

On the relationship between stomatal characters and atmospheric CO₂

Chantal D. Reid,¹ Hafiz Maherali,^{1,6} Hyrum B. Johnson,² Stanley D. Smith,³
Stan D. Wullschleger,⁴ and Robert B. Jackson^{1,5}

Received 19 May 2003; revised 11 July 2003; accepted 5 August 2003; published 4 October 2003.

[1] Leaf stomatal characters influence the response of terrestrial evapotranspiration to climate change and are used as proxies for the reconstruction of past atmospheric [CO₂]. We examined the phenotypic response of stomatal index (SI), density (SD) and aperture (AP) to rising atmospheric CO₂ in 15 species after four years exposure to a field CO₂ gradient (200 to 550 μmol mol⁻¹ atmospheric [CO₂]) or at three Free Air CO₂ Enrichment (FACE) sites. Along the CO₂ gradient, SI and SD showed no evidence of a decline to increasing [CO₂], while AP decreased slightly. There was no significant change in SI, SD or AP with CO₂ across FACE experiments. Without evolutionary changes, SI and SD may not respond to atmospheric [CO₂] in the field and are unlikely to decrease in a future high CO₂ world. **INDEX TERMS:** 0315 Atmospheric Composition and Structure: Biosphere/atmosphere interactions; 1615 Global Change: Biogeochemical processes (4805); 1851 Hydrology: Plant ecology; 3344 Meteorology and Atmospheric Dynamics: Paleoclimatology. **Citation:** Reid, C. D., H. Maherali, H. B. Johnson, S. D. Smith, S. D. Wullschleger, and R. B. Jackson, On the relationship between stomatal characters and atmospheric CO₂, *Geophys. Res. Lett.*, 30(19), 1983, doi:10.1029/2003GL017775, 2003.

1. Introduction

[2] Various proxies are used to infer past atmospheric CO₂ concentrations and their consequences for the earth's climate [Crowley and Berner, 2001]. Stomatal density (SD; # stomata per unit area) and index (SI; # stomata divided by the sum of stomatal and epidermal cells #) of plants are used because paleontological data [Woodward, 1987; Retallack, 2001] and growth chamber experiments [Royer, 2001] suggest they decline with increasing CO₂ up to 600 μmol CO₂ mol⁻¹ [Beerling and Royer, 2002]. Stomatal density and the size of the stomatal aperture (AP; length between the junctions of the guard cells at each end of the stoma) partially determine leaf conductance to H₂O and CO₂, a key physiological variable in coupled atmosphere-biosphere models [Sellers et al., 1996; Kürschner et al., 1997; Collatz et al., 2000]. Studies in natural CO₂ vents, which provide a

long-term experimental system for adaptation to elevated CO₂, suggest that many species do not alter their SD or SI in response to elevated atmospheric CO₂ [Jones et al., 1995; Bettarini et al., 1998], although some increase AP [Tognetti et al., 2000]. However, there are few direct experimental tests of the CO₂ - SD/SI/AP relationship in the field. Furthermore, to our knowledge, no field experiment has tested the effect of low atmospheric CO₂ concentration on stomatal structure and function, a critical window for paleo-reconstruction. Here, we present data for 15 annuals and perennial herb and woody species suggesting that, in the absence of evolutionary changes, SD and SI do not decrease with increasing CO₂ under field conditions but AP may.

2. Methods

[3] We examined the response of SD, SI, and AP to CO₂ in the field using a unique four-year experiment that maintained a continuous CO₂ gradient from paleo to future atmospheric CO₂ [200 to 550 μmol mol⁻¹; Gill et al., 2002] and three long-term Free Air CO₂ Enrichment (FACE) experiments. We examined woody, herbaceous, and annual species because previous analyses of paleo-correlations between CO₂ concentrations and stomatal characters have relied primarily on woody perennial species but have also used herbs [Wooller and Agnew, 2002]. In addition, the annual species *Arabidopsis thaliana* has been instrumental in attempts to elucidate the role of CO₂ signaling on stomatal development [Gray et al., 2001; Lake et al., 2001].

[4] The continuous CO₂ gradient and other environmental variables in the tunnel system have been described previously [Johnson et al., 2000]. Briefly, it consisted of two parallel elongated chambers (1 m tall × 1 m wide × 60 m long) in a Texas grassland, with atmospheric CO₂ maintained from ambient to pre-industrial concentrations (365 to 200 μmol mol⁻¹) along one chamber and from elevated to ambient CO₂ along the other (550 to 350 μmol mol⁻¹). The different CO₂ concentrations along the gradient were maintained by varying the rate and direction of air flow through the chamber. Chamber temperature was controlled to track outside ambient temperatures. Although tunnel daytime temperatures were lower than ambient and differed between the pre-industrial and elevated chambers in the first year of operation [Johnson et al., 2000], temperature control was enhanced in subsequent years so that both chambers were at ambient temperature [H. W. Polley, USDA/ARS, Temple, TX, unpublished data]. In the 4th year of CO₂ exposure, fully expanded sun leaves from each species were sampled in the Spring (*Bromus japonicus*, *Solanum dimidiatum*, *Sorghum halepense*) or Fall (*Convolvulus equitans*, *Paspalum pubiflorum*, *Solidago canadensis*). The dominant *Bothriochloa ischaemum* was sampled during both periods and showed similar results. Only data for

¹Department of Biology, Duke University, Durham, NC, USA.

²USDA-ARS, Temple, TX, USA.

³Department of Biological Sciences, University of Nevada, Las Vegas, NV, USA.

⁴Oak Ridge National Laboratory, Oak Ridge, TN, USA.

⁵Nicholas School of the Environment and Earth Sciences, Duke University, Durham, NC, USA.

⁶Currently at Department of Botany, University of Guelph, Guelph, ON, Canada N1G 2W1.

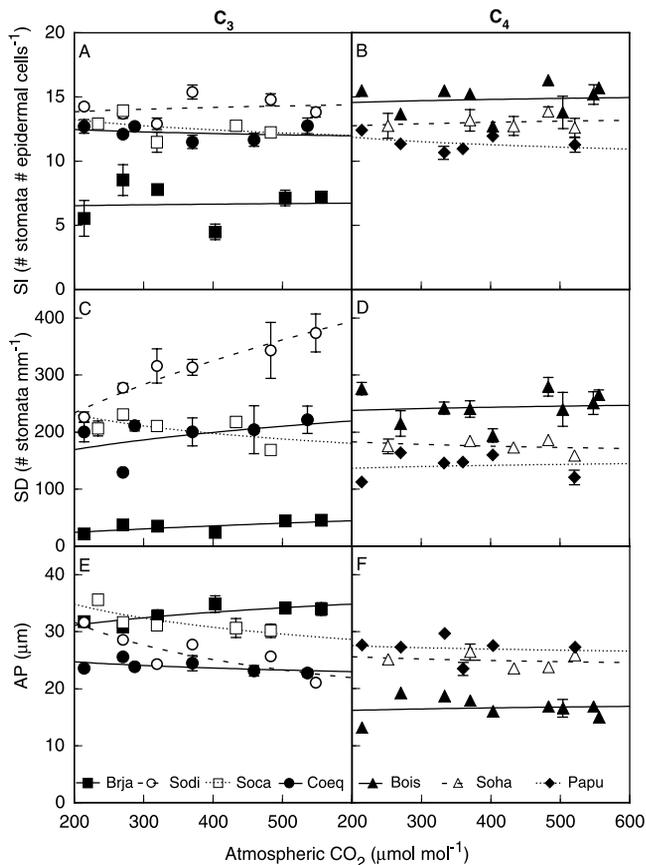


Figure 1. Stomatal characters of dominant species grown at subambient to elevated CO₂ in tunnel chambers for four years. Abaxial stomatal index (SI, the ratio of # stomata to the sum of stomata and epidermal cells #; (A–B), stomatal density (SD, # stomata per unit leaf area; (C–D), and the length of the stomatal aperture (AP, length between junction of the guard cells; E–F) are shown for C₃ grass and forbs (A, C, E): Brja, *Bromus japonicus*; Coeq, *Convolvulus equitans*; Soca, *Solidago canadensis*; Sodi, *Solanum dimidiatum*; and C₄ grasses (B, D, F): Bois, *Bothriochloa ischaemum*; Papu, *Paspalum pubiflorum*; Soha, *Sorghum halepense*. Data were fitted to a power model in SAS 6.12 (SAS Inst. Cary, NC). (n = 6, mean ± standard error).

species growing along the entire gradient from pre-industrial to elevated CO₂ are presented here.

[5] Free-Air CO₂ Enrichment (FACE) technology is used in a *Pinus taeda* plantation at the Duke Forest, Durham, NC

[DeLucia *et al.*, 1999], a desert scrub community at the Nevada Test Site, Las Vegas, NV [Smith *et al.*, 2000], and a *Liquidambar styraciflua* plantation at the Oak Ridge National Environmental Research Park, Oak Ridge, TN [Norby *et al.*, 2001]. All three FACE systems had been operating for at least four growing seasons when the leaves were sampled. CO₂ fumigation began in August 1996 at the Duke Forest, in April 1997 at the Nevada Test Site, and in April 1998 at the Oak Ridge forest. At the Duke Forest, fully expanded leaves were sampled in October 2000 (*L. styraciflua* and *Lonicera japonica*) and May 2001 (*Parthenocissus quinquefolia*, *Polygonatum biflorum*, *P. taeda*). *P. taeda* needles were sampled from the top of the forest canopy using mechanical lifts; the other species were sampled from the forest understory. At the Nevada desert site, plants were sampled in early May 2001. At Oak Ridge, upper canopy leaves of *L. styraciflua* were sampled in May 2001.

[6] At each field site, casts of fully expanded mature leaves were made by pressing leaf sections onto a microscope slide covered with polyvinylsiloxane dental impression material ('Extrude' Medium, Kerr Manufacturing Co, Orange, CA, USA; Williams and Greene [1988]). For short leaves (<2 cm length), the whole leaf was used. For longer leaves, a 1-cm length starting 2 cm up from the petiole base was used to minimize variability due to position [Poole *et al.*, 1996]. Each impression was analyzed at 100× or 400× on a light microscope interfaced with a solid state TV camera (Model CCD-72-SX; DAGE-MTI Inc., Michigan City, IN, USA) using an image analysis program (NIH Image 1.58; U.S. National Institutes of Health; <http://rsb.info.nih.gov/nih-image/>). Stomatal and epidermal cell counts were done on 3 to 6 fields-of-view per slide (depending on the variation in the counts) and were averaged for each slide. For the Oak Ridge site, the upper canopy leaves of 3 to 4 trees in each of three replicate ambient and elevated CO₂ rings were harvested. Leaves were placed into plastic envelopes and transported immediately to the laboratory, where leaf surface impressions were taken using clear fingernail polish. The number of stomata within each of three randomly selected fields (0.05 mm²) per leaf was counted using a light microscope, and stomatal density was averaged per ring.

3. Results and Discussion

[7] Along the CO₂ gradient, SI was not correlated with CO₂ either for any single species ($P > 0.18$ in all species, Figures 1a and 1b) or for the pooled data ($P = 0.87$, Figure 2a). Although SI is preferred to SD, because it is

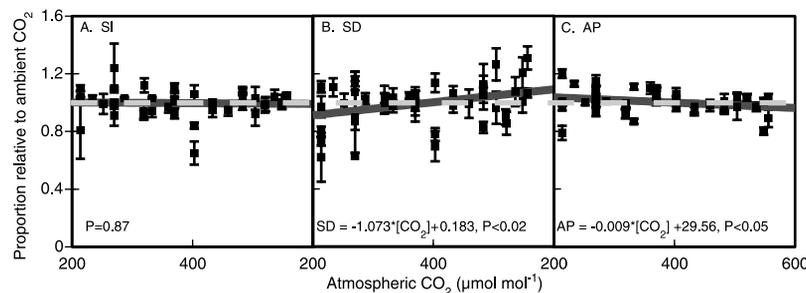


Figure 2. Relative change in stomatal characters of dominant species as a function of growth [CO₂] after four years in tunnel chambers. For all species pooled, each SI, SD, or AP at growth CO₂ is expressed as a proportion of SI, SD or AP at ambient CO₂. (n = 6 for mean ± standard error; dashed line = 1).

Table 1. Abaxial Stomatal Index (SI), Density (SD) and Length of the Stomatal Aperture (AP) of Woody and Herbaceous Species Grown at Current and Elevated Atmospheric CO₂ (FACE) for at Least Four Years

| Species | SI | | SD ^a | | AP | | Location |
|---|-------------------|----------|-----------------------------------|----------|-------------------|----------|--------------------------------------|
| | Ambient Avg. (##) | % Change | Ambient Avg. (#mm ⁻²) | % Change | Ambient Avg. (μm) | % Change | |
| <i>Eriogonum trichopes</i> (forb) ^b | 14.7 (0.3) | -0.4 | 312 (31) | +2.8 | 18.2 (1.5) | -1.1 | Nevada Test Site, NV ^d |
| <i>Larrea tridentata</i> (shrub) ^b | N/A | N/A | 121 (10) | +6.4 | 19.6 (0.4) | +11.2* | Nevada Test Site, NV |
| <i>Lepidium lasiocarpum</i> (forb) ^b | 13.5 (0.6) | +4.6 | 246 (16) | +15.6 | 14.8 (0.5) | -1.3 | Nevada Test Site, NV |
| <i>Liquidambar styraciflua</i> (tree) | N/A | N/A | 461 (22) ^c | -3.0 | N/A | N/A | Overstory, Oak Ridge TN ^e |
| <i>Liquidambar styraciflua</i> (tree) | 11.4 (0.4) | -8.1 | 264 (16) | -24.5** | 16.1 (0.4) | +3.0 | Understory, Durham NC ^f |
| <i>Lonicera japonica</i> (vine) | 11.9 (0.3) | -0.6 | 237 (10) | +6.8 | 17.5 (0.3) | +1.5 | Understory, Durham NC |
| <i>Parthenocissus quinquefolia</i> (vine) | 8.7 (0.2) | -0.15 | 121 (5) | +16.6 | 21.0 (0.5) | +2.2 | Understory, Durham NC |
| <i>Pinus taeda</i> (tree) | N/A | N/A | 112 (5) | +1.1 | 33.4 (1.2) | -1.8 | Overstory, Durham NC |
| <i>Polygonatum biflorum</i> (forb) | 16.4 (1.2) | +10.8 | 49 (8) | +1.5 | 27.5 (1.3) | +5.2 | Understory, Durham NC |

The % Change from ambient CO₂ (ca. 360 μmol mol⁻¹) in SI, SD or AP is shown for plants grown at elevated CO₂ (ca. 550–560 μmol mol⁻¹). In each FACE site, leaves of six individuals per species were used for each replicate ambient or elevated CO₂ ring (n = 18, mean ± standard error). CO₂ effect is significant at P < 0.05* and at P < 0.005** using ANOVA.

^aRatio of abaxial to adaxial SD was near 1 for amphistomatous species and was not affected by CO₂.

^bAmphistomatous leaf.

^cn = 4.

^dSmith et al. [2000].

^eNorby et al. [2001].

^fDeLucia et al. [1999].

unaffected by changes in epidermal cell expansion with CO₂ or other environmental factors [Retallack, 2001; Royer, 2001], SD is easier to measure and is often used when epidermal cells cannot be counted accurately. SD showed little evidence of the predicted decline with CO₂. Analyzed for species individually, SD increased significantly with CO₂ (Figure 1c) for *Br. japonicus* (r² = 0.243, P < 0.005) and *S. dimidiatum* (r² = 0.252, P < 0.005), opposite the direction predicted, and decreased only for one species, *S. canadensis* (r² = 0.180, P < 0.05). With all species pooled, SD showed a weak positive correlation with CO₂ (P = 0.02, Figure 2B), again opposite the direction predicted. AP showed a weak negative correlation with CO₂ (P < 0.05, Figure 2c) suggesting some compensation between stomatal number and size. However, individually, only *S. canadensis* and *S. dimidiatum* showed significant decreases in AP with increasing CO₂ (r² = 0.216, P < 0.004; and r² = 0.542, P < 0.0001, respectively, Figure 1e), while AP for *Br. japonicus* increased significantly (r² = 0.149, P < 0.01, Figure 1e).

[8] Results from the FACE experiments provide no evidence for a decline in SI or AP generally (P = 0.62 and P = 0.33, respectively) or for any species individually (Table 1). Similarly, the pooled SD data provided no evidence for a decline in SD at high CO₂ (P = 0.83, Table 1), also in contrast to current predictions. In fact, in seven of eight species, mean SD tended to be higher at elevated CO₂ (though not significantly), and only *L. styraciflua* when grown in the understory showed decreased SD (Table 1). These data suggest no significant morphological stomatal adjustments from current ambient to elevated CO₂. They also suggest that evolutionary timescales may be required for such responses to projected future CO₂ concentrations, as occurred for a few species in CO₂ vent studies [Jones et al., 1995; Tognetti et al., 2000]. Many trees will experience almost a doubling of atmospheric CO₂ in their current lifetime without the opportunity for such evolutionary changes.

[9] Our long-term experiments suggest that no general association between SD, SI or AP and future atmospheric

CO₂ is evident in the field. Available data on SD and SI from short-term CO₂ enrichment studies in open-top field chambers [Royer, 2001] and long-term records from near CO₂ springs [Bettarini et al., 1998] are consistent with this conclusion. Individually, species responded from pre-industrial to future CO₂ concentrations by a combination of change in stomatal size and number rather than solely by SD, and many species in all four experiments decrease their stomatal conductance in response to increased CO₂ despite the lack of decrease in SD or SI [e.g., Maherali et al., 2002; Nowak et al., 2001].

[10] Although our experiments do not incorporate long-term evolutionary effects, they reinforce some caveats about using stomatal characters in paleo-reconstruction of atmospheric CO₂, especially when historical factors such as precipitation, temperature, canopy position and leaf age are poorly characterized. Numerous studies have reported the effects of environmental variables other than CO₂ on stomatal development and morphology. For example, light availability and quality [e.g., Tichá, 1982; Schoch et al., 1984; Liu-Gitz et al., 2000] affect stomatal development and, hence, a plant can have leaves with different SD depending on leaf position in the canopy [Ceulemans et al., 1995]. Furthermore, signaling of light and CO₂ for stomatal development has been explored and appears similar [Lake et al., 2001]. Likewise, exposure to drought during leaf development reduces SD [Ciha and Brun, 1975; Awada et al., 2002], and, in altitudinal gradients where atmospheric CO₂ decreases, SD is determined by precipitation rather than CO₂ concentration [Körner et al., 1986]. Therefore, environmental factors other than CO₂ concentration may be stronger determinants of stomatal characters in natural settings.

[11] Predictions of reduced SD, SI, or AP with rising atmospheric CO₂ are implicit in coupled biosphere-atmosphere models as a mechanism for declining stomatal conductance [Sellers et al., 1996; Kürschner et al., 1997; Aasamaa et al., 2001]. These relationships are also being incorporated into projected forest responses to pollutants

such as ozone [Evans *et al.*, 1996]. Our results indicate that SI, SD, and AP are unlikely to decline in response to future high CO₂, and may not have responded significantly to low CO₂ in the past. Stronger field-based evidence supporting the relationships should emerge before stomatal characters are incorporated into biosphere-atmosphere models.

[12] **Acknowledgments.** We thank C. W. Cook for assistance. The research was supported by the National Institute for Global Environmental Change program, DOE, and the National Science Foundation.

References

- Aasamaa, K., A. Söber, and M. Rahi, Leaf anatomical characteristics associated with shoot hydraulic conductance, stomatal conductance and stomatal sensitivity to changes of leaf water status in temperate deciduous trees, *Aust. J. Plant Physiol.*, **28**, 765–774, 2001.
- Awada, T., L. E. Moser, W. H. Schacht, and P. E. Reece, Stomatal variability of native warm-season grasses from the Nebraska Sandhills, *Can. J. Plant Science*, **82**, 349–355, 2002.
- Beerling, D. J., and D. L. Royer, Reading a CO₂ signal from fossil stomata, *New Phytol.*, **153**, 387–397, 2002.
- Bettarini, I., F. P. Vaccari, and F. Miglietta, Elevated CO₂ concentrations and stomatal density: Observations from 17 plant species growing in a CO₂ spring in central Italy, *Glob. Change Biol.*, **4**, 17–22, 1998.
- Ceulemans, R., L. Van Praet, and X. N. Jiang, Effect of CO₂ enrichment, leaf position and clone on stomatal index and epidermal cell density in poplar (*Populus*), *New Phytol.*, **131**, 99–107, 1995.
- Ciha, A. J., and W. A. Brun, Stomatal size and frequency in soybeans, *Crop Sci.*, **15**, 309–313, 1975.
- Collatz, G. J., L. Bounoua, S. O. Los, D. A. Randall, I. Y. Fung, and P. J. Sellers, A mechanism for the influence of vegetation on the response of the diurnal temperature range to changing climate, *Geophys. Res. Lett.*, **27**, 3381–3384, 2000.
- Crowley, T. J., and R. A. Berner, CO₂ and climate change, *Science*, **292**, 870–872, 2001.
- DeLucia, E. H., J. G. Hamilton, S. L. Naidu, R. B. Thomas, J. A. Andrews, A. Finzi, M. Lavine, R. Matamala, J. E. Mohan, G. R. Hendrey, and W. H. Schlesinger, Net primary production of a forest ecosystem with experimental CO₂ enrichment, *Science*, **284**, 1177–1179, 1999.
- Evans, L. S., K. Albury, and N. Jennings, Relationships between anatomical characteristics and ozone sensitivity of leaves of several herbaceous dicotyledonous plant species at Great Smoky Mountains National Park, *Env. Exp. Bot.*, **36**, 413–420, 1996.
- Gill, R. A., H. W. Polley, H. B. Johnson, L. J. Anderson, H. Maherali, and R. B. Jackson, Nonlinear grassland responses to past and future atmospheric CO₂, *Nature*, **417**, 279–282, 2002.
- Gray, J. E., G. H. Holroyd, F. M. van der Lee, A. R. Bahrami, P. C. Sijmons, F. I. Woodward, W. Schuch, and A. M. Hetherington, The *HIC* signaling pathway links CO₂ perception to stomatal development, *Nature*, **408**, 713–716, 2001.
- Johnson, H. B., H. W. Polley, and R. P. Whitis, Elongated chambers for field studies across atmospheric CO₂ gradients, *Funct. Ecol.*, **14**, 388–396, 2000.
- Jones, M. B., J. C. Brown, A. Raschi, and F. Miglietta, The effects on *Arbutus unedo* L. of long-term exposure to elevated CO₂, *Glob. Change Biol.*, **1**, 295–302, 1995.
- Körner, Ch., P. Bannister, and A. F. Mark, Altitudinal variation in stomatal conductance, nitrogen content and leaf anatomy in different plant life forms in New Zealand, *Oecologia*, **69**, 577–588, 1986.
- Kürschner, W. M., F. Wagner, E. H. Visscher, and H. Visscher, Predicting the response of leaf stomatal frequency to a future CO₂-enriched atmosphere: Constraints from historical observations, *Geol. Rund.*, **86**, 512–517, 1997.
- Lake, J. A., W. P. Quick, D. J. Beerling, and F. I. Woodward, Signals from mature to new leaves, *Nature*, **411**, 154, 2001.
- Liu-Gitz, L., S. J. Britz, and W. P. Wergin, Blue light inhibits stomatal development in soybean isolines containing kaempferol-3-O-2^G-glycosyl-gentiobioside (K9), a unique flavonoid glycoside, *Plant Cell & Environ.*, **23**, 883–891, 2000.
- Maherali, H., C. D. Reid, H. W. Polley, H. B. Johnson, and R. B. Jackson, Stomatal acclimation over a subambient to elevated CO₂ gradient in a C₃/C₄ grassland, *Plant Cell & Environ.*, **25**, 557–566, 2002.
- Norby, R. J., D. E. Todd, J. Fults, and D. W. Johnson, Allometric determination of tree growth in a CO₂-enriched sweetgum stand, *New Phytol.*, **150**, 477–487, 2001.
- Nowak, R. S., L. A. DeFalco, C. S. Wilcox, D. N. Jordan, J. S. Coleman, J. R. Seemann, and S. D. Smith, Leaf conductance decreased under free-air CO₂ enrichment (FACE) for three perennials in the Nevada desert, *New Phytol.*, **150**, 449–458, 2001.
- Poole, I., J. D. B. Weyers, T. Lawson, and J. A. Raven, Variations in stomatal density and index: Implications for palaeoclimatic reconstructions, *Plant Cell & Environ.*, **19**, 705–712, 1996.
- Retallack, G. J., A 300-million-year record of atmospheric carbon dioxide from fossil plant cuticles, *Nature*, **411**, 287–290, 2001.
- Royer, D. L., Stomatal density and stomatal index as indicators of paleoatmospheric CO₂ concentration, *Rev. Paleobot. Palynol.*, **114**, 1–28, 2001.
- Schoch, P. G., R. Jacques, A. Lecharny, and M. Sibi, Dependence of the stomatal index on environmental factors during stomatal differentiation in leaves of *Vigna sinenses* L. II. Effect of different light quality, *J. Exp. Bot.*, **35**, 1405–1409, 1984.
- Sellers, P. J., L. Bounoua, G. J. Collatz, D. A. Randall, D. A. Dazlich, S. O. Los, J. A. Berry, I. Fung, C. J. Tucker, C. B. Field, and T. G. Jensen, Comparison of radiative and physiological effects of doubled atmospheric CO₂ on climate, *Science*, **271**, 1402–1406, 1996.
- Smith, S. D., T. E. Huxman, S. F. Zitzer, T. N. Charlet, D. C. Housman, J. S. Coleman, L. K. Fenstermaker, J. R. Seemann, and R. S. Nowak, Elevated CO₂ increases productivity and invasive species success in an arid ecosystem, *Nature*, **408**, 79–82, 2000.
- Tichá, I., Photosynthetic characteristics during ontogenesis of leaves. 7. Stomata density and sizes, *Photosynthetica*, **16**, 375–471, 1982.
- Tognetti, R. R., A. Minnocci, J. Peñuelas, A. Raschi, and M. B. Jones, Comparative field water relations of three Mediterranean shrub species co-occurring at a natural CO₂ vent, *J. Exp. Bot.*, **51**, 1135–1146, 2000.
- Williams, M. H., and P. B. Greene, Sequential scanning electron microscopy of a growing plant meristem, *Protoplasma*, **147**, 77–79, 1988.
- Woodward, F. I., Stomatal numbers are sensitive to increases in CO₂ from preindustrial levels, *Nature*, **327**, 617–618, 1987.
- Wooller, M. J., and A. D. Q. Agnew, Changes in graminoid stomatal morphology over the last glacial-interglacial transition: Evidence from Mount Kenya, East Africa, *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, **177**, 123–136, 2002.

R. B. Jackson and C. D. Reid, Department of Biology, Duke University, Durham, NC 27708, USA. (chantal@duke.edu)

H. B. Johnson, USDA-ARS, Temple, TX 76502, USA.

H. Maherali, Department of Botany, University of Guelph, Guelph, ON, Canada N1G 2W1.

S. D. Smith, Department of Biological Sciences, University of Nevada, Las Vegas, NV 89154, USA.

S. D. Wullschleger, Oak Ridge National Laboratory, Oak Ridge, TN 37831, USA.