46
IHP	Copper	24.044	0.72	0.56
LMB	Copper	18.564	0.56	0.43
LWC	Copper	24.379	0.73	0.56
MWC	Copper	68.843	2.06	1.59
backg	Copper	27.183	0.81	0.63
backg	Cyanide	0.038	0.00	
IHP	Di-n-butylphthalate	0.009	0.00	0.00
LMB	Di-n-butylphthalate	0.020	0.00	0.00
LWC	Di-n-butylphthalate	0.014	0.00	0.00
MWC	Di-n-butylphthalate	0.007	0.00	0.00
IHP	Lead	24.109	1.37	0.14
LMB	Lead	5.833	0.33	0.03
LWC	Lead	15.684	0.89	0.09
MWC	Lead	27.456	1.56	0.16
backg	Lead	13.513	0.77	0.08
IHP	Lithium	4.986	0.24	0.12
LMB	Lithium	3.945	0.19	0.10
LWC	Lithium	2.463	0.12	0.06
MWC	Lithium	3.530	0.17	0.09
backg	Manganese	0.462	0.00	0.00
IHP	Manganese	241.392	1.25	0.39
LMB	Manganese	1021.870	5.28	1.64
LWC	Manganese	170.370	0.88	0.27
MWC	Manganese	141.617	0.73	0.23
IHP	Mercury	85.769	1219.52	243.90
LMB	Mercury	3.296	46.86	9.37
LWC	Mercury	13.084	186.03	37.21
MWC	Mercury	18.647	265.13	53.03
backg	Mercury	1.471	20.91	4.18
LMB	Methylene chloride	0.002	0.00	0.00
MWC	Methylene chloride	0.000	0.00	0.00
IHP	Molybdenum	6.425	20.78	2.08

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure*	HO	HO
LMB	Molybdenum	10.608	34.31	3.43
LWC	Molybdenum	5.352	17.31	1.73
MWC	Molybdenum	6.818	22.05	2.21
IHP	Nickel	266.737	3.03	1.52
LMB	Nickel	100.067	1.14	0.57
LWC	Nickel	49.278	0.56	0.28
MWC	Nickel	119.102	1.35	0.68
backg	Nickel	136.708	1.56	0.78
IHP	Selenium	1.355	14.99	1.50
LMB	Selenium	2.505	27.72	2.77
LWC	Selenium	2.965	32.81	3.28
MWC	Selenium	7.994	88.45	8.85
backg	Selenium	2.924	32.35	3.24
IHP	Strontium	6.628	0.01	
LMB	Strontium	5.660	0.01	
LWC	Strontium	3.257	0.01	
MWC	Strontium	7.717	0.01	
IHP	Thallium	0.067	4.08	0.41
LMB	Thallium	0.113	6.88	0.69
LWC	Thallium	0.063	3.84	0.38
MWC	Thallium	0.082	4.98	0.50
backg	Thallium	0.068	4.13	0.41
IHP	Tin	0.869	0.03	0.02
LMB	Tin	0.846	0.03	0.02
LWC	Tin	0.566	0.02	0.01
MWC	Tin	0.725	0.03	0.02
IHP	Vanadium	3.551	8.29	0.83
LMB	Vanadium	4.729	11.04	1.10
LWC	Vanadium	2.402	5.61	0.56
MWC	Vanadium	3.431	8.01	0.80
backg	Vanadium	9.032	21.08	2.11
IHP	Zinc	242.420	0.69	0.34
LMB	Zinc	165.616	0.47	0.24
LWC	Zinc	380.413	1.08	0.54
MWC	Zinc	709.220	2.02	1.01
backg	Zinc	257.140	0.73	0.37
LMB	bis(2-Ethylhexyl)phthalate	0.020	0.00	0.00
LWC	bis(2-Ethylhexyl)phthalate	0.008	0.00	0.00
MWC	bis(2-Ethylhexyl)phthalate	0.016	0.00	0.00

* Biota estimates = UCL soil x UCL uptake factor.

Table C.13. Point Estimates of Contaminant Exposure for White-footed Mice in WAG 2.

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
IHP	Acetone	0.001	0.00	0.00
MWC	Acetone	0.002	0.00	0.00
IHP	Aluminum	249.575	119.67	11.97
LMB	Aluminum	282.459	135.43	13.54
LWC	Aluminum	146.194	70.10	7.01
MWC	Aluminum	204.066	97.84	9.78
backg	Aluminum	538.693	258.29	25.83
LMB	Antimony	0.040	0.02	0.00
MWC	Antimony	0.012	0.09	0.01
backg	Antimony	0.019	0.14	0.01
LMB	Aroclor-1248	0.000	0.00	0.00
IHP	Aroclor-1254	0.104	1.71	0.17
LMB	Aroclor-1254	0.003	0.05	0.01
IHP	Aroclor-1260	0.279	4.59	0.46
LWC	Aroclor-1260	0.267	4.40	0.44
MWC	Aroclor-1260	1.299	21.40	2.14
IHP	Arsenic	0.212	1.56	0.16
LMB	Arsenic	0.353	2.59	0.26
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.440	3.23	0.32
IHP	Barium	4.791	0.45	0.12
LMB	Barium	7.930	0.74	0.20
LWC	Barium	3.145	0.29	0.08
MWC	Barium	4.121	0.38	0.10
backg	Barium	5.312	0.49	0.13
IHP	Benzo(a)pyrene	0.001	0.00	0.00
LWC	Benzo(a)pyrene	0.000	0.00	0.00
MWC	Benzo(a)pyrene	0.000	0.00	0.00
backg	Beryllium	0.007	0.01	
IHP	Beryllium	0.075	0.06	
LMB	Beryllium	0.065	0.05	
LWC	Beryllium	0.053	0.04	
MWC	Beryllium	0.069	0.05	
IHP	Boron	0.037	0.00	0.00
LMB	Boron	0.046	0.00	0.00
LWC	Boron	0.031	0.00	0.00
MWC	Boron	0.053	0.00	0.00
IHP	Cadmium	1.247	4.58	0.46
LMB	Cadmium	1.061	3.90	0.39
LWC	Cadmium	0.924	3.40	0.34
MWC	Cadmium	0.999	3.67	0.37

Table12 (Continued)

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
backg	Cadmium	0.422	1.55	0.16
IHP	Chloroform	0.001	0.00	0.00
MWC	Chloroform	0.000	0.00	0.00
backg	Chromium	14.099	2.15	0.54
IHP	Chromium	16.240	2.48	0.62
LMB	Chromium	11.264	1.72	0.43
LWC	Chromium	25.340	3.87	0.97
MWC	Chromium	27.480	4.19	1.05
IHP	Copper	3.360	0.11	0.09
LMB	Copper	2.594	0.09	0.07
LWC	Copper	3.407	0.11	0.09
MWC	Copper	9.621	0.32	0.24
backg	Copper	3.799	0.13	0.10
backg	Cyanide	0.002	0.00	
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.001	0.00	0.00
LWC	Di-n-butylphthalate	0.001	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	2.712	0.17	0.02
LMB	Lead	0.656	0.04	0.00
LWC	Lead	1.764	0.11	0.01
MWC	Lead	3.089	0.19	0.02
backg	Lead	1.521	0.10	0.01
IHP	Lithium	0.469	0.02	0.01
LMB	Lithium	0.371	0.02	0.01
LWC	Lithium	0.232	0.01	0.01
MWC	Lithium	0.332	0.02	0.01
backg	Manganese	0.063	0.00	0.00
IHP	Manganese	23.641	0.13	0.04
LMB	Manganese	100.080	0.57	0.18
LWC	Manganese	16.688	0.09	0.03
MWC	Manganese	13.871	0.08	0.02
IHP	Mercury	10.927	170.97	34.20
LMB	Mercury	0.420	6.57	1.31
LWC	Mercury	1.667	26.08	5.22
MWC	Mercury	2.375	37.17	7.43
backg	Mercury	0.188	2.94	0.59
LMB	Methylene chloride	0.000	0.00	0.00
MWC	Methylene chloride	0.000	0.00	0.00
IHP	Molybdenum	0.796	2.83	0.28

Table12 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
LMB	Molybdenum	1.314	4.68	0.47
LWC	Molybdenum	0.663	2.36	0.24
MWC	Molybdenum	0.845	3.01	0.30
IHP	Nickel	33.937	0.42	0.21
LMB	Nickel	12.732	0.16	0.08
LWC	Nickel	6.270	0.08	0.04
MWC	Nickel	15.153	0.19	0.09
backg	Nickel	17.395	0.22	0.11
IHP	Selenium	0.171	2.08	0.21
LMB	Selenium	0.316	3.85	0.39
LWC	Selenium	0.374	4.56	0.46
MWC	Selenium	1.009	12.28	1.23
backg	Selenium	0.369	4.49	0.45
IHP	Strontium	0.698	0.00	
LMB	Strontium	0.601	0.00	
LWC	Strontium	0.354	0.00	
MWC	Strontium	0.811	0.00	
IHP	Thallium	0.003	0.18	0.02
LMB	Thallium	0.004	0.30	0.03
LWC	Thallium	0.003	0.17	0.02
MWC	Thallium	0.003	0.22	0.02
backg	Thallium	0.003	0.18	0.02
IHP	Tin	0.034	0.00	0.00
LMB	Tin	0.034	0.00	0.00
LWC	Tin	0.022	0.00	0.00
MWC	Tin	0.029	0.00	0.00
IHP	Vanadium	0.366	0.94	0.09
LMB	Vanadium	0.487	1.25	0.12
LWC	Vanadium	0.247	0.63	0.06
MWC	Vanadium	0.353	0.91	0.09
backg	Vanadium	0.931	2.39	0.24
IHP	Zinc	34.870	0.11	0.05
LMB	Zinc	23.823	0.07	0.04
LWC	Zinc	54.719	0.17	0.09
MWC	Zinc	102.015	0.32	0.16
backg	Zinc	36.984	0.12	0.06
LMB	bis(2-Ethylhexyl)phthalate	0.001	0.00	0.00
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
MWC	bis(2-Ethylhexyl)phthalate	0.001	0.00	0.00

^a Biota estimates = UCL soil x UCL uptake factor.

Table C.14. Point Estimates of Contaminant Exposure for White-tailed Deer in WAG 2.

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
IHP	Acetone	0.000	0.00	0.00
MWC	Acetone	0.000	0.00	0.00
IHP	Aluminum	24.289	82.91	8.29
LMB	Aluminum	27.482	93.81	9.38
LWC	Aluminum	14.226	48.56	4.86
MWC	Aluminum	19.856	67.77	6.78
backg	Aluminum	52.415	178.91	17.89
LMB	Antimony	0.008	0.03	0.00
MWC	Antimony	0.003	0.14	0.01
backg	Antimony	0.004	0.22	0.02
LMB	Aroclor-1248	0.000	0.00	0.00
IHP	Aroclor-1254	0.001	0.12	0.01
LMB	Aroclor-1254	0.000	0.00	0.00
IHP	Aroclor-1260	0.000	0.03	0.00
LWC	Aroclor-1260	0.000	0.03	0.00
MWC	Aroclor-1260	0.001	0.16	0.02
IHP	Arsenic	0.008	0.40	0.04
LMB	Arsenic	0.013	0.67	0.07
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.016	0.84	0.08
IHP	Barium	1.234	0.82	0.22
LMB	Barium	2.042	1.35	0.37
LWC	Barium	0.809	0.54	0.15
MWC	Barium	1.061	0.70	0.19
backg	Barium	1.367	0.91	0.25
IHP	Benzo(a)pyrene	0.000	0.00	0.00
LWC	Benzo(a)pyrene	0.000	0.00	0.00
MWC	Benzo(a)pyrene	0.000	0.00	0.00
backg	Beryllium	0.001	0.01	
IHP	Beryllium	0.002	0.01	
LMB	Beryllium	0.001	0.01	
LWC	Beryllium	0.001	0.01	
MWC	Beryllium	0.002	0.01	
IHP	Boron	0.008	0.00	0.00
LMB	Boron	0.009	0.00	0.00
LWC	Boron	0.006	0.00	0.00
MWC	Boron	0.011	0.00	0.00
IHP	Cadmium	0.045	1.18	0.12
LMB	Cadmium	0.038	1.00	0.10
LWC	Cadmium	0.034	0.88	0.09
MWC	Cadmium	0.036	0.95	0.09
backg	Cadmium	0.016	0.42	0.04

Table14 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
IHP	Chloroform	0.000	0.00	0.00
MWC	Chloroform	0.000	0.00	0.00
backg	Chromium	0.108	0.12	0.03
IHP	Chromium	0.124	0.13	0.03
LMB	Chromium	0.086	0.09	0.02
LWC	Chromium	0.193	0.21	0.05
MWC	Chromium	0.209	0.23	0.06
IHP	Copper	0.293	0.07	0.05
LMB	Copper	0.226	0.05	0.04
LWC	Copper	0.297	0.07	0.05
MWC	Copper	0.838	0.20	0.15
backg	Copper	0.332	0.08	0.06
backg	Cyanide	0.000	0.00	
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.000	0.00	0.00
LWC	Di-n-butylphthalate	0.000	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	0.127	0.06	0.01
LMB	Lead	0.031	0.01	0.00
LWC	Lead	0.082	0.04	0.00
MWC	Lead	0.144	0.06	0.01
backg	Lead	0.072	0.03	0.00
IHP	Lithium	0.041	0.02	0.01
LMB	Lithium	0.032	0.01	0.01
LWC	Lithium	0.020	0.01	0.00
MWC	Lithium	0.029	0.01	0.01
backg	Manganese	0.014	0.00	0.00
IHP	Manganese	12.328	0.50	0.15
LMB	Manganese	52.187	2.11	0.65
LWC	Manganese	8.699	0.35	0.11
MWC	Manganese	7.232	0.29	0.09
IHP	Mercury	0.082	9.11	1.82
LMB	Mercury	0.003	0.35	0.07
LWC	Mercury	0.013	1.39	0.28
MWC	Mercury	0.018	1.98	0.40
backg	Mercury	0.002	0.18	0.04
LMB	Methylene chloride	0.000	0.00	0.00
MWC	Methylene chloride	0.000	0.00	0.00
IHP	Molybdenum	0.003	0.07	0.01
LMB	Molybdenum	0.005	0.12	0.01

Table 14 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL	LOAEL
			HO	HO
LWC	Molybdenum	0.002	0.06	0.01
MWC	Molybdenum	0.003	0.07	0.01
IHP	Nickel	0.362	0.03	0.02
LMB	Nickel	0.136	0.01	0.01
LWC	Nickel	0.067	0.01	0.00
MWC	Nickel	0.162	0.01	0.01
backg	Nickel	0.188	0.02	0.01
IHP	Selenium	0.008	0.69	0.07
LMB	Selenium	0.015	1.28	0.13
LWC	Selenium	0.017	1.51	0.15
MWC	Selenium	0.047	4.06	0.41
backg	Selenium	0.017	1.51	0.15
IHP	Strontium	0.021	0.00	
LMB	Strontium	0.022	0.00	
LWC	Strontium	0.020	0.00	
MWC	Strontium	0.023	0.00	
IHP	Thallium	0.001	0.25	0.03
LMB	Thallium	0.001	0.43	0.04
LWC	Thallium	0.000	0.24	0.02
MWC	Thallium	0.001	0.31	0.03
backg	Thallium	0.001	0.26	0.03
IHP	Tin	0.007	0.00	0.00
LMB	Tin	0.007	0.00	0.00
LWC	Tin	0.004	0.00	0.00
MWC	Tin	0.006	0.00	0.00
IHP	Vanadium	0.034	0.62	0.06
LMB	Vanadium	0.045	0.83	0.08
LWC	Vanadium	0.023	0.42	0.04
MWC	Vanadium	0.033	0.60	0.06
backg	Vanadium	0.087	1.59	0.16
IHP	Zinc	1.636	0.04	0.02
LMB	Zinc	1.117	0.02	0.01
LWC	Zinc	2.564	0.06	0.03
MWC	Zinc	4.781	0.11	0.05
backg	Zinc	1.744	0.04	0.02
LMB	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
MWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00

^a: Biota estimates=UCL soil x UCL uptake factor.

Table C.15. Point Estimates of Contaminant Exposure for Red Fox in WAG 2.

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure*	HO	HO
IHP	Acetone	0.000	0.00	0.00
MWC	Acetone	0.000	0.00	0.00
IHP	Aluminum	88.024	159.61	15.96
LMB	Aluminum	99.626	180.65	18.07
LWC	Aluminum	51.563	93.50	9.35
MWC	Aluminum	71.975	130.51	13.05
backg	Aluminum	190.001	344.53	34.45
LMB	Antimony	0.033	0.06	0.01
MWC	Antimony	0.003	0.09	0.01
backg	Antimony	0.006	0.18	0.02
LMB	Aroclor-1248	0.000	0.01	0.00
IHP	Aroclor-1254	0.016	0.17	0.03
LMB	Aroclor-1254	0.000	0.01	0.00
IHP	Aroclor-1260	0.151	1.57	0.32
LWC	Aroclor-1260	0.145	1.51	0.31
MWC	Aroclor-1260	0.705	7.33	1.49
IHP	Arsenic	0.043	1.18	0.12
LMB	Arsenic	0.071	1.97	0.20
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.088	2.46	0.25
IHP	Barium	1.923	0.68	0.18
LMB	Barium	3.184	1.12	0.30
LWC	Barium	1.261	0.44	0.12
MWC	Barium	1.654	0.58	0.16
backg	Barium	2.131	0.75	0.20
IHP	Benzo(a)pyrene	0.001	0.00	0.00
LWC	Benzo(a)pyrene	0.000	0.00	0.00
MWC	Benzo(a)pyrene	0.000	0.00	0.00
backg	Beryllium	0.006	0.02	
IHP	Beryllium	0.012	0.03	
LMB	Beryllium	0.010	0.03	
LWC	Beryllium	0.008	0.02	
MWC	Beryllium	0.011	0.03	
IHP	Boron	0.025	0.00	0.00
LMB	Boron	0.031	0.00	0.00
LWC	Boron	0.028	0.00	0.00
MWC	Boron	0.041	0.00	0.00
IHP	Cadmium	0.194	2.70	0.27
LMB	Cadmium	0.166	2.30	0.23
LWC	Cadmium	0.144	2.01	0.20
MWC	Cadmium	0.156	2.17	0.22

Table 15 (Continued)

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
backg	Cadmium	0.066	0.92	0.09
IHP	Chloroform	0.000	0.00	0.00
MWC	Chloroform	0.000	0.00	0.00
backg	Chromium	2.088	1.21	0.30
IHP	Chromium	2.405	1.39	0.35
LMB	Chromium	1.668	0.96	0.24
LWC	Chromium	3.752	2.17	0.54
MWC	Chromium	4.069	2.35	0.59
IHP	Copper	1.817	0.23	0.17
LMB	Copper	1.403	0.17	0.13
LWC	Copper	1.842	0.23	0.18
MWC	Copper	5.204	0.65	0.50
backg	Copper	2.053	0.26	0.20
backg	Cyanide	0.001	0.00	
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.001	0.00	0.00
LWC	Di-n-butylphthalate	0.001	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	0.663	0.16	0.02
LMB	Lead	0.160	0.04	0.00
LWC	Lead	0.431	0.10	0.01
MWC	Lead	0.755	0.18	0.02
backg	Lead	0.372	0.09	0.01
IHP	Lithium	0.166	0.03	0.02
LMB	Lithium	0.131	0.03	0.01
LWC	Lithium	0.082	0.02	0.01
MWC	Lithium	0.117	0.02	0.01
backg	Manganese	0.018	0.00	0.00
IHP	Manganese	11.836	0.25	0.08
LMB	Manganese	50.104	1.08	0.33
LWC	Manganese	8.353	0.18	0.06
MWC	Manganese	6.943	0.15	0.05
IHP	Mercury	2.926	284.05	170.50
LMB	Mercury	0.112	10.92	6.55
LWC	Mercury	0.446	43.33	26.01
MWC	Mercury	0.636	61.75	37.06
backg	Mercury	0.050	4.87	2.92
LMB	Methylene chloride	0.000	0.00	0.00
MWC	Methylene chloride	0.000	0.00	0.00
IHP	Molybdenum	0.102	1.37	0.14

Table 15 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
LMB	Molybdenum	0.168	2.26	0.23
LWC	Molybdenum	0.085	1.14	0.11
MWC	Molybdenum	0.108	1.45	0.15
IHP	Nickel	5.220	0.25	0.12
LMB	Nickel	1.959	0.09	0.05
LWC	Nickel	0.965	0.05	0.02
MWC	Nickel	2.331	0.11	0.06
backg	Nickel	2.677	0.13	0.06
IHP	Selenium	0.038	1.76	0.18
LMB	Selenium	0.071	3.25	0.33
LWC	Selenium	0.084	3.86	0.39
MWC	Selenium	0.226	10.39	1.04
backg	Selenium	0.083	3.81	0.38
IHP	Strontium	0.140	0.00	
LMB	Strontium	0.122	0.00	
LWC	Strontium	0.075	0.00	
MWC	Strontium	0.162	0.00	
IHP	Thallium	0.002	0.61	0.06
LMB	Thallium	0.004	1.03	0.10
LWC	Thallium	0.002	0.57	0.06
MWC	Thallium	0.003	0.74	0.07
backg	Thallium	0.002	0.62	0.06
IHP	Tin	0.031	0.00	0.00
LMB	Tin	0.030	0.00	0.00
LWC	Tin	0.020	0.00	0.00
MWC	Tin	0.026	0.00	0.00
IHP	Vanadium	0.102	0.99	0.10
LMB	Vanadium	0.136	1.32	0.13
LWC	Vanadium	0.069	0.67	0.07
MWC	Vanadium	0.098	0.96	0.10
backg	Vanadium	0.259	2.52	0.25
IHP	Zinc	15.871	0.19	0.09
LMB	Zinc	10.844	0.13	0.06
LWC	Zinc	24.910	0.29	0.15
MWC	Zinc	46.438	0.55	0.27
backg	Zinc	16.825	0.20	0.10
LMB	bis(2-Ethylhexyl)phthalate	0.001	0.00	0.00
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
MWC	bis(2-Ethylhexyl)phthalate	0.001	0.00	0.00

^a Biota estimates=UCL soil x UCL uptake factor.

Table C.16. Point Estimates of Contaminant Exposure Hawk.

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
IHP	Acetone	0.000		
MWC	Acetone	0.000		
IHP	Aluminum	8.089	0.07	0.18
LMB	Aluminum	9.145	0.08	0.21
LWC	Aluminum	4.736	0.04	0.11
MWC	Aluminum	6.608	0.06	0.15
backg	Aluminum	17.444	0.16	0.39
LMB	Antimony	0.001	0.00	0.00
MWC	Antimony	0.002		
backg	Antimony	0.003		
LMB	Aroclor-1248	0.000		
IHP	Aroclor-1254	0.000	0.00	0.00
LMB	Aroclor-1254	0.000	0.00	0.00
IHP	Aroclor-1260	0.142	0.79	0.08
LWC	Aroclor-1260	0.136	0.75	0.08
MWC	Aroclor-1260	0.660	3.67	0.37
IHP	Arsenic	0.004	0.00	0.00
LMB	Arsenic	0.006	0.00	0.00
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.008	0.00	0.00
IHP	Barium	0.947	0.05	0.02
LMB	Barium	1.568	0.08	0.04
LWC	Barium	0.622	0.03	0.01
MWC	Barium	0.815	0.04	0.02
backg	Barium	1.050	0.05	0.03
IHP	Benzo(a)pyrene	0.000		
LWC	Benzo(a)pyrene	0.000		
MWC	Benzo(a)pyrene	0.000		
backg	Beryllium	0.000		
IHP	Beryllium	0.000		
LMB	Beryllium	0.000		
LWC	Beryllium	0.000		
MWC	Beryllium	0.000		
IHP	Boron	0.003	0.00	0.00
LMB	Boron	0.003	0.00	0.00
LWC	Boron	0.000	0.00	0.00
MWC	Boron	0.002	0.00	0.00
IHP	Cadmium	0.021	0.01	0.00
LMB	Cadmium	0.018	0.01	0.00
LWC	Cadmium	0.016	0.01	0.00
MWC	Cadmium	0.017	0.01	0.00

Table16 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
backg	Cadmium	0.008	0.01	0.00
IHP	Chloroform	0.000		
MWC	Chloroform	0.000		
backg	Chromium	0.438	0.44	0.09
IHP	Chromium	0.504	0.50	0.10
LMB	Chromium	0.350	0.35	0.07
LWC	Chromium	0.786	0.79	0.16
MWC	Chromium	0.853	0.85	0.17
IHP	Copper	1.501	0.03	0.02
LMB	Copper	1.159	0.02	0.02
LWC	Copper	1.522	0.03	0.02
MWC	Copper	4.301	0.09	0.07
backg	Copper	1.696	0.04	0.03
backg	Cyanide	0.000		
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.000	0.00	0.00
LWC	Di-n-butylphthalate	0.000	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	0.204	0.18	0.02
LMB	Lead	0.049	0.04	0.00
LWC	Lead	0.132	0.12	0.01
MWC	Lead	0.232	0.20	0.02
backg	Lead	0.115	0.10	0.01
IHP	Lithium	0.045		
LMB	Lithium	0.035		
LWC	Lithium	0.022		
MWC	Lithium	0.032		
backg	Manganese	0.012	0.00	
IHP	Manganese	0.903	0.00	
LMB	Manganese	3.822	0.00	
LWC	Manganese	0.639	0.00	
MWC	Manganese	0.530	0.00	
IHP	Mercury	1.904	317.34	29.75
LMB	Mercury	0.073	12.20	1.14
LWC	Mercury	0.290	48.40	4.54
MWC	Mercury	0.414	68.99	6.47
backg	Mercury	0.033	5.44	0.51
LMB	Methylene chloride	0.000		
MWC	Methylene chloride	0.000		
IHP	Molybdenum	0.000	0.00	0.00

Table16 (Continued)

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
LMB	Molybdenum	0.000	0.00	0.00
LWC	Molybdenum	0.000	0.00	0.00
MWC	Molybdenum	0.000	0.00	0.00
IHP	Nickel	1.222	0.02	0.01
LMB	Nickel	0.459	0.01	0.00
LWC	Nickel	0.226	0.00	0.00
MWC	Nickel	0.546	0.01	0.01
backg	Nickel	0.628	0.01	0.01
IHP	Selenium	0.012	0.02	0.01
LMB	Selenium	0.022	0.04	0.02
LWC	Selenium	0.026	0.05	0.03
MWC	Selenium	0.070	0.14	0.07
backg	Selenium	0.026	0.05	0.03
IHP	Strontium	0.006		
LMB	Strontium	0.009		
LWC	Strontium	0.012		
MWC	Strontium	0.006		
IHP	Thallium	0.000		
LMB	Thallium	0.000		
LWC	Thallium	0.000		
MWC	Thallium	0.000		
backg	Thallium	0.000		
IHP	Tin	0.000	0.00	0.00
LMB	Tin	0.000	0.00	0.00
LWC	Tin	0.000	0.00	0.00
MWC	Tin	0.000	0.00	0.00
IHP	Vanadium	0.000	0.00	
LMB	Vanadium	0.000	0.00	
LWC	Vanadium	0.000	0.00	
MWC	Vanadium	0.000	0.00	
backg	Vanadium	0.001	0.00	
IHP	Zinc	13.754	0.95	0.10
LMB	Zinc	9.397	0.65	0.07
LWC	Zinc	21.587	1.49	0.16
MWC	Zinc	40.244	2.78	0.31
backg	Zinc	14.577	1.01	0.11
LMB	bis(2-Ethylhexyl)phthalate	0.000	0.00	
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	
MWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	

^a Biota estimates=UCL soil x UCL uptake factor.

Table C.17. Point Estimates of Contaminant Exposure for Wild Turkey in WAG 2.

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
IHP	Acetone	0.000		
MWC	Acetone	0.000		
IHP	Aluminum	75.241	0.69	1.69
LMB	Aluminum	85.165	0.78	1.91
LWC	Aluminum	44.077	0.40	0.99
MWC	Aluminum	61.527	0.56	1.38
backg	Aluminum	162.420	1.48	3.65
LMB	Antimony	0.032	0.00	0.00
MWC	Antimony	0.001		
backg	Antimony	0.003		
LMB	Aroclor-1248	0.000		
IHP	Aroclor-1254	0.008	0.05	0.00
LMB	Aroclor-1254	0.000	0.00	0.00
IHP	Aroclor-1260	0.012	0.07	0.01
LWC	Aroclor-1260	0.011	0.06	0.01
MWC	Aroclor-1260	0.055	0.30	0.03
IHP	Arsenic	0.029	0.01	0.00
LMB	Arsenic	0.049	0.01	0.00
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.061	0.01	0.00
IHP	Barium	2.096	0.10	0.05
LMB	Barium	3.470	0.17	0.08
LWC	Barium	1.371	0.07	0.03
MWC	Barium	1.803	0.09	0.04
backg	Barium	2.320	0.11	0.06
IHP	Benzo(a)pyrene	0.001		
LWC	Benzo(a)pyrene	0.000		
MWC	Benzo(a)pyrene	0.000		
backg	Beryllium	0.006		
IHP	Beryllium	0.008		
LMB	Beryllium	0.007		
LWC	Beryllium	0.006		
MWC	Beryllium	0.007		
IHP	Boron	0.022	0.00	0.00
LMB	Boron	0.029	0.00	0.00
LWC	Boron	0.028	0.00	0.00
MWC	Boron	0.038	0.00	0.00
IHP	Cadmium	0.060	0.04	0.00
LMB	Cadmium	0.051	0.03	0.00
LWC	Cadmium	0.044	0.03	0.00
MWC	Cadmium	0.048	0.03	0.00

Table 17 (Continued)

Location	Contaminant	Total Exposure*	NOAEL HO	LOAEL HO
backg	Cadmium	0.020	0.01	0.00
IHP	Chloroform	0.000		
MWC	Chloroform	0.000		
backg	Chromium	0.735	0.73	0.15
IHP	Chromium	0.847	0.85	0.17
LMB	Chromium	0.587	0.59	0.12
LWC	Chromium	1.321	1.32	0.26
MWC	Chromium	1.433	1.43	0.29
IHP	Copper	0.344	0.01	0.01
LMB	Copper	0.265	0.01	0.00
LWC	Copper	0.349	0.01	0.01
MWC	Copper	0.985	0.02	0.02
backg	Copper	0.389	0.01	0.01
backg	Cyanide	0.001		
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.001	0.01	0.00
LWC	Di-n-butylphthalate	0.001	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	0.438	0.39	0.04
LMB	Lead	0.106	0.09	0.01
LWC	Lead	0.285	0.25	0.03
MWC	Lead	0.499	0.44	0.04
backg	Lead	0.246	0.22	0.02
IHP	Lithium	0.133		
LMB	Lithium	0.105		
LWC	Lithium	0.066		
MWC	Lithium	0.094		
backg	Manganese	0.007	0.00	
IHP	Manganese	22.130	0.02	
LMB	Manganese	93.679	0.10	
LWC	Manganese	15.613	0.02	
MWC	Manganese	12.981	0.01	
IHP	Mercury	0.563	93.82	8.80
LMB	Mercury	0.022	3.61	0.34
LWC	Mercury	0.086	14.33	1.34
MWC	Mercury	0.122	20.40	1.91
backg	Mercury	0.010	1.62	0.15
LMB	Methylene chloride	0.000		
MWC	Methylene chloride	0.000		
IHP	Molybdenum	0.042	0.01	0.00

Table 17 (Continued)

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
LMB	Molybdenum	0.069	0.02	0.00
LWC	Molybdenum	0.035	0.01	0.00
MWC	Molybdenum	0.045	0.01	0.00
IHP	Nickel	1.953	0.03	0.02
LMB	Nickel	0.733	0.01	0.01
LWC	Nickel	0.361	0.00	0.00
MWC	Nickel	0.872	0.01	0.01
backg	Nickel	1.001	0.01	0.01
IHP	Selenium	0.009	0.02	0.01
LMB	Selenium	0.016	0.03	0.02
LWC	Selenium	0.019	0.04	0.02
MWC	Selenium	0.051	0.10	0.05
backg	Selenium	0.019	0.04	0.02
IHP	Strontium	0.089		
LMB	Strontium	0.076		
LWC	Strontium	0.044		
MWC	Strontium	0.104		
IHP	Thallium	0.002		
LMB	Thallium	0.004		
LWC	Thallium	0.002		
MWC	Thallium	0.003		
backg	Thallium	0.002		
IHP	Tin	0.031	0.00	0.00
LMB	Tin	0.030	0.00	0.00
LWC	Tin	0.020	0.00	0.00
MWC	Tin	0.026	0.00	0.00
IHP	Vanadium	0.090	0.01	
LMB	Vanadium	0.119	0.01	
LWC	Vanadium	0.061	0.01	
MWC	Vanadium	0.087	0.01	
backg	Vanadium	0.228	0.02	
IHP	Zinc	2.375	0.16	0.02
LMB	Zinc	1.622	0.11	0.01
LWC	Zinc	3.726	0.26	0.03
MWC	Zinc	6.946	0.48	0.05
backg	Zinc	2.521	0.17	0.02
LMB	bis(2-Ethylhexyl)phthalate	0.001	0.00	
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	
MWC	bis(2-Ethylhexyl)phthalate	0.001	0.00	

^a Biota estimates = UCL soil x UCL uptake factor.

Table C.18. Point Estimates of Contaminant Exposure of Mink to Small Mammals in WAG2.

Location	Contaminant	Total Exposure ^a	NOAEL HO	LOAEL HO
IHP	Acetone	0.000	0.00	0.00
MWC	Acetone	0.000	0.00	0.00
IHP	Aluminum	5.216	6.49	0.65
LMB	Aluminum	5.885	7.33	0.73
LWC	Aluminum	3.050	3.80	0.38
MWC	Aluminum	4.254	5.30	0.53
backg	Aluminum	11.229	13.98	1.40
LMB	Antimony	0.002	0.00	0.00
MWC	Antimony	0.004	0.07	0.01
backg	Antimony	0.006	0.11	0.01
LMB	Aroclor-1248	0.000	0.00	0.00
IHP	Aroclor-1254	0.000	0.00	0.00
LMB	Aroclor-1254	0.000	0.00	0.00
IHP	Aroclor-1260	0.091	0.65	0.13
LWC	Aroclor-1260	0.087	0.62	0.13
MWC	Aroclor-1260	0.424	3.03	0.62
IHP	Arsenic	0.002	0.05	0.00
LMB	Arsenic	0.004	0.08	0.01
MWC	Arsenic	0.000	0.00	0.00
backg	Arsenic	0.005	0.10	0.01
IHP	Barium	0.611	0.15	0.04
LMB	Barium	1.011	0.24	0.07
LWC	Barium	0.404	0.10	0.03
MWC	Barium	0.526	0.13	0.03
backg	Barium	0.680	0.16	0.04
IHP	Benzo(a)pyrene	0.000	0.00	0.00
LWC	Benzo(a)pyrene	0.000	0.00	0.00
MWC	Benzo(a)pyrene	0.000	0.00	0.00
backg	Beryllium	0.000	0.00	
IHP	Beryllium	0.000	0.00	
LMB	Beryllium	0.000	0.00	
LWC	Beryllium	0.000	0.00	
MWC	Beryllium	0.000	0.00	
IHP	Boron	0.005	0.00	0.00
LMB	Boron	0.005	0.00	0.00
LWC	Boron	0.000	0.00	0.00
MWC	Boron	0.004	0.00	0.00
IHP	Cadmium	0.014	0.13	0.01
LMB	Cadmium	0.012	0.11	0.01
LWC	Cadmium	0.010	0.10	0.01
MWC	Cadmium	0.011	0.10	0.01

Table 18 (Continued)

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
backg	Cadmium	0.006	0.05	0.01
IHP	Chloroform	0.000	0.00	0.00
MWC	Chloroform	0.000	0.00	0.00
backg	Chromium	0.282	0.11	0.03
IHP	Chromium	0.324	0.13	0.03
LMB	Chromium	0.225	0.09	0.02
LWC	Chromium	0.505	0.20	0.05
MWC	Chromium	0.548	0.22	0.05
IHP	Copper	0.965	0.08	0.06
LMB	Copper	0.745	0.06	0.05
LWC	Copper	0.979	0.08	0.06
MWC	Copper	2.764	0.24	0.18
backg	Copper	1.091	0.09	0.07
backg	Cyanide	0.000	0.00	
IHP	Di-n-butylphthalate	0.000	0.00	0.00
LMB	Di-n-butylphthalate	0.000	0.00	0.00
LWC	Di-n-butylphthalate	0.000	0.00	0.00
MWC	Di-n-butylphthalate	0.000	0.00	0.00
IHP	Lead	0.131	0.02	0.00
LMB	Lead	0.032	0.01	0.00
LWC	Lead	0.085	0.01	0.00
MWC	Lead	0.149	0.02	0.00
backg	Lead	0.074	0.01	0.00
IHP	Lithium	0.029	0.00	0.00
LMB	Lithium	0.023	0.00	0.00
LWC	Lithium	0.014	0.00	0.00
MWC	Lithium	0.020	0.00	0.00
backg	Manganese	0.021	0.00	0.00
IHP	Manganese	0.582	0.01	0.00
LMB	Manganese	2.465	0.04	0.01
LWC	Manganese	0.414	0.01	0.00
MWC	Manganese	0.343	0.01	0.00
IHP	Mercury	1.223	81.56	48.93
LMB	Mercury	0.047	3.13	1.88
LWC	Mercury	0.187	12.44	7.46
MWC	Mercury	0.266	17.73	10.64
backg	Mercury	0.021	1.42	0.85
LMB	Methylene chloride	0.000	0.00	0.00
MWC	Methylene chloride	0.000	0.00	0.00
IHP	Molybdenum	0.000	0.00	0.00

Table18 (Continued)

Location	Contaminant	Total	NOAEL	LOAEL
		Exposure ^a	HO	HO
LMB	Molybdenum	0.000	0.00	0.00
LWC	Molybdenum	0.000	0.00	0.00
MWC	Molybdenum	0.000	0.00	0.00
IHP	Nickel	0.785	0.03	0.01
LMB	Nickel	0.295	0.01	0.00
LWC	Nickel	0.145	0.00	0.00
MWC	Nickel	0.351	0.01	0.01
backg	Nickel	0.406	0.01	0.01
IHP	Selenium	0.008	0.24	0.02
LMB	Selenium	0.014	0.45	0.04
LWC	Selenium	0.017	0.53	0.05
MWC	Selenium	0.045	1.43	0.14
backg	Selenium	0.017	0.53	0.05
IHP	Strontium	0.010	0.00	
LMB	Strontium	0.015	0.00	
LWC	Strontium	0.021	0.00	
MWC	Strontium	0.010	0.00	
IHP	Thallium	0.000	0.00	0.00
LMB	Thallium	0.000	0.00	0.00
LWC	Thallium	0.000	0.00	0.00
MWC	Thallium	0.000	0.00	0.00
backg	Thallium	0.000	0.00	0.00
IHP	Tin	0.000	0.00	0.00
LMB	Tin	0.000	0.00	0.00
LWC	Tin	0.000	0.00	0.00
MWC	Tin	0.000	0.00	0.00
IHP	Vanadium	0.000	0.00	0.00
LMB	Vanadium	0.000	0.00	0.00
LWC	Vanadium	0.000	0.00	0.00
MWC	Vanadium	0.000	0.00	0.00
backg	Vanadium	0.002	0.01	0.00
IHP	Zinc	8.840	0.07	0.04
LMB	Zinc	6.039	0.05	0.02
LWC	Zinc	13.870	0.11	0.06
MWC	Zinc	25.859	0.21	0.11
backg	Zinc	9.380	0.08	0.04
LMB	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
LWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00
MWC	bis(2-Ethylhexyl)phthalate	0.000	0.00	0.00

^a Biota estimates = UCL soil x UCL uptake factor.

Table C.19. Results of Monte Carlo Simulation of Exposure for wildlife in WAG 2

Species	Analyte	Mean	STD	80th Percen.	%>NOAEL	%>LOAEL
Short-tailed Shrew	Aroclor 1260	0.624	0.265	0.823	> 95%	35-40%
	Arsenic	0.976	0.177	1.126	> 95%	< 5%
	Barium	20.617	2.437	22.551	> 95%	< 5%
	Cadmium	3.475	0.856	4.222	> 95%	70%
	Chromium	56.165	11.929	65.749	> 95%	> 95%
	Copper	9.414	2.423	11.387	< 5%	< 5%
	Manganese	214.645	51.290	258.606	65%	< 5%
	Mercury	6.753	4.272	9.921	> 95%	> 95%
	Molybdenum	5.362	0.351	5.661	< 5%	< 5%
	Nickel	48.614	15.456	60.623	< 5%	< 5%
	Selenium	1.093	0.297	1.335	> 95%	75%
	Vanadium	2.341	0.175	2.486	> 95%	< 5%
	Zinc	134.327	24.675	154.142	< 5%	< 5%
White-footed Mice	Aroclor 1260	0.084	0.041	0.117	70%	< 5%
	Chromium	7.054	1.482	8.246	60-65%	< 5%
	Mercury	0.856	0.546	1.264	> 95%	80-85%
	Selenium	0.135	0.037	0.165	90-95%	< 5%
Red Fox	Aroclor 1260	0.049	0.022	0.066	< 5%	< 5%
	Mercury	0.241	0.126	0.337	> 95%	65-70 %
	Selenium	0.034	0.006	0.039	> 95%	< 5%
Mink ¹	Mercury	0.011	0.005	0.015	~ 20%	< 5%
Mink ²	Mercury	0.098	0.061	0.145	> 95%	90-95%
White-tailed Deer	Mercury	0.010	0.004	0.013	55-60%	< 5%
Belted Kingfisher ¹	Mercury	0.072	0.011	0.080	> 95%	70-75%
Red-tailed Hawk	Mercury	0.153	0.092	0.229	> 95%	80-85%
Wild Turkey	Mercury	0.055	0.028	0.075	> 95%	30-35%

¹ Exposure through consumption of fish only.² Exposure through consumption of small mammals, vegetation, invertebrates, and soil.

Table C.20. Experimental Information for Derivation of Mammalian NOAEL's and LOAEL's

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation
Acetone	NA	rat	10 ² 90 d.	50 ² 90 d.	liver, kidney damage	EPA 1986c
Aluminum	AlCl ₃	mouse	1.93 ¹ 3 gen.	19.3 3 gen.	reproduction	Ondreicka et al. 1966
Antimony	potassium tartrate	mouse	0.125 lifetime	1.25 lifetime	reproduction	Schroeder et al. 1968
Aroclor-1248	NA	rhesus monkey	0.01 ¹ 14 mo.	0.1 14 mo.	reproduction	Barsotti et al. 1976
Aroclor-1254	NA	mink	0.14 4.5 mo.	0.69 4.5 mo.	reproduction	Aulerich and Ringer 1977
Arsenic	As+3	mouse	0.126 ¹ 3 gen.	1.26 3 gen.	reproduction	Schroeder and Mitchner 1971
Barium	chloride	rat	5.1 16 mo.		growth, hypertension	Perry et al. 1983
Barium	chloride	rat		19.8 ² 10 d.	mortality	Borzelleca et al. 1988
Benzene	NA	mouse	26.36 ¹ 6-12 d, gest.	263.6 6-12 d, gest.	reproduction	Nawrot and Staples 1979
Benzo(a)pyrene	NA	mouse	1 ¹ 7-16 d, gest	10 7-16 d, gest	reproduction	Mackenzie and Angevine 1981
Beryllium	sulfate	rat	0.66 1126 d.		longevity/ wt. loss	Schroeder and Mitchner 1975

Table 6.20 (Continued)

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation
Bis(2-ethylhexyl) phthalate	NA	mouse	18.3 105 d.	183 105 d.	reproduction	Lamb et al. 1987
Boron	boric acid, borax	rat	28 3 gen.	93.6 3 gen.	reproduction	Weir and Fisher 1972
Cadmium	CdCl ₂	mouse	2.518 2 gen.	0.1913 2 gen.	reproduction	Schroeder and Mitchner 1971
Chloroform	NA	rat	15 ² 13 wk.	41 ² 13 wk.	liver, kidney, gonad cond.	Palmer et al. 1979
Chromium	Cr ⁺⁶	rat	3.28 1 yr.		wt. loss, food consumption	Mackenzie et al. 1958
Chromium	Cr ⁺⁶	rat		13.14 ² 3 mo.	Mortality	Steven et al. 1976
Copper	sulfate	mink	11.71 1 yr.	15.14 1 yr.	reproduction	Aulerich et al. 1982
Cyanide	potassium cyanide	rat	68.7 gest. lact.		reproduction	Tewe and Maner 1981
1,2 Dichloroethene	NA	mouse	45.2 ² 90 d.		body, organ weight, blood chem. hepatic function	Palmer et al. 1979
Di-n-butylphthalate	NA	mouse	550 105 d.	1833 105 d.	reproduction	Lamb et al. 1987
Lead	acetate	rat	8 3 gen.	80 3 gen.	reproduction	Azar et al. 1973
Lithium	carbonate	rat	9.4 gest.	18.8 gest.	reproduction	Marathe and Thomas 1986

Table 6.20 (Continued)

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation
Manganese	oxide	rat	88 224 d., gest.	284 224 d., gest.	reproduction	Laskey et al. 1982
Mercury	methyl	mink	0.015 ² 93 d.	93 d.	mortality	Wobeser et al. 1976
Mercury	methyl	rat	3 gen.	0.16 3 gen.	reproduction	Verschuuren et al. 1976
Methylene Chloride	NA	rat	5.85 2 yr	50 2 yr	Liver Histology	NCA 1982
Molybdenum	molybdate	mouse	0.26 ¹ 3 gen.	2.6 3 gen.	reproduction	Schroeder and Mitchner 1971
Nickel	sulfate	rat	40 3 gen.	80 3 gen.	reproduction	Ambrose et al. 1976
Selenium	selenate	mouse	0.075 ¹ 3 gen.	0.75 3 gen.	reproduction	Schroeder and Mitchner 1971
Strontium	chloride	rat	263 3 yr.	3 yr.	body weight, bone changes	Skoryna 1981
1,1,2,2-Tetrachloroethene	NA	mouse	1.4 ² 6 wk.	7 ² 6 wk.	Hepatotoxicity	Buben and O'Flaherty 1985
Thallium	sulfate	rat	0.0074 ^{1,2} 60 d.	0.074 ^{1,2} 60 d.	reproduction	Formigli et al. 1986
Tin	(TBTO)	mouse	23.4 6-15 d, gest.	35 6-15 d, gest.	reproduction	Davis et al. 1987
Trichloroethene	NA	mouse	0.7 ^{1,2} 6 wk.	7 ² 6 wk.	hepatotoxicity	Buben and O'Flaherty 1985

Table 6.20 (Continued)

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation
1,1,1 Trichloroethane	NA	mouse	1000 3 gen.	3 gen.	reproduction	Lane et al. 1982
Vanadium	NaVO ₃	rat	0.21 ¹ 60 d. + gest.	2.1 60 d. + gest.	reproduction	Domingo et al. 1986
Vinyl chloride	NA	rat	0.17 ¹ lifetime	1.7 lifetime	longevity, mortality	Feron et al. 1981
Zinc	oxide	rat	160 gest.	320 gest.	reproduction	Schlicker and Cox 1968

¹ Estimated NOAEL; LOAEL to NOAEL factor of 10 applied.

² Estimated NOAEL; subchronic to chronic factor of 10 applied.

Table C-21. Estimated NOAELs and LOAELs for Mammalian Endpoints.

Contaminant	Estimated NOAEL's					Estimated LOAEL's				
	Shrew	Mouse	Mink	Fox	Deer	Shrew	Mouse	Mink	Fox	Deer
Acetone	21.978	19.972	7.692	5.281	2.806	109.892	99.858	38.458	26.405	14.028
Aluminum	2.295	2.086	0.803	0.551	0.293	22.952	20.856	8.032	5.515	2.930
Antimony	0.149	0.135	0.052	0.036	0.019	1.487	1.351	0.520	0.357	0.190
Aroclor 1248	0.043	0.039	0.015	0.010	0.005	0.427	0.388	0.150	0.103	0.055
Aroclor 1254	0.067	0.061	0.140	0.096	0.009	0.668	0.607	0.690	0.474	0.085
Arsenic	0.150	0.136	0.052	0.036	0.019	1.498	1.362	0.524	0.360	0.191
Barium	11.835	10.754	4.142	2.844	1.511	43.517	39.544	15.229	10.456	5.555
Benzo(a)pyrene	1.189	1.081	0.416	0.286	0.152	11.892	10.806	4.162	2.857	1.518
Beryllium	1.451	1.318	0.508	0.349	0.185					
Bis(2-ethylhexyl)phthalate	21.763	19.775	7.616	5.229	2.778	217.625	197.753	76.161	52.290	27.779
Boron	61.539	55.920	21.536	14.787	7.855	205.717	186.933	71.993	49.430	26.259
Cadmium	0.299	0.272	0.105	0.072	0.038	2.994	2.721	1.048	0.719	0.382
Chloroform	32.967	29.957	11.537	7.922	4.208	90.111	81.883	31.536	21.652	11.503
Chromium (Cr+6)	7.209	6.551	2.523	1.732	0.920	28.879	26.243	10.107	6.939	3.686
Copper	33.432	30.380	11.700	8.033	4.267	43.262	39.312	15.140	10.395	5.522
Cyanide	141.890	128.941	49.659	34.095	18.113					
1,2-Dichloroethene	53.752	48.844	18.811	12.915	6.861					
Di-n-butyl phthalate	654.066	594.341	228.899	157.157	83.490	2179.822	1980.776	762.858	523.761	278.249
Lead	17.583	15.977	6.153	4.225	2.244	175.826	159.772	61.533	42.248	22.444
Lithium	20.660	18.773	7.230	4.964	2.637	41.319	37.546	14.460	9.928	5.274
Manganese	193.409	175.749	67.686	46.473	24.688	624.140	567.191	218.441	149.980	79.676
Methylmercury	0.070	0.064	0.015	0.010	0.009	0.352	0.320	0.123	0.085	0.045
Methylene Chloride	12.857	11.683	4.500	3.089	1.641	109.892	99.858	38.458	26.405	14.028
Molybdenum	0.309	0.281	0.108	0.074	0.039	3.092	2.810	1.082	0.743	0.395
Nickel	87.913	79.886	30.766	21.124	11.222	175.826	159.772	61.533	42.248	22.444
Selenium (Sodium selenite)	0.090	0.082	0.032	0.022	0.012	0.904	0.821	0.316	0.217	0.115
Strontium	578.029	525.250	202.289	138.890	73.785					

1,1,2,2-Tetrachloroethene	1.665	1.513	0.583	0.400	0.213	8.324	7.564	2.913	2.000	1.063
Thallium	0.016	0.015	0.006	0.004	0.002	0.164	0.149	0.058	0.039	0.021
Tin	27.828	25.287	9.739	6.686	3.552	41.622	37.822	14.566	10.001	5.313
1,1,1-Trichloroethane	1235.930	1123.080	432.530	296.970	157.760					
Trichloroethene	0.832	0.756	0.291	0.200	0.106	8.324	7.564	2.913	2.000	1.063
Vanadium	0.428	0.389	0.150	0.103	0.055	4.285	3.894	1.500	1.030	0.547
Vinyl Chloride	0.374	0.340	0.131	0.090	0.048	3.736	3.395	1.308	0.898	0.477
Zinc	351.653	319.544	123.066	84.496	44.888	703.306	639.088	246.131	168.992	89.776

Table C.22 Estimated NOAEL's and LOAEL's for Avian Endpoints

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation	Estimated NOAEL (mg/kg/d)	Estimated LOAEL (mg/kg/d)
Aluminum	$Al_2(SO_4)_3$	ringed dove	109.7 4 mo.		reproduction	Carriere et al. 1986	109.7	
Aluminum	chloride	day-old white leghorn chicks	2 wks.	44.5 ² 2 wks.	mortality	Storer and Nelson 1968		44.5
Aroclor 1254	NA	Ring-necked pheasant	0.18 ² 17 wk	1.8 17 wk	reproduction	Dahlgren et al. 1972	0.18	1.8
Arsenic	arsenite	mallard duck	5.14 128d	12.84 128d	mortality	USFWS 1964	5.14	12.84
Arsenic	Paris Green	brown-headed cowbird	2.46 7 mo.	7.38 7 mo.	mortality	USFWS 1969	2.46	7.38
Barium	hydroxide	1-day old chicks	20.8 ² 4 wks.	41.7 ² 4 wks.	mortality	Johnson et al. 1960	20.8	41.7
bis(2-ethylhexyl) Phthalate	NA	ringed dove	1.1 4 wks.	4 wks.	reproduction	Peakall 1974	1.1	
Boron	boric acid	mallard duck	28.8 3 wks. prior, during, 3 wks. post reprod.	100 3 wks. prior, during, 3 wks. post reprod.	reproduction	Smith and Anders 1989	28.8	100
Cadmium	$CdCl_2$	mallard duck	1.45 90 d	20 90 d	reproduction	White and Finley 1978	1.45	20

Table 6.22 (Continued)

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration		LOAEL (mg/kg/d) and Duration	Endpoint	Citation	Estimated LOAEL (mg/kg/d)	
			1	5				1	5
Chromium	Cr ³⁺	black duck	10 mo.	10 mo.	5	reproduction	Haseltine et al., unpubl. data	1	5
Copper	oxide	chicken	33.2 10 wk	46.97 10 wk	46.97 10 wk	growth/ mortality	Mehring et al. 1960	33.2	46.97
Di-n-butylphthalate	NA	ringed dove	0.11 ¹ 4 wks.	1.1 4 wks.	1.1 4 wks.	reproduction	Peakall 1974	0.11	
Lead	metal	American Kestrel	3.85 7 mo.			reproduction	Pattee 1984	3.85	
Lead	acetate	Japanese quail		11.3 12 wk		reproduction	Edens et al. 1976		11.3
Manganese	oxide	Japanese quail	977 75 d.		75 d.	growth, aggressiveness	Laskey and Edens 1985	977	
Mercury	methyl	mallard duck	0.0064 ² 3 gen.	0.064 3 gen.	0.064 3 gen.	reproduction	Heinz 1979	0.0064	0.064
Molybdenum	sodium Mo	chicken	3.5 ¹ 21 d.	35.3 21 d.	35.3 21 d.	reproduction	Lepore and Miller 1965	3.5	35.3
Nickel	sulfate	mallard duck	77.4 90 d	107 90 d	107 90 d	mortality, growth, behavior	Cain and Pafford 1981	77.4	107
Selenium	selenite	mallard duck	0.5 10 wk	1.0 10 wk	1.0 10 wk	reproduction	Heinz et al. 1987	0.5	1.0
Tin	(TBTO)	Japanese quail	6.8 6 wks.	16.9 6 wks.	16.9 6 wks.	reproduction	Schlatterer et al. 1993	6.8	16.9

Table 6.22 (Continued)

Contaminant	Form	Test Species	NOAEL (mg/kg/d) and Duration	LOAEL (mg/kg/d) and Duration	Endpoint	Citation	Estimated NOAEL (mg/kg/d)	Estimated LOAEL (mg/kg/d)
Vanadium	vanadyl sulfate	mallard duck	11.4 12 wks.	12 wks.	mortality, body weight, blood chem.	White and Dieter 1978	11.4	
Zinc	zinc sulfate	chicken	14.5 44 wk	130.9 44 wk	reproduction	Stahl et al. 1990	14.5	1309

¹ Estimated NOAEL: LOAEL to NOAEL factor of 10 applied.

² Estimated NOAEL: subchronic to chronic factor of 10 applied.

Table C.23 Water Benchmark Values for Wildlife Endpoints

Contaminant	Estimated NOAEL's					Estimated LOAEL's						
	Shrew	Mouse	Mink	Fox	Hawk	Shrew	Mouse	Mink	Fox	Deer	Kingfisher	Hawk
Acetone	99.90	66.57	77.69	62.54	42.84	499.51	332.86	388.47	312.69	214.20		
Aluminum	10.43	6.95	8.11	6.53	4.47	104.33	69.52	81.13	65.31	44.74	411.63	782.92
Antimony	0.68	0.45	0.53	0.42	0.29	6.76	4.50	5.26	4.23	2.90		
Aroclor 1248	0.19	0.13	0.15	0.12	0.08	1.94	1.29	1.51	1.22	0.83		
Aroclor 1254	0.30	0.20	1.41	1.14	0.13	3.08	2.02	6.97	5.61	1.30	16.65	31.67
Arsenic(Sodium Arsenite)	0.68	0.45	0.53	0.43	0.29	6.81	4.54	5.30	4.26	2.92	118.77	225.90
Barium	53.80	35.85	41.84	33.68	23.07	197.81	131.81	153.83	123.83	84.83	385.73	733.66
Benzo(a)pyrene	5.41	3.60	4.20	3.38	2.32	54.06	36.02	42.04	33.84	23.18		
Beryllium	6.59	4.39	5.13	4.13	2.83							
Bis(2-ethylhexyl)phthalate	98.92	65.92	76.93	61.92	42.42	989.21	659.18	769.30	619.23	424.20		
Boron	279.72	186.40	217.54	175.11	119.95	935.08	623.11	727.21	585.36	400.99	925.00	1759.38
Cadmium	0.08	0.05	0.06	0.05	0.03	0.10	0.07	0.08	0.06	0.04	185.00	351.88
Carbon Tetrachloride	159.84	106.52	124.31	100.06	68.55							
Chloroform	149.85	99.86	116.54	93.81	64.26	409.60	272.94	318.54	256.41	175.65		
Chromium (Cr+3)	27343.00	18220.70	21264.60	17116.70	11725.50							
Chromium (Cr+6)	32.77	21.84	25.48	20.51	14.05	131.27	87.48	102.09	82.18	56.29	46.25	87.97
Copper	151.96	101.27	118.18	95.13	65.17	196.64	131.04	152.93	123.10	84.33	570.73	1085.53
1,2-Dichloroethene	244.33	162.81	190.01	152.95	104.78							
Di-n-butyl phthalate	2973.03	1981.14	2312.11	1861.07	1274.92	9908.28	6602.58	7705.64	6202.44	4248.94	10.18	19.35
Lead (Metal)	79.92	53.26	62.15	50.03	34.27	35.61	67.74					
Lead (Acetate)						10.45	19.88					
Lithium	93.91	62.58	73.03	58.79	40.27	799.21	532.57	621.54	500.31	342.73	104.53	198.81
Manganese	879.13	585.83	683.70	550.34	377.00	187.82	125.16	146.06	117.57	80.54		
Methylmercury	0.32	0.21	0.15	0.12	0.14	1.60	1.07	1.24	1.00	0.69	0.59	1.13
Methylene Chloride	58.44	38.94	45.45	36.59	25.06	499.51	332.86	388.47	312.69	214.20		
Molybdenum	1.41	0.94	1.09	0.88	0.60	14.05	9.37	10.93	8.80	6.03	326.53	621.06
Nickel	399.61	266.29	310.77	250.15	171.36	799.21	532.57	621.54	500.31	342.73	989.75	1882.53
Selenium(Sodium Selenite)	0.41	0.27	0.32	0.26	0.18	4.11	2.74	3.20	2.57	1.76	9.25	17.59
Strontium	2627.41	1750.84	2043.32	1644.75	1126.71							
1,1,2,2-Tetrachloroethene	7.57	5.04	5.89	4.74	3.25	37.84	25.21	29.43	23.69	16.23		
Thallium	0.08	0.05	0.06	0.05	0.03	0.75	0.50	0.58	0.47	0.32		

Tin	126.49	84.29	98.37	79.18	54.24	62.90	119.64	189.19	126.07	147.13	118.43	1.13	156.33	297.33
1,1,1-Trichloroethane	5617.86	3743.60	4368.99	3516.75	2409.04									
Trichloroethene	3.78	2.52	2.94	2.37	1.62			37.84	25.21	29.43	23.69	16.23		
Vanadium	1.95	1.30	1.52	1.22	0.84	105.45	200.57	19.48	12.98	15.15	12.19	8.35		
Vinyl Chloride	1.70	1.13	1.32	1.06	0.73			16.98	11.32	13.21	10.63	7.23		
Zinc	1598.42	1065.15	1243.09	1000.61	685.45	134.13	255.11	3196.84	2130.29	2486.17	2001.22	1370.90	1211.75	2304.78
Zirconium	9.41	6.27	7.32	5.89	4.03									

Table C.24 Contaminant concentrations (mg/kg) found in Kingfisher egg shells and feathers found on the ORR.

Matrix	Burrow ^a	As	Cd	Se	Pb	Hg	⁶⁰ Co (pCi/g)	¹³⁷ Cs (pCi/g)
egg shell	CRD	0.135	<0.0333 ^b	1.58	2.0	<0.020	<7.45	<9.09
egg shell	WOC	0.0536	0.0583	1.41	5.31	0.182	<1.89	58.1_ 1.7
feathers	CRU	0.074	0.0132	5.72	0.657	1.03	<0.21	<0.18
feathers	CRU	0.0449	<0.0102	6.54	1.42	1.01	<0.18	<0.19
feathers	CRU	0.052	<0.010	5.72	1.67	1.04	<0.17	<0.18
feathers	CRU	0.0755	0.0755	6.83	1.91	0.726	<0.25	<0.16

^a CRD = Clinch River downstream of WOL Embayment; WOC = White Oak Creek downstream of WCK 3.5; CRU = Clinch River upstream of Oak Ridge Reservation.

^b Less than values are below minimum detection limit.

Table C.25. Contaminant concentrations in tissues of the three kingfishers found on the ORR

Bird No.	Watershed and Location	Organ	¹³⁷Cs (pCi/g)	Cd (mg/kg)^a	Pb (mg/kg)^a	Se (mg/kg)	Hg (mg/kg)
1	East Fork Poplar Creek, Lake Reality	whole body	< 2				
		feathers		ND	2.67	5.38	13.9
		kidney		4.04	ND	5.81	8.65
		liver		0.95	ND	2.71	3.69
		heart		ND	ND	1.25	1.1
		muscle		ND	ND	ND	0.572
2	East Fork Poplar Creek	feathers		7.21	1.86	5.63	4.55
		kidney		0.40	ND	3.14	1.46
		liver		0.23	ND	3.45	0.955
		heart		ND	ND	2.01	0.594
		muscle	3	ND	ND	1.04	0.805
3	White Oak Creek, Bldg. 4505	whole body	13,690				
		feathers		0.34	4.88	7.29	2.72
		kidney	69	1.53	0.42	6.01	26.8
		liver	76	0.90	0.40	7.5	17.6
		heart	81	ND	ND	2.2	9.52
		muscle	151	ND	0.58	1.84	6.34

^aND= Nondetect: As- <0.40 mg/kg, Cd- <0.20 mg/kg, Pb- <0.40 mg/kg, and Se-<0.40 mg/kg.

Table C.26 Estimated Densities of Wildlife Populations within WAG 2

Endpoint	Population Density ¹	Units	Amount of Suitable Habitat	Units	Estimated Population Size ¹
Short-tailed Shrew	23 ha		60.56 ha		1393
White-footed Mouse	57 ha		60.62 ha		3455
White-tailed Deer	0.1704 ha		60.62 ha		10
Red Fox	0.077 ha		60.62 ha		5
Red-Tailed Hawk	0.03 ha		60.62 ha		2
Wild Turkey	0.0426 ha		60.62 ha		3
Mink	0.6 stream km		6.4 stream km		4
Belted Kingfisher	0.4 stream km		6.4 stream km		3

¹Number of individuals

Table 6.27. Summary of the number of individuals of wildlife endpoints estimated to be experiencing adverse effects in WAG 2

Species	Analyte	%>LOAEL	Number in Area	Number Adversely Affected	Percent Adversely Affected
Short-tailed Shrew	Aroclor 1260	35-40%	1393	488-557	35-40%
	Arsenic	< 5%	1393	0	0%
	Barium	< 5%	1393	0	0%
	Cadmium	70%	1393	975	70%
	Chromium	> 95%	1393	1393	100%
	Copper	< 5%	1393	0	0%
	Manganese	< 5%	1393	0	0%
	Mercury	> 95%	1393	1393	100%
	Molybdenum	< 5%	1393	0	0%
	Nickel	< 5%	1393	0	0%
	Selenium	75%	1393	1045	75%
	Vanadium	< 5%	1393	0	0%
	Zinc	< 5%	1393	0	0%
White-footed Mice	Aroclor 1260	< 5%	3455	0	0%
	Chromium	< 5%	3455	0	0%
	Mercury	80-85%	3455	3455	100%
	Selenium	< 5%	3455	0	0%
Red Fox	Aroclor 1260	< 5%	5	0	0%
	Mercury	65-70 %	5	3	60%
	Selenium	< 5%	5	0	0%
Mink ¹	Mercury	< 5%	4	0	0%
Mink ²	Mercury	90-95%	4	4	100%
White-tailed Deer	Mercury	< 5%	10	0	0%
Belted Kingfisher ¹	Mercury	70-75%	3	2	66%
Red-tailed Hawk	Mercury	80-85%	2	2	100%
Wild Turkey	Mercury	30-35%	3	1	33%

¹ Exposure through consumption of fish only.

² Exposure through consumption of small mammals, vegetation, invertebrates, and soil.

Table 6.28 (Continued)

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
1C	COPPER	0.0037	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1WC	COPPER	0.007	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
HRT	COPPER	0.002	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
UMB	COPPER	0.0149	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
W4T	COPPER	0.014	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	COPPER	0.0081	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	COPPER	0.0039	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1C	LEAD	0.002	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
BWC	LEAD	0.004	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
W4T	LEAD	0.0021	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	LEAD	0.0033	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	LEAD	0.002	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1C	MANGANESE	0.0339	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1WC	MANGANESE	0.0227	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
BWC	MANGANESE	0.008	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
HRT	MANGANESE	0.0509	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
NWT	MANGANESE	0.1486	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
RAC	MANGANESE	5.3231	0.006	0.009	0.008	0.010	0.014	0.000	0.001	0.000
UMB	MANGANESE	0.0745	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
W4T	MANGANESE	1.3784	0.002	0.002	0.002	0.003	0.004	0.000	0.000	0.000
WOL	MANGANESE	0.1769	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	MANGANESE	3.2234	0.004	0.006	0.005	0.006	0.009	0.000	0.000	0.000
RAC	MOLYBDENUM	0.012	0.009	0.013	0.011	0.014	0.020	0.000	0.000	0.000
W4T	MOLYBDENUM	0.0195	0.014	0.021	0.018	0.022	0.032	0.000	0.001	0.000
RAC	Methylene chloride	0.002	0.000	0.000	0.000	0.000	0.000			
W4T	NICKEL	0.0152	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	NICKEL	0.0141	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1C	STRONTIUM	0.0952	0.000	0.000	0.000	0.000	0.000			
1WC	STRONTIUM	0.1335	0.000	0.000	0.000	0.000	0.000			
BWC	STRONTIUM	0.036	0.000	0.000	0.000	0.000	0.000			
HRT	STRONTIUM	0.1074	0.000	0.000	0.000	0.000	0.000			
NWT	STRONTIUM	0.1691	0.000	0.000	0.000	0.000	0.000			
RAC	STRONTIUM	0.2323	0.000	0.000	0.000	0.000	0.000			
UMB	STRONTIUM	0.1997	0.000	0.000	0.000	0.000	0.000			
W4T	STRONTIUM	0.2301	0.000	0.000	0.000	0.000	0.000			
WOL	STRONTIUM	0.1048	0.000	0.000	0.000	0.000	0.000			
WOL	STRONTIUM	0.1164	0.000	0.000	0.000	0.000	0.000			
RAC	Trichloroethene	0.001	0.000	0.000	0.000	0.000	0.001			
W4T	Trichloroethene	0.001	0.000	0.000	0.000	0.000	0.001			
UMB	VANADIUM	0.002	0.001	0.002	0.001	0.002	0.002	0.000	0.000	0.000

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
WOL	VANADIUM	0.0038	0.002	0.003	0.003	0.003	0.005	0.000	0.000	0.000
1C	ZINC	0.0283	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
1WC	ZINC	0.2617	0.000	0.000	0.000	0.000	0.000	0.001	0.002	0.001
HRT	ZINC	0.0065	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
UMB	ZINC	0.114	0.000	0.000	0.000	0.000	0.000	0.000	0.001	0.000
W4T	ZINC	0.0127	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	ZINC	0.0528	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WOL	ZINC	0.0152	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
W4T	cis-1,2-Dichloroethene	0.0029	0.000	0.000	0.000	0.000	0.000			

Table C.29. NOAEL hazard quotients for water contaminants in White Oak Creek seeps.

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
WOCET	ALUMINUM	3.7796	0.362	0.544	0.466	0.579	0.845	0.001	0.004	0.002
MBTRIB-3	ALUMINUM	0.7349	0.070	0.106	0.091	0.113	0.164	0.000	0.001	0.000
MID. DRAIN.	ALUMINUM	0.1081	0.010	0.016	0.013	0.017	0.024	0.000	0.000	0.000
SW2-5	ALUMINUM	0.0977	0.009	0.014	0.012	0.015	0.022	0.000	0.000	0.000
SW5-2	ALUMINUM	0.078	0.007	0.011	0.010	0.012	0.017	0.000	0.000	0.000
SW5-4	ALUMINUM	0.0508	0.005	0.007	0.006	0.008	0.011	0.000	0.000	0.000
EAST SEEP	ALUMINUM	1.2622	0.121	0.182	0.156	0.193	0.282	0.000	0.001	0.001
MV-1	ALUMINUM	0.2748	0.026	0.040	0.034	0.042	0.061	0.000	0.000	0.000
SW7-3	ALUMINUM	0.548	0.053	0.079	0.068	0.084	0.122	0.000	0.001	0.000
SW7-5	ALUMINUM	1.1221	0.108	0.161	0.138	0.172	0.251	0.000	0.001	0.001
SW7-6	ALUMINUM	0.761	0.073	0.109	0.094	0.117	0.170	0.000	0.001	0.000
SW7-8	ALUMINUM	0.412	0.039	0.059	0.051	0.063	0.092	0.000	0.000	0.000
WCTTRIB-1	ALUMINUM	0.957	0.092	0.138	0.118	0.147	0.214	0.000	0.001	0.000
SW2-1	ALUMINUM	0.186	0.018	0.027	0.023	0.028	0.042	0.000	0.000	0.000
WCTTRIB-2	ALUMINUM	0.194	0.019	0.028	0.024	0.030	0.043	0.000	0.000	0.000
SW9-2	ALUMINUM	1.7906	0.172	0.258	0.221	0.274	0.400	0.001	0.002	0.001
MV-3	ALUMINUM	0.162	0.016	0.023	0.020	0.025	0.036	0.000	0.000	0.000
SW2-6	ALUMINUM	0.2012	0.019	0.029	0.025	0.031	0.045	0.000	0.000	0.000
SW2-7	ALUMINUM	0.038	0.004	0.005	0.005	0.006	0.008	0.000	0.000	0.000
BTT	ALUMINUM	0.0401	0.004	0.006	0.005	0.006	0.009	0.000	0.000	0.000
SW4-2	ALUMINUM	0.0415	0.004	0.006	0.005	0.006	0.009	0.000	0.000	0.000
WAG4 MS1	ALUMINUM	0.3133	0.030	0.045	0.039	0.048	0.070	0.000	0.000	0.000
WAG4 T2A	ALUMINUM	0.7102	0.068	0.102	0.088	0.109	0.159	0.000	0.001	0.000
RS-1	ALUMINUM	0.298	0.029	0.043	0.037	0.046	0.067	0.000	0.000	0.000
RS-3A	ALUMINUM	12.7297	1.220	1.831	1.569	1.949	2.845	0.004	0.013	0.007
RS-3B	ALUMINUM	1.8185	0.174	0.262	0.224	0.278	0.406	0.001	0.002	0.001
SW7-2	ALUMINUM	1.0665	0.102	0.153	0.131	0.163	0.238	0.000	0.001	0.001
SW4-1	ANTIMONY	0.045	0.067	0.100	0.086	0.106	0.155			
MID. DRAIN.	ARSENIC	0.001	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
EAST SEEP	ARSENIC	0.003	0.004	0.007	0.006	0.007	0.010	0.000	0.000	0.000
MV-1	ARSENIC	0.002	0.003	0.004	0.004	0.005	0.007	0.000	0.000	0.000
SW7-3	ARSENIC	0.003	0.004	0.007	0.006	0.007	0.010	0.000	0.000	0.000
SW2-6	ARSENIC	0.001	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW2-7	ARSENIC	0.001	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW4-1	ARSENIC	0.006	0.009	0.013	0.011	0.014	0.021	0.000	0.000	0.000
RS-3A	ARSENIC	0.0202	0.030	0.044	0.038	0.047	0.069	0.000	0.000	0.000
WOCET	Acetone	0.009	0.000	0.000	0.000	0.000	0.000			
SNNT	Acetone	0.008	0.000	0.000	0.000	0.000	0.000			
MV-1	Acetone	0.001	0.000	0.000	0.000	0.000	0.000			
MV-2	Acetone	0.003	0.000	0.000	0.000	0.000	0.000			
SW7-5	Acetone	0.014	0.000	0.000	0.000	0.000	0.000			

Table 6.29. (Continued)

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
SW2-1	Acetone	0.011	0.000	0.000	0.000	0.000	0.000			
SW2-4	Acetone	0.014	0.000	0.000	0.000	0.000	0.000			
WCTRI-2	Acetone	0.002	0.000	0.000	0.000	0.000	0.000			
SW2-2	Acetone	0.001	0.000	0.000	0.000	0.000	0.000			
SW6-1	Acetone	0.003	0.000	0.000	0.000	0.000	0.000			
WOCET	BARIUM	0.1121	0.002	0.003	0.003	0.003	0.005	0.000	0.001	0.000
WCTRI-3	BARIUM	0.062	0.001	0.002	0.001	0.002	0.003	0.000	0.000	0.000
MBTRI-3	BARIUM	0.1328	0.002	0.004	0.003	0.004	0.006	0.000	0.001	0.000
MID. DRAIN.	BARIUM	0.1126	0.002	0.003	0.003	0.003	0.005	0.000	0.001	0.000
SW2-5	BARIUM	0.3012	0.006	0.008	0.007	0.009	0.013	0.000	0.002	0.001
SW5-2	BARIUM	0.064	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW5-4	BARIUM	0.127	0.002	0.004	0.003	0.004	0.006	0.000	0.001	0.000
EAST SEEP	BARIUM	0.036	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
MV-1	BARIUM	0.16	0.003	0.004	0.004	0.005	0.007	0.000	0.001	0.000
SW7-3	BARIUM	0.0219	0.000	0.001	0.001	0.001	0.001	0.000	0.000	0.000
SW7-5	BARIUM	0.0704	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW7-6	BARIUM	0.049	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
SW7-8	BARIUM	0.03	0.001	0.001	0.001	0.001	0.001	0.000	0.000	0.000
WCTRI-1	BARIUM	0.13974	0.003	0.004	0.003	0.004	0.006	0.000	0.001	0.000
SW2-1	BARIUM	0.084	0.002	0.002	0.002	0.002	0.004	0.000	0.000	0.000
SW2-3	BARIUM	0.038	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
SW2-4	BARIUM	0.32395	0.006	0.009	0.008	0.010	0.014	0.001	0.002	0.001
WCTRI-2	BARIUM	0.038	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
SW9-1	BARIUM	0.14653	0.003	0.004	0.004	0.004	0.006	0.000	0.001	0.000
SW9-2	BARIUM	0.28725	0.005	0.008	0.007	0.009	0.012	0.000	0.001	0.001
MBTRI-2A	BARIUM	0.05	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
MV-3	BARIUM	0.049	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
SW2-2	BARIUM	0.104	0.002	0.003	0.002	0.003	0.005	0.000	0.001	0.000
SW2-6	BARIUM	0.16817	0.003	0.005	0.004	0.005	0.007	0.000	0.001	0.000
SW2-7	BARIUM	0.14008	0.003	0.004	0.003	0.004	0.006	0.000	0.001	0.000
BTT	BARIUM	0.30888	0.006	0.009	0.007	0.009	0.013	0.000	0.002	0.001
SW4-1	BARIUM	0.3623	0.007	0.010	0.009	0.011	0.016	0.001	0.002	0.001
SW4-2	BARIUM	0.42776	0.008	0.012	0.010	0.013	0.019	0.001	0.002	0.001
WAG4 MS1	BARIUM	0.15379	0.003	0.004	0.004	0.005	0.007	0.000	0.001	0.000
WAG4 T2A	BARIUM	0.10552	0.002	0.003	0.003	0.003	0.005	0.000	0.001	0.000
RS-1	BARIUM	0.036	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
RS-3A	BARIUM	0.0612	0.001	0.002	0.001	0.002	0.003	0.000	0.000	0.000
RS-3B	BARIUM	0.029	0.001	0.001	0.001	0.001	0.001	0.000	0.000	0.000
SW7-1	BARIUM	0.07831	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW7-2	BARIUM	0.032	0.001	0.001	0.001	0.001	0.001	0.000	0.000	0.000

Table 6.29. (Continued)

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
SW7-8	MANGANESE	0.126	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WCTRIB-1	MANGANESE	1.1979	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
SW2-1	MANGANESE	0.992	0.001	0.002	0.001	0.002	0.003	0.000	0.000	0.000
SW2-3	MANGANESE	0.093	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW2-4	MANGANESE	5.8866	0.007	0.010	0.009	0.011	0.016	0.000	0.001	0.000
WCTRIB-2	MANGANESE	0.024	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW9-1	MANGANESE	0.3602	0.000	0.001	0.001	0.001	0.001	0.000	0.000	0.000
SW9-2	MANGANESE	2.7056	0.003	0.005	0.004	0.005	0.007	0.000	0.000	0.000
MBTRIB-2A	MANGANESE	0.142	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
MV-3	MANGANESE	0.019	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW2-2	MANGANESE	0.796	0.001	0.001	0.001	0.001	0.002	0.000	0.000	0.000
SW2-6	MANGANESE	1.5626	0.002	0.003	0.002	0.003	0.004	0.000	0.000	0.000
SW2-7	MANGANESE	1.9677	0.002	0.003	0.003	0.004	0.005	0.000	0.000	0.000
BTT	MANGANESE	0.0117	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW4-1	MANGANESE	12.9311	0.015	0.022	0.019	0.024	0.034	0.000	0.001	0.001
SW4-2	MANGANESE	1.0607	0.001	0.002	0.002	0.002	0.003	0.000	0.000	0.000
WAG4 MS1	MANGANESE	0.3384	0.000	0.001	0.000	0.001	0.001	0.000	0.000	0.000
WAG4 T2A	MANGANESE	0.2263	0.000	0.000	0.000	0.000	0.001	0.000	0.000	0.000
RS-1	MANGANESE	1.35	0.002	0.002	0.002	0.002	0.004	0.000	0.000	0.000
RS-3A	MANGANESE	0.3108	0.000	0.001	0.000	0.001	0.001	0.000	0.000	0.000
RS-3B	MANGANESE	0.1381	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW7-1	MANGANESE	6.4531	0.007	0.011	0.009	0.012	0.017	0.000	0.001	0.000
SW7-2	MANGANESE	0.2533	0.000	0.000	0.000	0.000	0.001	0.000	0.000	0.000
RS-3A	MERCURY	0.0011	0.003	0.005	0.007	0.009	0.008	0.006	0.019	0.010
EAST SEEP	MOLYBDENUM	0.015	0.011	0.016	0.014	0.017	0.025	0.000	0.000	0.000
SW7-3	MOLYBDENUM	0.0142	0.010	0.015	0.013	0.016	0.024	0.000	0.000	0.000
BTT	MOLYBDENUM	0.1145	0.081	0.122	0.105	0.130	0.190	0.001	0.004	0.002
SW4-2	MOLYBDENUM	0.008	0.006	0.009	0.007	0.009	0.013	0.000	0.000	0.000
WAG4 MS1	MOLYBDENUM	0.0074	0.005	0.008	0.007	0.008	0.012	0.000	0.000	0.000
RS-3A	MOLYBDENUM	0.0517	0.037	0.055	0.047	0.059	0.086	0.000	0.002	0.001
WOCET	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
5NNT	Methylene chloride	0.013	0.000	0.000	0.000	0.000	0.001			
MID. DRAIN.	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
MV-1	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
SW7-3	Methylene chloride	0.0037	0.000	0.000	0.000	0.000	0.000			
SW7-5	Methylene chloride	0.03	0.001	0.001	0.001	0.001	0.001			
SW2-1	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
SW2-3	Methylene chloride	0.0083	0.000	0.000	0.000	0.000	0.000			
SW2-4	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
SW9-2	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			

Table 6.29. (Continued)

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
SW7-2	Methylene chloride	0.001	0.000	0.000	0.000	0.000	0.000			
MID. DRAIN.	NICKEL	0.006	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW2-5	NICKEL	0.013	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW5-4	NICKEL	0.0138	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
EAST SEEP	NICKEL	0.0319	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW7-3	NICKEL	0.028	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW7-5	NICKEL	0.0206	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW2-6	NICKEL	0.0139	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW2-7	NICKEL	0.008	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
BTT	NICKEL	0.048	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
SW4-2	NICKEL	8.5222	0.021	0.032	0.027	0.034	0.050	0.004	0.012	0.006
WAG4 MS1	NICKEL	0.0426	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
WAG4 T2A	NICKEL	0.0303	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
RS-3A	NICKEL	0.1031	0.000	0.000	0.000	0.000	0.001	0.000	0.000	0.000
EAST SEEP	SELENIUM	0.001	0.002	0.004	0.003	0.004	0.006	0.000	0.000	0.000
SW7-3	SELENIUM	0.003	0.007	0.011	0.009	0.012	0.017	0.000	0.001	0.000
RS-3A	SELENIUM	0.002	0.005	0.007	0.006	0.008	0.011	0.000	0.000	0.000
WOCET	STRONTIUM	0.154	0.000	0.000	0.000	0.000	0.000			
WTRIB-3	STRONTIUM	0.21	0.000	0.000	0.000	0.000	0.000			
MBTRIB-3	STRONTIUM	0.226	0.000	0.000	0.000	0.000	0.000			
MID. DRAIN.	STRONTIUM	0.207	0.000	0.000	0.000	0.000	0.000			
SW2-5	STRONTIUM	0.343	0.000	0.000	0.000	0.000	0.000			
SW5-2	STRONTIUM	0.201	0.000	0.000	0.000	0.000	0.000			
SW5-4	STRONTIUM	0.203	0.000	0.000	0.000	0.000	0.000			
EAST SEEP	STRONTIUM	0.132	0.000	0.000	0.000	0.000	0.000			
MV-1	STRONTIUM	0.169	0.000	0.000	0.000	0.000	0.000			
SW7-3	STRONTIUM	0.031	0.000	0.000	0.000	0.000	0.000			
SW7-5	STRONTIUM	0.114	0.000	0.000	0.000	0.000	0.000			
SW7-8	STRONTIUM	0.069	0.000	0.000	0.000	0.000	0.000			
WTRIB-1	STRONTIUM	0.383	0.000	0.000	0.000	0.000	0.000			
SW2-1	STRONTIUM	0.189	0.000	0.000	0.000	0.000	0.000			
SW2-3	STRONTIUM	0.067	0.000	0.000	0.000	0.000	0.000			
SW2-4	STRONTIUM	0.597	0.000	0.000	0.000	0.000	0.001			
WTRIB-2	STRONTIUM	0.061	0.000	0.000	0.000	0.000	0.000			
SW9-1	STRONTIUM	0.115	0.000	0.000	0.000	0.000	0.000			
SW9-2	STRONTIUM	0.197	0.000	0.000	0.000	0.000	0.000			
MBTRIB-2A	STRONTIUM	0.188	0.000	0.000	0.000	0.000	0.000			
MV-3	STRONTIUM	0.145	0.000	0.000	0.000	0.000	0.000			
SW2-2	STRONTIUM	0.124	0.000	0.000	0.000	0.000	0.000			
SW2-6	STRONTIUM	0.231	0.000	0.000	0.000	0.000	0.000			

Table 6.29. (Continued)

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
RS-3A	ZINC	0.04944	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
RS-3B	ZINC	0.009	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
MID. DRAIN.	cis-1,2-Dichloroethene	0.005	0.000	0.000	0.000	0.000	0.000			
SW4-1	cis-1,2-Dichloroethene	0.047	0.000	0.000	0.000	0.000	0.000			
SW4-2	cis-1,2-Dichloroethene	0.066	0.000	0.000	0.000	0.000	0.001			
WAG4 MS1	cis-1,2-Dichloroethene	0.002	0.000	0.000	0.000	0.000	0.000			
WAG4 T2A	cis-1,2-Dichloroethene	0.014	0.000	0.000	0.000	0.000	0.000			
FRENCH DR S	cis-1,2-Dichloroethene	0.14	0.001	0.001	0.001	0.001	0.001			
WAG6 MS2	cis-1,2-Dichloroethene	0.001	0.000	0.000	0.000	0.000	0.000			
WAG6 MS3B	cis-1,2-Dichloroethene	0.001	0.000	0.000	0.000	0.000	0.000			

Table C.30. LOAEL hazard quotients for water contaminants in White Oak Creek seeps.

Location	Contaminant	95% UCL	Shrew	Mouse	Mink	Fox	Deer	Turkey	Kingfisher	Hawk
RS-3A	ALUMINUM	12.7297	0.122	0.183	0.157	0.195	0.285	0.009	0.031	0.016

Table C.31. Radionuclide-specific ground and water dose coefficients and radiation energies (Eckerman and Ryman 1993).

Radionuclide [a]	Dose coefficients			Half lives (d)		Energies (MeV/nt)		
	DF _{gd} 0-5 (Sv m ³ /s Bq)	DF _{water} (Sv m ³ /s Bq)	rad. 1/2 life	biol. 1/2 life	alpha	beta	gamma	
Americium-241	2.18e-19	1.88e-18	1.58e+05	2.00e+04	5.48e+00	5.20e-02	3.30e-02	
Cesium-137	3.07e-21	1.49e-20	1.10e+04	1.15e+02	0.00e+00	1.87e-01	0.00e+00	
Barium-137m	1.09e-17	6.26e-17	1.77e-03	6.50e+01	0.00e+00	6.50e-02	5.97e-01	
Cobalt-60	4.45e-17	2.74e-16	1.92e+03	9.50e+00	0.00e+00	9.70e-02	2.50e+00	
Curium-244	6.74e-22	1.15e-20	6.61e+03	2.40e+04	5.80e+00	9.00e-03	2.00e-03	
Plutonium-238	7.60e-22	1.14e-20	3.20e+04	6.50e+04	5.49e+00	1.10e-02	2.00e-03	
Plutonium-239/240	9.47e-22	1.04e-20	2.39e+06	6.50e+04	5.15e+00	9.00e-03	1.00e-03	
Strontium-90	2.95e-21	1.46e-20	1.06e+04	4.00e+03	0.00e+00	1.96e-01	0.00e+00	
Yttrium-90	8.31e-20	3.63e-19	2.67e+00	1.40e+04	0.00e+00	9.35e-01	0.00e+00	
Technetium-99	5.74e-22	3.14e-21	7.77e+07	1.00e+00	0.00e+00	1.01e-01	0.00e+00	
Thorium-228	3.14e-20	2.05e-19	6.98e+02	5.70e+04	5.40e+00	2.10e-02	3.00e-03	
Radium-224	1.78e-19	1.03e-18	3.66e+00	8.10e+03	5.67e+00	2.00e-03	1.00e-02	
Radon-220	7.08e-21	4.03e-20	6.44e-04	0.00e+00	6.29e+00	0.00e+00	0.00e+00	
Polonium-216	3.07e-22	1.80e-21	1.74e-06	3.00e+01	6.78e+00	0.00e+00	0.00e+00	
Lead-212	2.53e-18	1.52e-17	4.43e-01	1.46e+03	0.00e+00	1.76e-01	1.48e-01	
Bismuth-212	3.34e-18	2.00e-17	4.20e-02	5.00e+00	2.17e+00	4.72e-01	1.86e-01	
Thallium-208	5.79e-17	3.84e-16	2.13e-03	5.00e+00	0.00e+00	5.98e-01	3.38e+00	
Polonium-212	0.00e+00	0.00e+00	3.53e-12	3.00e+01	8.79e+00	0.00e+00	0.00e+00	
Thorium-230	5.22e-21	3.94e-20	2.81e+07	5.70e+04	4.67e+00	1.50e-02	2.00e-03	
Thorium-232	2.36e-21	1.99e-20	5.13e+12	5.70e+04	4.00e+00	1.20e-02	1.00e-03	
Radium-228	0.00e+00	0.00e+00	2.10e+03	8.10e+03	0.00e+00	1.70e-02	0.00e+00	
Actinium-228	1.73e-17	1.04e-16	2.55e-01	2.40e+04	0.00e+00	4.75e-01	9.71e-01	
Uranium-233/234	3.56e-21	2.70e-20	5.79e+07	1.00e+02	4.79e+00	9.50e-03	1.50e-03	
Uranium-235	2.65e-18	1.59e-17	2.57e+11	1.00e+02	4.40e+00	4.90e-02	1.56e-01	
Thorium-231	1.59e-19	1.18e-18	1.06e+00	5.70e+04	0.00e+00	1.65e-01	2.60e-02	
Uranium-238	5.45e-22	7.95e-21	1.63e+12	1.00e+02	4.19e+00	1.00e-02	1.00e-03	
Thorium-234	1.07e-19	7.64e-19	2.41e+01	5.70e+04	0.00e+00	6.00e-02	9.00e-03	
Protactinium-234m	2.70e-19	1.52e-18	8.13e-04	4.10e+04	0.00e+00	8.22e-01	1.20e-02	

Protactinium-234	3.41e-17	2.03e-16	2.79e-01	4.10e+04	0.00e+00	4.94e-01	1.92e+00
[a] Indented radionuclides are short-lived daughter products of the parent radionuclides.							

Table C-32. Literature-derived bioaccumulation factors for mammals (MammBAF) and Oak Ridge Reservation-specific soil-to-tissue uptake factors for plants (browse, canopy, and herbaceous vegetation), earthworms, and small mammals (MammUF) used in estimation of dose rates in the White Oak Creek radiological assessment.

Radionuclide	Browse	Canopy	Herb	Earthworm	MammBAF	MammUF	Fish BCF (L/g)
Americium-241	2.49e+00 m	5.70e+00 m	2.18e+00 m	3.15e+00 m	2.00e-03 l	2.2682 m	0.03
Cesium-137	1.37e-02 l	1.37e-02 l	1.37e-02 l	1.00e+01 p	1.00e+01 l		2
Barium-137m	1.31e-01 e	2.41e-01 e	1.26e-01 e	1.19e-01 e	1.00e-02 l	0.045 e	0.004
Cobalt-60	5.00e-03 e	4.40e-03 e	9.90e-03 e	0.00e+00 e	5.00e-03 l	0.0099 e	0.3
Curium-244	1.09e+00 s	1.07e+00 m	1.09e+00 m	8.86e-01 m	1.00e-03 l	2.6933 m	0.03
Plutonium-238	1.80e+00 s	5.20e+00 s	5.00e+00 s	2.50e+00 s	5.00e-04 l	2.7333 s	0.03
Plutonium-239/240	1.80e+00 m	5.20e+00 m	5.00e+00 m	2.50e+00 m	5.00e-04 l	2.7333 m	0.03
Strontium-90	2.05e-01 l	2.05e-01 l	2.05e-01 l	1.69e-01 m	5.00e+00 l		0.06
Yttrium-90	1.00e-03 l	1.00e-03 l	1.00e-03 l	5.00e-02 p	5.00e-02 l		0.03
Technetium-99	2.08e+02 l	2.08e+02 l	2.08e+02 l	2.08e+02 p	5.00e-05 l		0.02
Thorium-228	1.37e-04 l	1.37e-04 l	1.37e-04 l	5.00e-03 p	5.00e-03 l		0.1
Radium-224	9.30e-03 l	9.30e-03 l	9.30e-03 l	5.00e-02 p	5.00e-02 l		0.05
Radon-220							
Polonium-216	9.12e-05 l	9.12e-05 l	9.12e-05 l	2.50e-01 p	2.50e-01 l		0.05
Lead-212	1.08e-02 e	6.15e-02 e	2.54e-02 e	3.81e-01 e	2.00e-02 l	0.0267 e	0.3
Bismuth-212	8.75e-03 l	8.75e-03 l	8.75e-03 l	2.00e-02 p	2.00e-02 l		0.01
Thallium-208	5.80e-03 s	5.80e-03 s	5.80e-03 e	0.00e+00 e	2.00e+00 l		
Polonium-212	9.12e-05 l	9.12e-05 l	9.12e-05 l	2.50e-01 p	2.50e-01 l		0.05
Thorium-230	1.37e-04 l	1.37e-04 l	1.37e-04 l	5.00e-03 p	5.00e-03 l		0.1
Thorium-232	1.37e-04 l	1.37e-04 l	1.37e-04 l	5.00e-03 p	5.00e-03 l		0.1
Radium-228	9.30e-03 l	9.30e-03 l	9.30e-03 l	5.00e-02 p	5.00e-02 l		0.05
Actinium-228	8.75e-04 l	8.75e-04 l	8.75e-04 l	1.25e-03 p	1.25e-03 l		
Uranium-233/234	1.00e-04 e	6.00e-04 e	1.50e-03 e	6.30e-02 e	1.50e-02 l	0.0003 e	0.01
Uranium-235	1.00e-04 e	6.00e-04 e	1.50e-03 e	6.30e-02 e	1.50e-02 l	0.0003 e	0.01
Thorium-231	1.37e-04 l	1.37e-04 l	1.37e-04 l	5.00e-03 p	5.00e-03 l		0.1
Uranium-238	1.00e-04 e	6.00e-04 e	1.50e-03 e	6.30e-02 e	1.50e-02 l	0.0003 e	0.01
Thorium-234	1.37e-04 l	1.37e-04 l	1.37e-04 l	5.00e-03 p	5.00e-03 l		0.1
Protactinium-234m	2.00e-03 l	2.00e-03 l	2.00e-03 l	2.00e-03 p	5.00e-08 l		0.01

Protactinium-234	2.00e-03	l	2.00e-03	l	2.00e-03	l	2.00e-03	p	5.00e-08	l	0.01
------------------	----------	---	----------	---	----------	---	----------	---	----------	---	------

m = Uptake factor measured for given radionuclide
s = Assumed uptake is same as that for related isotope for which measured value was available
e = Uptake factor for elemental form used for radionuclide
l = Literature value from IAEA (1994), NCRP (1989), or Baes et al. (1984). Literature biotransfer factors (d/kg) have been converted to unitless uptake factors assuming a cow ingestion rate of 50 kg wet food/d.
p = Uptake factor for earthworms was unavailable. Used the larger of the herbaceous plant and small mammal values. See Sec. 6.1 for further discussion of derivation of uptake factors.

Table C.33. Overall dose rates from internal and external exposures for plants, earthworms, and terrestrial wildlife exposed to radionuclides in soil at WAG2 Intermediate Holding Pond (IHP).

Receptor	Dose rate (pCi/g)	Primary contributors (% of total dose rate)	
Plants	4340	Plutonium-239/240	(66.6%)
		Cesium-137	(17.1%)
		Americium-241	(10.7%)
		Plutonium-238	(3.0%)
Earthworms	2850	Plutonium-239/240	(43.9%)
		Cesium-137	(26.0%)
		Americium-241	(22.5%)
		Plutonium-238	(2.3%)
Short-tailed shrew	2283	Plutonium-239/240	(60.1%)
		Americium-241	(20.3%)
		Cesium-137	(8.7%)
		Curium-244	(7.1%)
White-footed mouse	2177	Plutonium-239/240	(63.0%)
		Americium-241	(21.2%)
		Curium-244	(7.4%)
		Cesium-137	(4.7%)
Wild turkey	150	Cesium-137	(91.3%)
White-tailed deer	84	Cesium-137	(91.4%)
Red fox	326	Cesium-137	(94.6%)
Red-tailed hawk	283	Cesium-137	(94.0%)
Mink	180	Cesium-137	(95.3%)
Belted kingfisher ¹	64	Cesium-137	(98.7%)

¹ Kingfisher dose rate estimate is based on external exposures to radionuclides in soil. Water-related exposures (water and fish ingestion) will be added when water data become available.

Table C.34. Overall dose rates from internal and external exposures for plants, earthworms, and terrestrial wildlife exposed to radionuclides in soil at WAG2 Middle White Oak Creek (MWC).

Receptor	Dose rate (pCi/g)	Primary contributors (% of total dose rate)	
Plants	1400	Plutonium-239/240	(83.1%)
		Americium-241	(9.8%)
		Plutonium-238	(3.0%)
Earthworms	800	Plutonium-239/240	(62.9%)
		Americium-241	(23.7%)
		Cesium-137	(4.0%)
Short-tailed shrew	734	Plutonium-239/240	(75.0%)
		Americium-241	(18.6%)
		Plutonium-238	(3.3%)
White-footed mouse	723	Plutonium-239/240	(76.1%)
		Americium-241	(18.8%)
		Plutonium-238	(3.4%)
Wild turkey	11	Cesium-137	(53.8%)
White-tailed deer	5	Cesium-137	(63.4%)
Red fox	17	Cesium-137	(77.3%)
Red-tailed hawk	14	Cesium-137	(81.2%)
Mink	9	Cesium-137	(84.3%)
Belted kingfisher ¹	3	Cesium-137	(93.4%)

¹ Kingfisher dose rate estimate is based on external exposures to radionuclides in soil. Water-related exposures (water and fish ingestion) will be added when water data become available.

Table C.35. Overall dose rates from internal and external exposures for plants, earthworms, and terrestrial wildlife exposed to radionuclides in soil at WAG2 Lower White Oak Creek (LWC).

Receptor	Dose rate (pCi/g)	Primary contributors (% of total dose rate)	
Plants	1350	Plutonium-239/240	(74.0%)
		Americium-241	(12.2%)
		Cesium-137	(8.1%)
Earthworms	846	Plutonium-239/240	(51.4%)
		Americium-241	(27.2%)
		Cesium-137	(13.0%)
Short-tailed shrew	739	Plutonium-239/240	(64.3%)
		Americium-241	(22.4%)
		Curium-244	(6.2%)
White-footed mouse	722	Plutonium-239/240	(65.8%)
		Americium-241	(22.9%)
		Curium-244	(6.3%)
Wild turkey	24	Cesium-137	(84.2%)
White-tailed deer	13	Cesium-137	(85.6%)
Red fox	49	Cesium-137	(92.9%)
Red-tailed hawk	42	Cesium-137	(93.7%)
Mink	27	Cesium-137	(94.3%)
Belted kingfisher ¹	10	Cesium-137	(95.7%)

¹ Kingfisher dose rate estimate is based on external exposures to radionuclides in soil. Water-related exposures (water and fish ingestion) will be added when water data become available.

Table C.36. Overall dose rates from internal and external exposures for plants, earthworms, and terrestrial wildlife exposed to radionuclides in soil at WAG2 Lower Melton Branch (LMB).

Receptor	Dose rate (pCi/g)	Primary contributors (% of total dose rate)	
Plants	3010	Plutonium-239/240	(66.6%)
		Cesium-137	(17.1%)
		Americium-241	(10.7%)
		Plutonium-238	(3.0%)
Earthworms	3050	Strontium-90	(92.1%)
		Americium-241	(2.1%)
Short-tailed shrew	622	Strontium-90	(58.3%)
		Curium-244	(26.7%)
		Americium-241	(7.5%)
White-footed mouse	498	Strontium-90	(49.7%)
		Curium-244	(33.4%)
		Americium-241	(9.4%)
Wild turkey	366	Strontium-90	(96.2%)
White-tailed deer	275	Strontium-90	(96.9%)
Red fox	1087	Strontium-90	(97.6%)
Red-tailed hawk	1235	Strontium-90	(98.4%)
Mink	568	Strontium-90	(97.3%)
Belted kingfisher ¹	9	Cesium-137	(47.3%)
		Cobalt-60	(37.5%)

¹ Kingfisher dose rate estimate is based on external exposures to radionuclides in soil. Water-related exposures (water and fish ingestion) will be added when water data become available.

DISTRIBUTION

1. T. L. Ashwood
2. L. V. Asplund
3. D. M. Borders
4. H. L. Boston
5. W. D. Brickeen
6. B. B. Burgoa
7. R. B. Clapp
8. A. H. Curtis
9. M. F. P. DeLozier
10. A. F. Diefendorf
11. R. A. Efroymsen
12. C. J. Ford
13. S. B. Garland
14. C. S. Haase
15. D. S. Hicks
16. S. G. Hildebrand
17. D. D. Huff
18. D. S. Jones
19. K. L. Kaiser
20. R. H. Ketelle
21. E. A. Krispin
22. A. J. Kuhaida, Jr
23. D. M. Matteo
24. W. L. McCalla
25. P. L. Osborne
- 26-27. P. T. Owen
28. S. T. Puraker
29. D. E. Reichle
30. C. T. Rightmire
31. B. E. Sample
32. P. A. Schrandt
33. J. A. Shaakir-Ali
34. J. G. Smith
35. M. R. Smith
36. S. H. Stow
- 37-41. G. W. Suter
42. D. R. Watkins
43. C. J. E. Welsh
44. A. S. Will
45. Central Research Library
46. ER Document Management Center—RC
47. Office of Scientific and Technical Information, P. O. Box 62, Oak Ridge, TN 37831
48. P. A. Hoffmann, DOE Oak Ridge Operations Office, P. O. Box 2001, Oak Ridge, TN 37831-8541
49. G. D. Reed, University of Tennessee, Department of Civil and Environmental Engineering, 223 Perkins Hall, Knoxville, TN 37996-2010